# Annotation and differential expression analysis of non-coding RNAs in 16 freely accessible bat genomes 

Marie Lataretu, Friedrich-Schiller-Universität Jena
$14^{\text {th }}$ February, 2019

## Bats are cool!

Features

- The only flying mammals



## Bats are cool!

Features

- The only flying mammals
- Laryngeal echolocation



## Bats are cool!

Features

- The only flying mammals
- Laryngeal echolocation
- Vocal learning



## Bats are cool!

Features

- The only flying mammals
- Laryngeal echolocation
- Vocal learning
- Account for ~20 \% of all mammal species



## Bats are cool!

## Features

- The only flying mammals
- Laryngeal echolocation
- Vocal learning
- Account for ~ 20 \% of all mammal species
- Immunity against various pathogenic viruses



## Bats are cool!

## Features

- The only flying mammals
- Laryngeal echolocation
- Vocal learning
- Account for ~ 20 \% of all mammal species
- Immunity against various pathogenic viruses
- Show homosexual behavior ${ }^{1}$


[^0]
## Freely available genomes and annotations (today)

- Genomes: 32 of more than 1,300 species



## Freely available genomes and annotations (today)

- Genomes: 32 of more than 1,300 species
- Annotations: 11 of 32 species



## Freely available genomes (before 15 January 2019)



## Freely available annotations (before 15 January 2019)



## Freely available annotations (before 15 January 2019)



Maximal number of annotated RNAs for each RNA class.

## Hackaton



## Hackaton



1. Annotation of non-coding RNAs in 16 bats

## Hackaton



1. Annotation of non-coding RNAs in 16 bats
2. Differential expression analysis of non-coding RNAs

## Annotation of ncRNA in 16 bats

Coordinator
Martin


## Annotation of ncRNA in 16 bats

Coordinator
Martin

ncRNA classes

- tRNAs
- snoRNAs
- miRNAs
- IncRNAs
- Mitochondrial annotation
- And others (e.g. snRNAs)


## Annotation of ncRNA in 16 bats

rRNA

1. RNAmmer $(\mathrm{v} 1.2)^{2}$

- Hidden markov models
$\rightarrow 5.8 \mathrm{~S}, 18 \mathrm{~S}$ and 28 S rRNA
${ }^{2}$ K. Lagesen et al. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. 2007.


## Annotation of ncRNA in 16 bats

tRNA

1. tRNAscan-SE ${ }^{3}$

- Default parameters

2. Remove 'Undet' or 'Pseudo' types
[^1]
## Annotation of ncRNA in 16 bats

## snoRNA, miRNA and others



1. Gorap ${ }^{4}$ (uses Infernal) with alignments from the $\mathrm{Rfam}^{5}$ data base
[^2]
## Annotation of ncRNA in 16 bats

## snoRNA, miRNA and others



1. Gorap ${ }^{4}$ (uses Infernal) with alignments from the $\mathrm{Rfam}^{5}$ data base
2. For snoRNAs:

- Classification of C/D box and H/ACA box

[^3]
## Annotation of ncRNA in 16 bats

## miRNA



- miRDeep2 $(v 2 \cdot 0.0 .8)^{6}$
- Input:
- Combined smallRNA-Seq data set ${ }^{7}$
- Mapped to each individual bat assembly

[^4]
## Annotation of ncRNA in 16 bats

## IncRNA

Data

- LNCipedia (v5.2) ${ }^{8}$ data base

- High confidence set:
- 107,039 transcript of potential human IncRNAs

[^5]
## Annotation of ncRNA in 16 bats

## IncRNA

Data

- LNCipedia (v5.2) ${ }^{8}$ data base
- High confidence set:
- 107,039 transcript of potential human IncRNAs

Tool

1. $\mathrm{BLASTn}^{9}\left(\mathrm{v} 2.7 .1+, 1 \mathrm{e}^{-10}\right)$
[^6]
## Annotation of ncRNA in 16 bats

## IncRNA

Data

- LNCipedia (v5.2) ${ }^{8}$ data base
- High confidence set:
- 107,039 transcript of potential human IncRNAs

Tool

1. BLASTn $^{9}\left(\mathrm{v} 2.7 .1+, 1 \mathrm{e}^{-10}\right)$
2. Filter and re-structure the result
$\rightarrow$ Gene - transcript - exon structure
$\rightarrow$ Indroduce IncRNA hot spots
[^7]
## Annotation of ncRNA in 16 bats

## Mitochondrial annotation

Data


- 10 NCBI mitogenomes
- 1 blasted mitogenome
- Rearrange the mitogenome

[^8]
## Annotation of ncRNA in 16 bats

## Mitochondrial annotation

Data


- 10 NCBI mitogenomes
- 1 blasted mitogenome
- Rearrange the mitogenome


## Tool

- MITOS2 ${ }^{10}$
$\rightarrow$ Protein coding and non-coding RNA annotation

[^9]
## Annotation of ncRNA in 16 bats

Finalization

- Check gtf format



## Annotation of ncRNA in 16 bats

Finalization

- Check gtf format
- Merge all annotations for each bat
- Check for overlaps:


1. Within the new annotations
2. In the existing NCBI annotations

## Annotation of ncRNA in 16 bats

Finalization

- Check gtf format
- Merge all annotations for each bat
- Check for overlaps:


1. Within the new annotations
2. In the existing NCBI annotations

- Produce nice html tables for each annotation
- Automated csv and xlsx generation


## Annotation of ncRNA in 16 bats

Finalization

- Check gtf format
- Merge all annotations for each bat
- Check for overlaps:


1. Within the new annotations
2. In the existing NCBI annotations

- Produce nice html tables for each annotation
- Automated csv and xlsx generation
tRNAs



## Results



Maximal number of newly annotated RNAs for each RNA class. Newly annotated IncRNAs: 286805

## Results

- Final annotation for each bat in gft format
- Annotations for each ncRNA class and bat $\rightarrow$ Compatible and useable annotations


## Hackaton



1. Annotation of non-coding RNAs in 16 bats
2. Differential expression analysis of non-coding RNAs

## Differential expression analysis of non-coding RNAs

Data

- 6 RNA-Seq data sets
- 98 samples in total
- From 4 different bat species


## Differential expression analysis of non-coding RNAs

Pipeline<br>- Preprocessing with Trimmomatic (v0.36) ${ }^{11}$

[^10]
## Differential expression analysis of non-coding RNAs

## Pipeline

- Preprocessing with Trimmomatic (v0.36) ${ }^{11}$
- Mapping with HISAT (v2.1.0) ${ }^{12}$
- Each sample individually
$\rightarrow 1568$ mappings in total

[^11]
## Differential expression analysis of non-coding RNAs

## Pipeline

- Preprocessing with Trimmomatic (v0.36) ${ }^{11}$
- Mapping with HISAT (v2.1.0) ${ }^{12}$
- Each sample individually $\rightarrow 1568$ mappings in total
- Counting with featureCounts (v1.6.3) ${ }^{13}$
- Only unique mapped reads

[^12]
## Differential expression analysis of non-coding RNAs

## Analysis

- Differential gene expression analyses with DESeq2 ${ }^{14}$
- DESeq2 normalization
$\rightarrow$ Pairwise comparisons
$\rightarrow$ Significantly ${ }^{15}$ differentially expressed ncRNAs

[^13]
## Differential expression analysis of non-coding RNAs

## Analysis

- Differential gene expression analyses with DESeq2 ${ }^{14}$
- DESeq2 normalization
$\rightarrow$ Pairwise comparisons
$\rightarrow$ Significantly ${ }^{15}$ differentially expressed ncRNAs
- TPM (transcripts per million) for each ncRNA in each sample $\rightarrow$ Normalized expression level of each ncRNA

[^14]
## Preliminary results

- RNA-Seq data set: Field-2015 ${ }^{16}$
- 5 mock samples
- 6 infected (white-nose syndrome, WNS) samples


[^15]
## Preliminary results



PCA on ncRNAs.

## Preliminary results



## Preliminary results



Expression levels of significantly differentially expressed genes.

## Preliminary results



Expression levels of significantly differentially expressed genes.

## What is next?

- Analyze the other RNA-Seq data sets
- Make the annotations and results available


## What is next?

- Analyze the other RNA-Seq data sets
- Make the annotations and results available
- Hack the 16 new NCBI assemblies
- Bat1K project ${ }^{17}$ : sequence the genomes of all living bat species

[^16]Thanks to


- Manja Marz
- Martin Hölzer
- Nelly Fernanda Mostajo Berrospi
- RNA Bioinformatics \& High-Throughput Analysis Jena



## Genome quality



Icarus plot of the 16 investigated bat species: assembly lengths, N50 and N75 values.


[^0]:    ${ }^{1}$ B. Bagemihl. Biological Exuberance: Animal Homosexuality and Natural Diversity. 1999.

[^1]:    ${ }^{3}$ T. M. Lowe and S. R. Eddy. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. 1997.

[^2]:    ${ }^{4}$ github.com/koriege/gorap
    ${ }^{5}$ I. Kalvari et al. Rfam 13.0: shifting to a genome-centric resource for non-coding RNA families. 2017.

[^3]:    ${ }^{4}$ github.com/koriege/gorap
    ${ }^{5}$ I. Kalvari et al. Rfam 13.0: shifting to a genome-centric resource for non-coding RNA families. 2017.

[^4]:    ${ }^{6}$ M. R. Friedländer et al. miRDeep2 accurately identifies known and hundreds of novel microRNA genes in seven animal clades. 2012.
    ${ }^{7}$ Unpublished data, provided by Friedemann Weber, Justus-Liebig-Universität Giessen

[^5]:    ${ }^{8}$ P.-J. Volders et al. LNCipedia 5: towards a reference set of human long non-coding RNAs. 2019.
    ${ }^{9}$ S. F. Altschul et al. Basic local alignment search tool. 1990.

[^6]:    ${ }^{8}$ P.-J. Volders et al. LNCipedia 5: towards a reference set of human long non-coding RNAs. 2019.
    ${ }^{9}$ S. F. Altschul et al. Basic local alignment search tool. 1990.

[^7]:    ${ }^{8}$ P.-J. Volders et al. LNCipedia 5: towards a reference set of human long non-coding RNAs. 2019.
    ${ }^{9}$ S. F. Altschul et al. Basic local alignment search tool. 1990.

[^8]:    ${ }^{10}$ M. Bernt et al. MITOS: Improved de novo metazoan mitochondrial genome annotation. 2013.

[^9]:    ${ }^{10}$ M. Bernt et al. MITOS: Improved de novo metazoan mitochondrial genome annotation. 2013.

[^10]:    ${ }^{11}$ A. M. Bolger et al. Trimmomatic: A flexible trimmer for Illumina sequence data. 2014.
    ${ }^{12}$ D. Kim et al. HISAT: a fast spliced aligner with low memory requirements. 2015.

[^11]:    ${ }^{11}$ A. M. Bolger et al. Trimmomatic: A flexible trimmer for Illumina sequence data. 2014.
    ${ }^{12}$ D. Kim et al. HISAT: a fast spliced aligner with low memory requirements. 2015.
    ${ }^{13}$ Y. Liao et al. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. 2014.

[^12]:    ${ }^{11}$ A. M. Bolger et al. Trimmomatic: A flexible trimmer for Illumina sequence data. 2014.
    ${ }^{12}$ D. Kim et al. HISAT: a fast spliced aligner with low memory requirements. 2015.
    ${ }^{13}$ Y. Liao et al. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. 2014.

[^13]:    ${ }^{14}$ M. I. Love et al. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. 2014.
    ${ }^{15}$ Adjusted p-value $<0.05$; absolute log 2 fold change $>2$

[^14]:    ${ }^{14}$ M. I. Love et al. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. 2014.
    ${ }^{15}$ Adjusted p-value $<0.05$; absolute $\log 2$ fold change $>2$

[^15]:    ${ }^{16}$ K. A. Field et al. The white-nose syndrome transcriptome: activation of anti-fungal host responses in wing tissue of hibernating little brown myotis. 2015.

[^16]:    ${ }^{17}$ https://bat1k.ucd.ie/

