

Curating and evaluating RNA structure assignments

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Outline:

- Part I: Semi-Automated RNA Sequence Editor (SARSE).
 - Curating RNA structural alignments.
 - Rnadbtools and SARSE.
 - Integrating Pfold and Pcluster.
 - The temperature of Rfam.
- Part II: R_K : The K category correlation coefficient.
 - Comparing two K category assignments.
 - Pearson's correlation coefficient and least square fitting.
 - Extending Pearson's correlation coefficient to two K dimensional tables the R_K coefficient.
 - Discretization, an extension of Matthews correlation coefficient.
 - Applications of R_K .

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Semi-Automated RNA Sequence Editor (SARSE)

Motivation:

- Good curated alignments to compare with predictions.
- Some years ago (still valid): Most ncRNA structural alignments have bad quality and obvious contain inconsistencies.
- Some databases even lack structural assignments corresponding to the multiple RNA alignment.
- Some years ago SRP RNA one of the best alignments, but had many inconsistencies. Old clean: from 20 to 3 pr sequence.
- Doing this kind of work is extremely useful to the community, but also extremely low prestige (and no funding :-(..).)
- Exist no good editor which include basic editing functions combined with structural consistency checks.

RNAdbtools: <http://rnadbtool.kvl.dk>

- Toolbox to conduct basic consistency checks.
- Highlights any tyupe of non-standard RNA pairs (and check, whether bases assigned to the same pair).
- Extends RNA stems where possible.
- Automated search and align0 realignment of global regions of blast hits. [Now outdated].
- Introduction of the column format: <http://colformat.kvl.dk>.
- Colformat motivation. Easy to work with while very flexible. [hence much man power in time are saved!]

We would like to have SARSE

- Make RNAdbtools interactive.
- Features: Split view; drop drag; highlight complement bases; unlimited undo/redo sessions; overview (click and jump to region); history window.
- Integrate your own commandline software into SARSE [by dumping data in the colformat].
- Extends RNA stems where possible.
- Automated search and align0 realignment of global regions of blast hits. [Now outdated, but similar stuff could be included in SARSE].
- SARSE: Jave based interface. Basic editor funtions directly incoorporated.
- Other software can be executed: RNAdbtools, pfold and pcluster [NEW !].
- Extensive documentation: <http://sarse.kvl.dk>.

Semi-Automated RNA Sequence Editor

Clean up RNA multiple structural alignments. <http://sarse.kvl.dk> (See intro)

The screenshot displays the Sarse (Semi-Automated RNA Sequence Editor) interface. The main window shows a multiple sequence alignment of HIV-1 RNA sequences. The alignment is presented in two columns, with sequence identifiers on the left and right. The sequences are color-coded by reliability: green for double-stranded regions and blue for single-stranded regions. The alignment is titled "result-1127982993814.col".

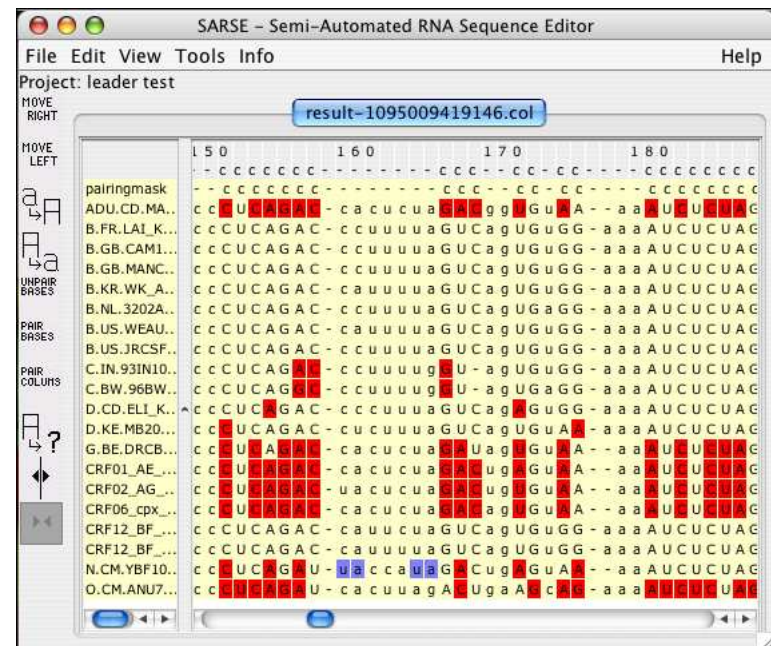
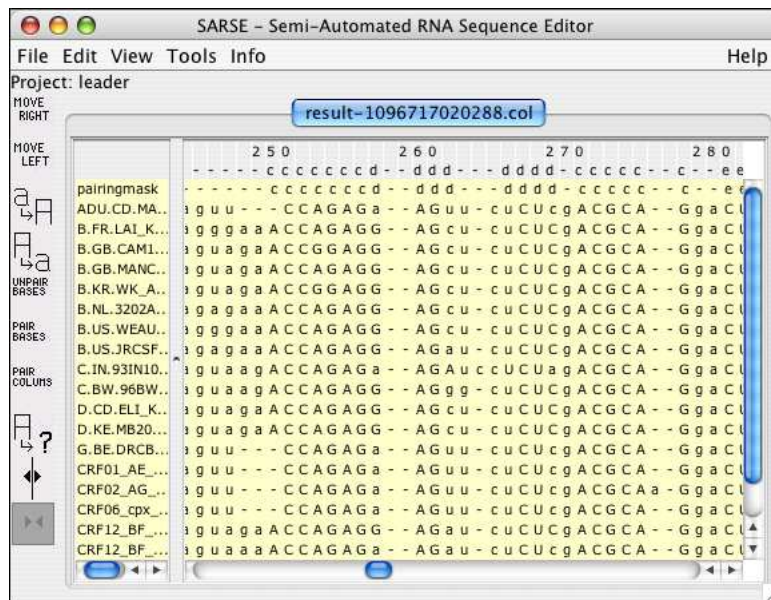
Below the alignment, there are two smaller windows: "Overview" showing a bar chart of sequence reliability and "History" showing a list of recent actions such as "Move left" and "Move right".

On the right side, a secondary structure prediction window titled "B_HXB2_K03455" shows a complex RNA fold with numbered regions (1-9). A legend below the structure indicates reliability levels: green for double-stranded (>90% and >70%) and blue for single-stranded (<70% and >70% and >90%).

At the bottom, a terminal window shows the command-line interface for the software, including the following commands and output:

```
[ebbe@momo-130207-dellp4 tutorial-data]$ ls
pcluster/ pfold/
[ebbe@momo-130207-dellp4 tutorial-data]$ cd pfold
[ebbe@momo-130207-dellp4 pfold]$ ls
00_README 01_HIV1_leader.fasta 02_HIV1_leader.aln 03_HIV1_leader2.txt 03_HIV1_leader.fasta 03_HIV1_leader.txt 04_HIV1_leader.col HIV-1/ test2/
[ebbe@momo-130207-dellp4 pfold]$ emacs 03
03_HIV1_leader2.txt 03_HIV1_leader.fasta 03_HIV1_leader.txt
[ebbe@momo-130207-dellp4 pfold]$ emacs 03_HIV1_leader2.txt
[ebbe@momo-130207-dellp4 pfold]$
```

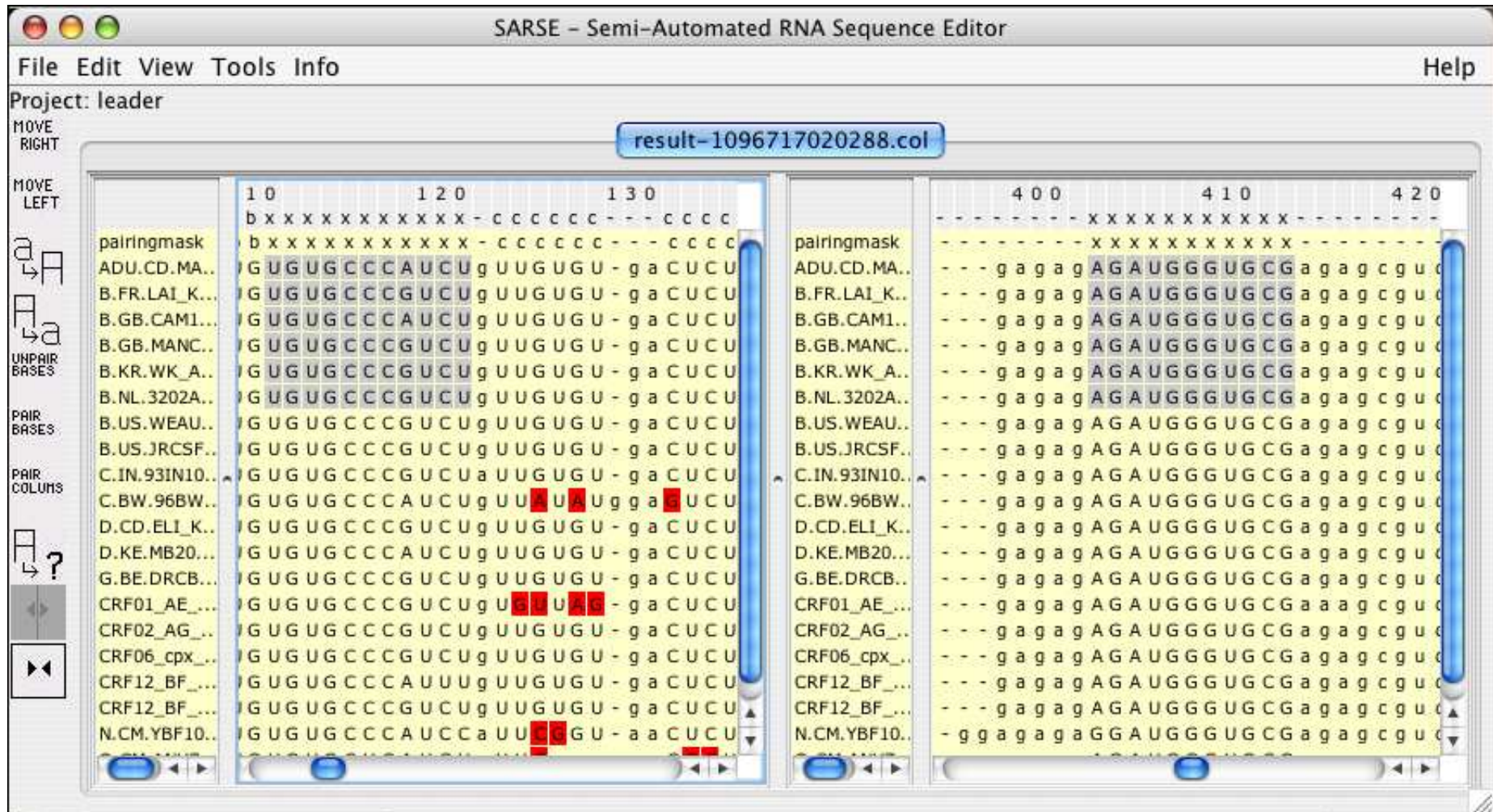
SARSE: Semi-Automated RNA Sequence Editor



- Invoke RNADBTOOLS to point out inconsistencies.
- Small overview box (not shown) gives a global view.
- Data from programs dumped in a projects directory.

SARSE: Semi-Automated RNA Sequence Editor

Split view



SARSE: Semi-Automated RNA Sequence Editor

Coloring according to Pfold reliability scores:

The screenshot displays the SARSE (Semi-Automated RNA Sequence Editor) application window. The title bar reads "SARSE - Semi-Automated RNA Sequence Editor". The menu bar includes "File", "Edit", "View", "Tools", "Info", and "Help".

At the top center, two buttons are visible: "ebbe test" and "test_ebbe.col".

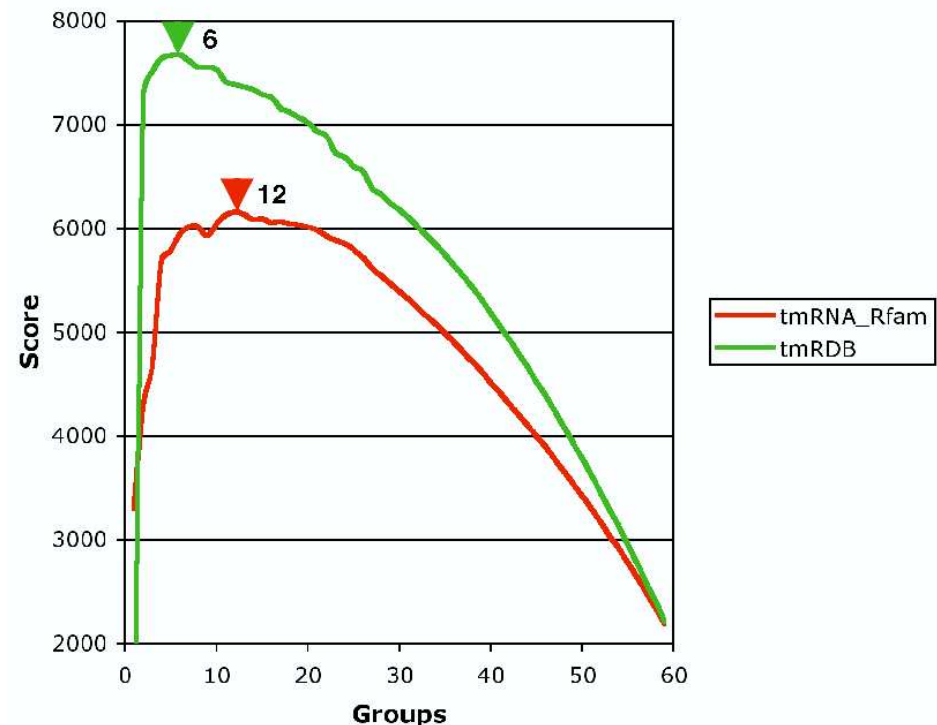
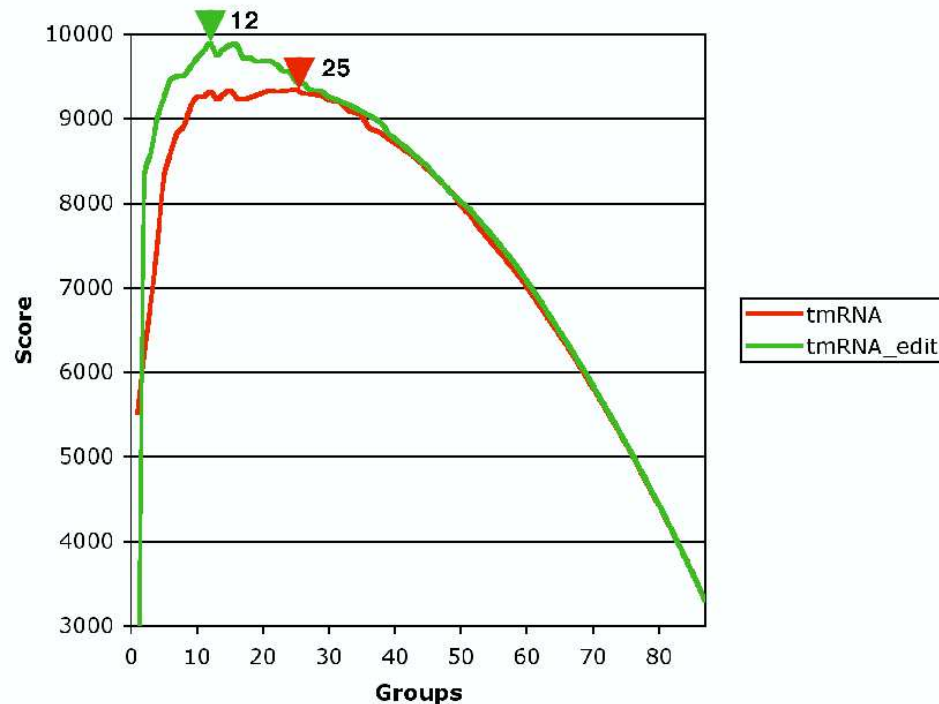
The main editing area is divided into two horizontal panels. The top panel shows a sequence alignment for positions 1 through 30. The bottom panel shows positions 30 through 60. Each panel has a list of sequence names on the left and a grid of colored cells representing the sequences. The colors range from green to red, indicating Pfold reliability scores. The top panel also shows a "pairingmask" row above the sequences, with '1' indicating paired positions and '0' indicating unpaired positions.

On the left side of the interface, there are several control buttons: "MOVE RIGHT", "MOVE LEFT", "PAIR BASES", "PAIR COLUMNS", "SINGLE SELECT", and "DOUBLE SELECT".

Semi-Automated RNA Sequence Editor: Pcluster

- pfold structure disrupted: poor alignment or variations in structure.
- Detect structural subgroups.
- Score: Reliability weighted sum of base pairs.
- Greedy heuristic method by joining subgroups with highest score.
- Extract: number of subgroups for max score.
- Heuristic to find "best" groupings by interpolation between max score point that score half of max point.

Semi-Automated RNA Sequence Editor: Pcluster



... and to something not so different:

You yeah I mean you: you train and test your prediction methods on these data

SARSE perspectives

- Adding even more programs. RNAalifold etc.
- Adding: auto fetching of sequences from databases.
- Web server set up.
- Web suites for specialist to curate their set of families.

Part II The K category correlation coefficient

Motivation:

- Predictions can yield more than dichotomies.
[Eg. protein secondary structure]
- RNA secondary structure predictions: bp, \neg bp, unassignable.
- Exists measure for comparing multiple categories
[eg: Escoufier (1973) and review by Baldi *et al.* (2000).]
- None of the measures have completely desired properties.
- Goal: an extension of Matthews correlation coefficient:

$$C = \frac{P_t N_t - P_f N_f}{\sqrt{(N_t + N_f)(N_t + P_f)(P_t + N_f)(P_t + P_f)}}$$

- Idea: Simple extension of Pearson's correlation coefficient.

Note on Matthews correlation coefficient for ncRNA evaluation

(... and to something *slightly* different!)

Basepair prediction N_t factor N larger than P_t, P_f, N_f . [$N(N-1)/2$ pairs of bases.]

$$\begin{aligned} C &= \frac{P_t N_t - P_f N_f}{N_t \sqrt{(1 + N_f/N_t)(1 + P_f/N_t)(P_t + N_f)(P_t + P_f)}} \\ &\approx \frac{P_t N_t - P_f N_f}{N_t \sqrt{(P_t + N_f)(P_t + P_f)}} \\ &= \frac{P_t}{\sqrt{(P_t + N_f)(P_t + P_f)}} \left[1 - \frac{P_f N_f}{P_t N_t} \right] \end{aligned}$$

where $N_f/N_t \rightarrow 0$ and $P_f/N_t \rightarrow 0$ for $N \rightarrow \infty$. For any reasonable prediction method ($P_t > 0$), with at least $P_t \sim P_f$ or $P_t \sim N_f$, we can write

$$C \approx \frac{P_t}{\sqrt{(P_t + N_f)(P_t + P_f)}} = \sqrt{\frac{P_t}{P_t + N_f} \frac{P_t}{P_t + P_f}},$$

Pearson's correlation coefficient and least square fitting

Pearson's correlation coefficient:

$$r = \frac{COV(X, Y)}{\sqrt{COV(X, X)COV(Y, Y)}}, \quad COV(X, Y) = \sum_{n=1}^N (X_n - \bar{X})(Y_n - \bar{Y})$$

For variables Y and X of length N least. Least square fitting in the coefficient b :

$$Y = a + bX$$

yield an expression for b . Conversely an similar expression can be obtained for fitting in the coefficient b' :

$$X = a' + b'Y$$

For a linear fit:

$$E = \sum_{n=1}^N (Y_n - (a + bX_n))^2$$

partial derivatives in a and b should be zero. It follows that

$$r^2 = bb', \quad b = \frac{COV(X, Y)}{COV(X, X)}$$

Extending Pearson's correlation coefficient to two K -dimensional tables, the R_K coefficient

Consider two $N \times K$ tables: $\underline{\underline{X}}$ and $\underline{\underline{Y}}$. Define

$$COV(\underline{\underline{X}}, \underline{\underline{Y}}) = \sum_{k=1}^K w_k COV(\underline{\underline{X}}_k, \underline{\underline{Y}}_k) = \frac{1}{K} \sum_{n=1}^N \sum_{k=1}^K (X_{nk} - \bar{X}_k)(Y_{nk} - \bar{Y}_k)$$

where $\bar{X}_k = \frac{1}{N} \sum_{n=1}^N X_{nk}$ and \bar{Y}_k are the respective means of column k . Use ("prior") $w_k = 1/K$.

$$R_K = \frac{COV(\underline{\underline{X}}, \underline{\underline{Y}})}{\sqrt{COV(\underline{\underline{X}}, \underline{\underline{X}})COV(\underline{\underline{Y}}, \underline{\underline{Y}})}}$$

Basic properties: $R_K \in [-1, 1]$; $R_1 = r$; $R_2 = r$, when $X_{n1} + X_{n2} = a$ and $Y_{n1} + Y_{n2} = b$. Hence R_2 reduces to Matthews correlation coefficient when X and Y components only take the values $\{0, 1\}$.

Relation to least square fitting

K related linear fits $\vec{Y} = \vec{a} + b\vec{X}$ over the N data points. $K = 1$: Pearson case.

Weighted difference in a cost function:

$$E = \sum_{n=1}^N \sum_{k=1}^K w_k (Y_{nk} - (a_k + bX_{nk}))^2$$

To obtain minimum. Require: $\partial E / \partial a_k = 0$ (for all $k = 1, \dots, K$) and $\partial E / \partial b = 0$.

After a little algebra:

$$\sum_{k=1}^K w_k \left(\sum_{n=1}^N X_{nk} Y_{nk} - N \bar{X}_k \bar{Y}_k \right) = b \left\{ \sum_{k=1}^K w_k \left(\sum_{n=1}^N X_{nk}^2 - N \bar{X}_k^2 \right) \right\}$$

yielding

$$b = \frac{COV(\underline{\underline{X}}, \underline{\underline{Y}})}{COV(\underline{\underline{X}}, \underline{\underline{X}})} \quad \text{and} \quad R_K^2 = bb'$$

The Discrete version of R_K

- The $K \times K$ confusion matrix $\underline{\underline{C}}$.
- Let C_{kl} be the number X_{nk} 's predicted to belong to class k , but belong to class l , $l \neq k$.
- For $K=2$: C_{11} : true positives; C_{22} : true negatives; C_{12} : false positives; C_{21} : false negatives.
- Well known observations:
 - $N = \sum_{kl} C_{kl}$.
 - $\bar{X}_k = \frac{1}{N} \sum_l C_{kl}$.
 - $\bar{Y}_k = \frac{1}{N} \sum_l C_{lk}$.
 - $C_{kk} = \sum_n X_{nk} Y_{nk}$.

The Discrete version of R_K

Plug in the known observations to R_K and obtain

$$R_K = \frac{\sum_{klm} C_{kk}C_{lm} - C_{kl}C_{mk}}{\sqrt{\sum_k \left(\sum_l C_{kl} \right) \left(\sum_{\substack{l' \\ k' \neq k}} C_{k'l'} \right)} \sqrt{\sum_k \left(\sum_l C_{lk} \right) \left(\sum_{\substack{l' \\ k' \neq k}} C_{l'k'} \right)}}$$

or equivalently

$$R_K = \frac{N \text{Tr}(\underline{\underline{C}}) - \sum_{kl} \underline{\underline{C}}_k \hat{\underline{\underline{C}}}_l}{\sqrt{N^2 - \sum_{kl} \underline{\underline{C}}_k (\hat{\underline{\underline{C}}}_l^\top)_l} \sqrt{N^2 - \sum_{kl} (\underline{\underline{C}}_k^\top)_k \hat{\underline{\underline{C}}}_l}}$$

- $\underline{\underline{C}}_k$ the k th row of $\underline{\underline{C}}$.
- $\hat{\underline{\underline{C}}}_l$ the l th column of $\underline{\underline{C}}$.
- $\underline{\underline{C}}^\top$ is $\underline{\underline{C}}$ transposed.

Applications of R_K

Comparison to other measures of evaluating protein secondary structure predictions [From EVA (Rost and Co-workers)]

- Numerous approaches for protein secondary structure prediction.
- Predicting the three classes, α -helix, β -sheet and coil.
- Comparing to Q_3 ranking, the fraction of correctly predictions over all three classes.
- Comparing to SOV (Segment Overlap), measure that take continuous stretches of helices and sheet into consideration in the evaluation.

Applications of R_K

- Eva (as of August 2003) have several classes of different set sizes.
- Each set covers different number of predictions methods.

R	1	0	0	1	0	0
L	1	0	0	1	0	0
R	1	0	0	1	0	0
V	1	0	0	1	0	0
H	1	0	0	1	0	0
Q	1	0	0	1	0	0
I	1	0	0	1	0	0
A	1	0	0	1	0	0
E	1	0	0	1	0	0
E	1	0	0	1	0	0
H	0	0	1	1	0	0
G	0	0	1	0	0	1
L	0	1	0	0	0	1
R	0	1	0	0	0	1
H	0	1	0	0	0	1
D	0	1	0	0	0	1
S	0	1	0	0	0	1
S	0	1	0	0	0	1
G	0	0	1	0	0	1
E	0	0	1	0	0	1
F	0	0	1	0	0	1
K	0	0	1	0	0	1
G	0	0	1	0	0	1
α	α	β	C	α	β	C

Set	method	R_3	rank	sov	Q_3
1	profsec	0.621	1	74.8	75.09
	psipred	0.619	1	73.6	75.11
	apssp2	0.613	1	71.4	74.74
	samt99_sec	0.613	1	71.1	74.68
	sspro2	0.598	1	69.1	73.81
	phdpsi	0.581	1	69.7	72.61
	jpred	0.570	1	70.3	71.81
	prospect	0.567	1	69.8	71.77
	prof_king	0.555	1	69.8	70.83
phd	0.526	2	64.5	69.01	
2	profsec	0.600	1	71.5	74.00
	samt99_sec	0.586	1	67.1	73.28
	psipred	0.579	1	69.8	72.86
	sspro2	0.573	1	67.4	72.50
	phdpsi	0.560	1	66.9	71.62
	prof_king	0.544	1	66.5	70.33
	jpred	0.536	1	66.7	69.92
phd	0.505	2	62.6	68.03	
3	profsec	0.600	1	71.5	74.00
	psipred	0.579	1	69.8	72.86
	samt99_sec	0.586	1	67.1	73.28
	phdpsi	0.560	1	66.9	71.62
	prof_king	0.544	1	66.5	70.33
	jpred	0.536	1	66.7	69.92
phd	0.505	2	62.6	68.03	
4	profsec	0.608	1	71.7	74.61
	psipred	0.591	1	71.1	73.69
	samt99_sec	0.591	1	68.9	73.69
	phdpsi	0.568	1	67.7	72.22
	jpred	0.545	1	67.5	70.63
phd	0.512	2	64.5	68.54	
5	psipred	0.608	1	71.0	74.82
	profsec	0.606	1	70.2	74.52
	samt99_sec	0.600	1	69.5	74.32
	phdpsi	0.566	2	66.6	72.13
	phd	0.533	3	64.8	69.95
6	psipred	0.617	1	72.1	75.44
	profsec	0.610	1	71.2	74.82
	phdpsi	0.565	2	67.5	72.10
	phd	0.540	3	65.9	70.43

Applications of R_K

RNA example of R_3 . First some background:

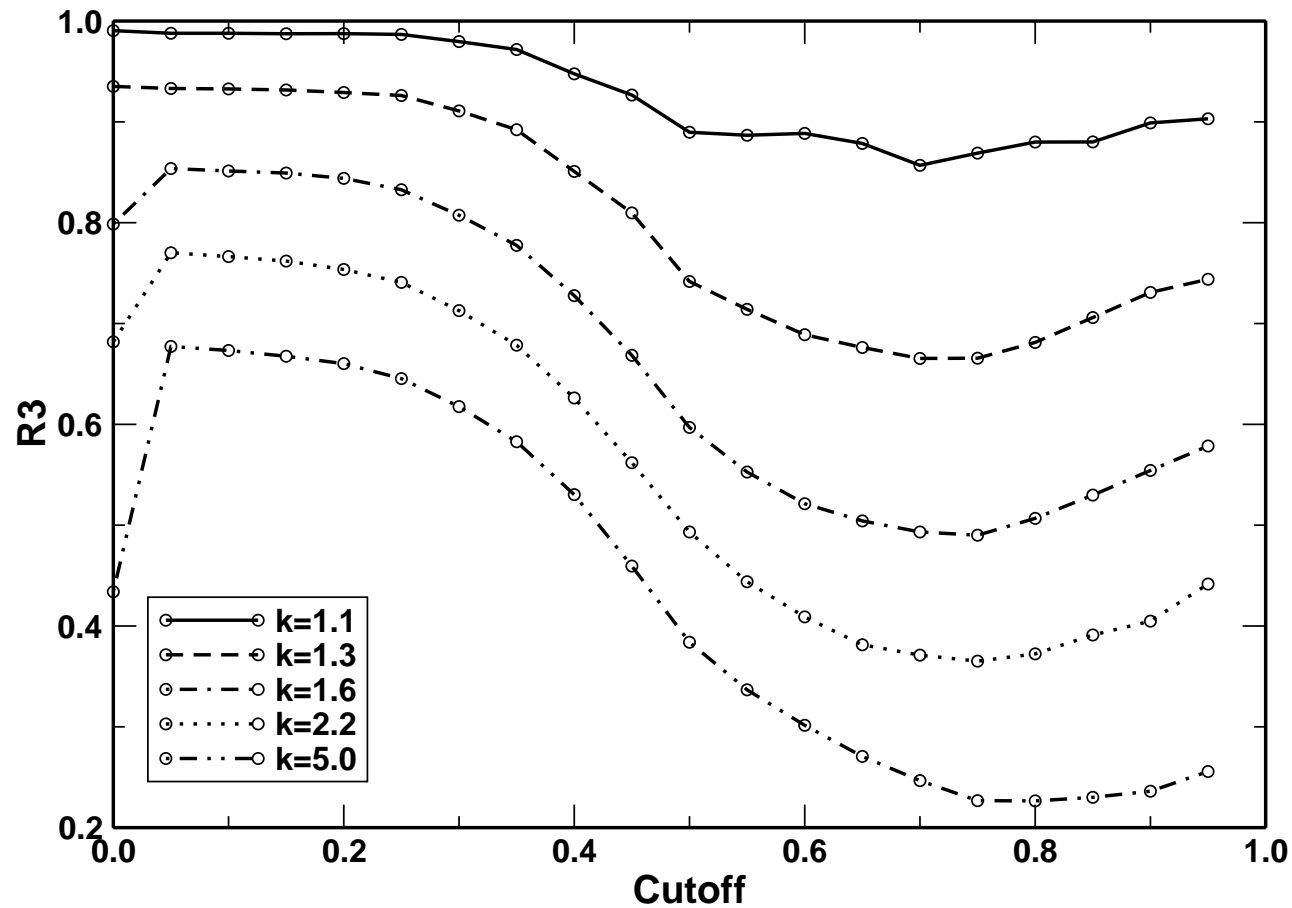
- Study applying pfold to structure prediction of the HIV leader (713 nt.) using 20 aligned sequences (Knudsen *et al.*, 2004).
- Study: Compare pfold predictions to predictions with perturbed rate values.
- Aim of study: can the evolutionary rates estimated from rRNA and tRNA be applied on the much faster evolving HIV-1 sequence.
[NOTE: this assumption is implicit for all prediction methods estimating parameters from *e.g.*, rRNA and tRNA, such as for QRNA.]
- The answer was yes; the HIV-1 prediction is fine, just within the limits stabil predictions obtained when perturbing the evolutionary rates.
- Rates were essentially perturbed by having 50% chance of making the rate k times larger or k times smaller [and adding some normalization constraints].

Applications of R_K

- Introducing a third category, the unknown or unassignable categories.
- pfold predictions uncertain for low reliability score.
- Statement: If pfold score low for positions that are manually hard to assign basepairs, the overall prediction should be higher than if an assignment was enforced on these positions.
- Use different reliability score cut-offs for sending a basepair in the third category "unknown". The reference structure is still the original pfold prediction.

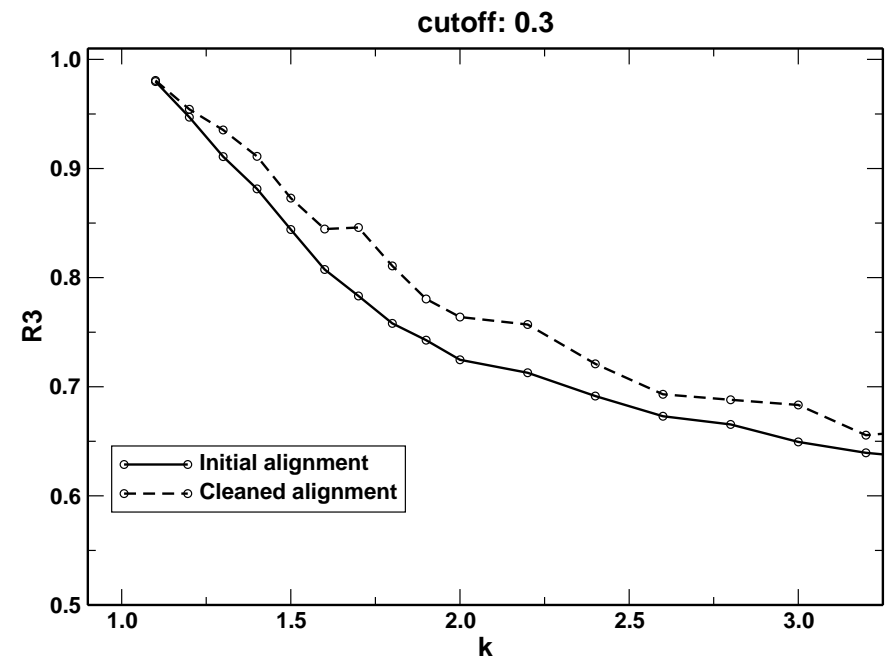
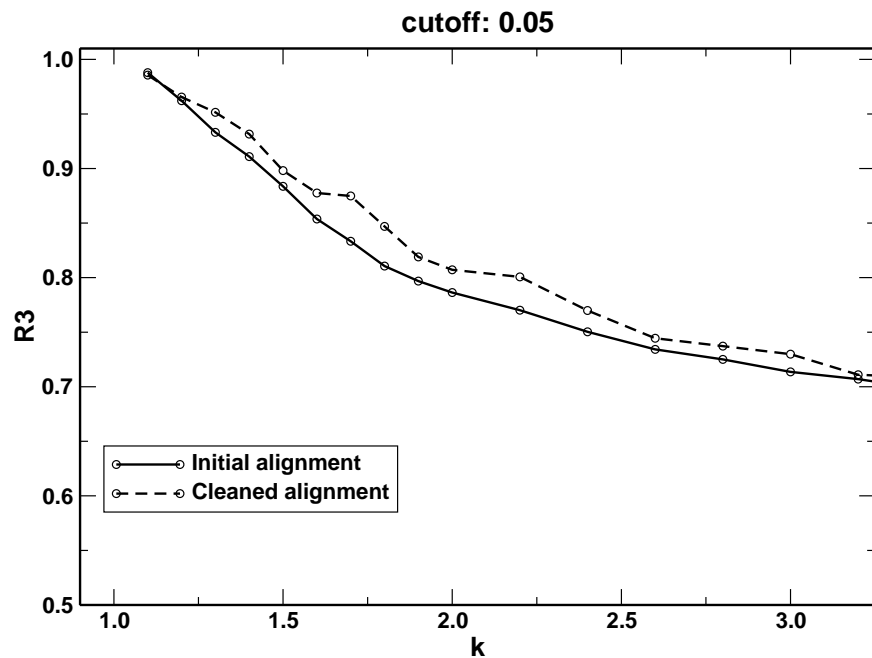
Applications of R_K

Rate variation performance for various cut-offs.



Applications of R_K

RNA structure predictions for different reliability (pfold) cut-offs for varying rate perturbations k . A comparison between initial and cleaned alignment.



Perspectives for R_K

Measures for comparing a predicted structure assignment to a curated structure assignment.

- Applying R_K to cases of $K > 3$.
- A measure as SOV would be needed to take prediction of entire helices into consideration.
- Further extension: Comparing $L \times N \times K$ tables and compute one correlation coefficient.