# Prediction of structural non-coding RNAs by comparative sequence analysis

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# Outline

- 1. Introduction to non-coding RNAs and motivation of this work
- 2. New algorithms for detection of structural non-coding RNAs
- 3. A large scale screen of the human genome
- 4. Other applications

# Non-coding RNAs

Non coding RNAs ("RNA genes") are transcripts that exert their function as RNA whithout being translated to protein.

- "Classical" examples:
  - Protein expression: transfer RNA, ribosomal RNA
  - Pre-mRNA splicing: spliceosomal RNAs
  - tRNA maturation: Ribonuclease P
  - Protein export: Signal recognition particle RNA
- New abundant classes of small non-coding RNAs: microRNAs, snoRNAs
- Many other examples are currently emerging in all organisms studied.

# Motivation



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- 1. A vast amount of genomic data is available
- 2. There are fewer protein coding genes than expected



3. Highly conserved non-coding DNA awaits functional annotation.



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- 4. The transcriptional map of the human genome is much more complex than expected.



Non-coding RNAs ?

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# Computational identification of non-coding RNAs

Based on a priori knowledge: find members of known families

- Sequence similiarity alone: BLASTN
- Sequence and additional motif information: specialized programs for e.g. tRNA or snoRNAs
- De novo prediction: find new genes and families
  - Unlike protein coding genes (ORFs, codon bias,...) ncRNAs lack strong statistical signals in primary sequence
  - The function of many ncRNA depend on a defined secondary structure

# Can secondary structure predictions be used for ncRNA detection?

# Significance of predicted RNA secondary structures: z-score statistics

- Has a natural occuring RNA sequence a lower MFE than random sequences of the same size and base composition?
  - 1. Calculate native MFE m.
  - 2. Calculate mean  $\mu$  and standard deviation  $\sigma$  of MFEs of a large number of shuffled random sequences.
  - 3. Express significance in standard deviations from the mean as *z*-score

$$z = \frac{m - \mu}{\sigma}$$

 Negative z-scores indicate that the native RNA is more stable than the random RNAs.

#### z-scores for 579 tRNAs



- ▶ Only 2% below a *z*-score threshold of -4.
- Native sequences are not clearly separated from the random bulk.

# Consensus folding using RNAalifold

- RNAalifold uses the same algorithms and energy parameters as RNAfold
- Energy contributions of the single sequences are averaged
- Covariance information (e.g. compensatory mutations) is incorporated in the energy model.
- It calculates a consensus MFE consisting of an energy term and a covariance term:

#### z-scores of consensus MFEs for tRNA alignments



 Alifoldz: Additional information from aligned sequences shifts MFE predictions towards significant levels.

#### The structure conservation index



The SCI is an efficient and convenient measure for secondary structure conservation.

#### Efficient calculation of stability z-scores

- The significance of a predicted MFE structure can be expressed as z-score which is normalized w.r.t. sequence length and base composition.
- Traditionally, z-scores are sampled by time-consuming random shuffling.
- The shuffling can be replaced by a Support Vector Machine regression calculation which is of the same accuracy.



#### SVM classification based on both scores



 Both scores separate native ncRNAs from controls in two dimensions.

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- RNAz: more accurate and faster than any other available programs.

# Screening the human genome



- Large scale comparative screen of mammals/vertebrates
- ► ≈ 5% of the best conserved non-coding regions
- ► → 438,788 alignments covering 82.64 MB (2.88% of the genome)

#### Statistics of detected structures



# Novel structural RNAs of known classes: mirRNAs and H/ACA snoRNAs



#### Novel structures of unknown function



# Other applications: Cyanobacterial ncRNAs



 I. Axmann, P. Kensche et al. (Genome Biol. 6:R73, 2005) identified and characterized 7 novel ncRNAs in cyanobacteria using Alifoldz. Other applications: Benchmarking alignment programs on structural RNAs



The SCI can be used to assess the quality of an alignment of a structural RNA (P. Gardner, A. Wilm & S. Washietl Nucleic Acids Res. 33:2433, 2005).

# Other applications

- RNAz screen of urochordate genomes (K. Missal, D. Rose, P.F. Stadler *Bioinformatics* 21: Suppl 2,ii77-ii78, 2005)
- RNAz screen of nematode genomes (K. Missal *et al. J. Exp. Zoolog. B*, in press).
- Prediction of putative miRNA precursors in the miRNAMap (Hsu *et al.*, submitted)

# Summary and Conclusions

- De novo ncRNA prediction is notoriously difficult.
- Single sequence methods are of limited statistical significance.
- Comparative approaches dramatically improve accuracy.
- RNAz is an accurate and efficient approach for predicting ncRNAs.
- RNAz used for the first comprehensive annotation of conserved RNA secondary structures in the human genome.
- The data provides a strong basis for further computational and experimental studies.
- The programs and methods presented here were successfully used in a variety of other applications.

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