

Fragment based detection of ncRNAs

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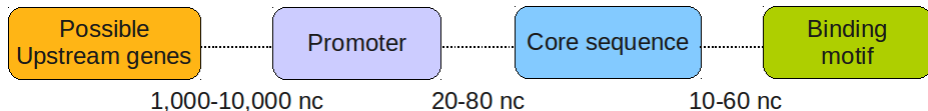
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Introduction

- detection of ncRNAs is an important task in genome annotation
- many ncRNA classes and families known
- finding members in other species can be rather complex
- in most cases a combination of tools is necessary
- candidates have to be scored



Complexity of ncRNA detection - Features

- sequence similarity
- secondary structure
- promoter data
- terminator data
- protein interaction sites
- RNA interaction
- synteny
- specific distances

Complexity of ncRNA detection - Tools

- sequence: blast, GotohScan
- motifs: RNABOB, fragrep, RNAmotif
- secondary structure: RNA Vienna Package, RNAshape
- RNA-interaction: Petcofold, RIPalign
- terminators: TranstermHP
- covariance models: Infernal

Bioinformatician

- build a basically new pipeline for each RNA class
 - 1 evaluate features
 - 2 find suitable tools
 - 3 plug them together
 - 4 combine and assess results
 - 5 score the results
- time intensive
- manual labor

Complexity of ncRNA detection - Result

Bioinformatician

- build a basically new pipeline for each RNA class
 - 1 evaluate features
 - 2 find suitable tools
 - 3 plug them together
 - 4 combine and assess results
 - 5 score the results
- time intensive
- manual labor

Biologist



Fragment based approach

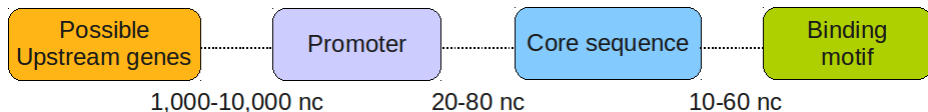
Deals with

- 1 evaluate features
- 2 (find suitable tools)
- 3 plug them together
- 4 combine and assess results
- 5 score the results

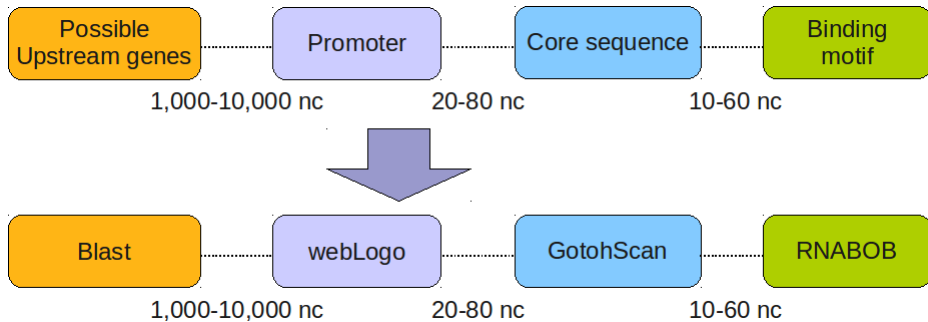
Applicable for

- bioinformatician with expert knowledge about the tools
 - command line tool
 - all-in-one online tool
- scientists with limited background in computer science
 - all-in-one online tool

Starting point



Starting point



Core problem

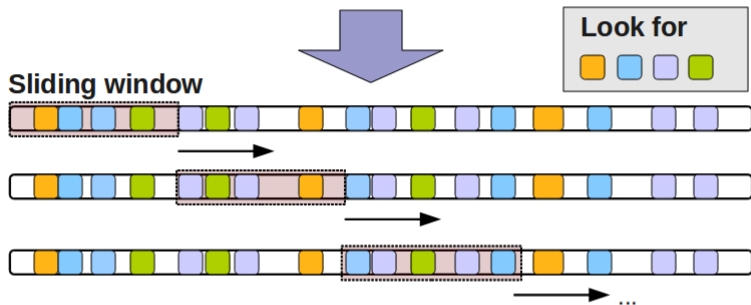
Tools



Combined positions on genome

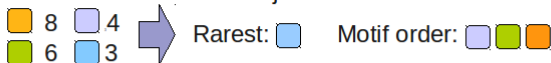


Sliding window



Evaluations step

How often does which object occur?



Recursion step (for each window)

For each rarest object
extend the window to match maximal spread
findNextMotif()



Example

Find rarest



Extend window

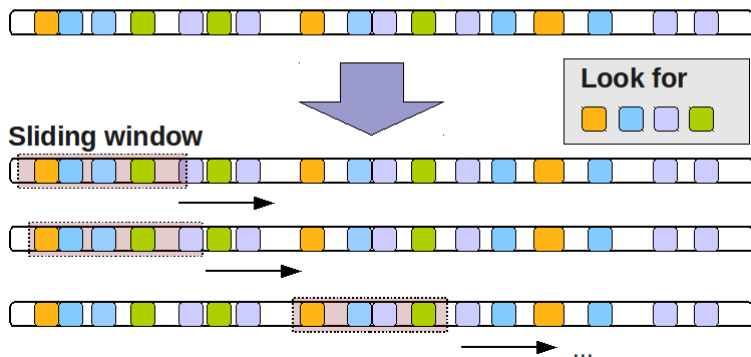
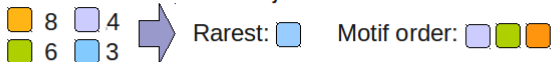


Find next...



Evaluations step

How often does which object occur?



Online tool

Search options

Submit Query

Motif 1			
Naming	Distance to motif 0	Program	Program details
Name: upstream_gene	Min.: <input type="text"/> Max.: <input type="text"/>	Blastn	E-Value: 1e-10 Fasta sequence(s): CCAACTCCGG TGAGCCTTAGTTGCTGGGACTGGAGGCTCT CCCTCTTGGCATTATCTCCAGATCGAGAA

Motif 2			
Naming	Distance to motif 1	Program	Program details
Name: promoter	Min.: 1000 Max.: 10000	Sequence	Sequence: TATAAT Strand: + only Mutations: 2

Motif 3			
Naming	Distance to motif 2	Program	Program details
Name: ncRNA	Min.: 10 Max.: 100	Sequence	Sequence: TGAGCCTTAGTGTCT Strand: + only Mutations: 5

Submit Query

Data format

```
motif: 1
name: upstream_gene
method: Blastn
evalue: 1e-10
fasta: EMBEDDED
>WMT1
TTGCCAACACAAGACTCGGTATTGATGCTACCCATTATCTCAACCACCTCCTGACCGACCCCAACTCCCG
TGAGCCTTTAGTTGCTGCGACTGGAGGTCTCCCTCTTGCCATTATCTCCAAGATCGAGAATGATTTGCGT
GCTCTCGAGCGCCATGCCATCAAACCCGTCTTCGTGTTCCCCGGCCTTCCGCTTGCTTCTCGACCTCTC
CTAAGGGTCCCAGATATCAAGGCTGAGCGAGAAAACAGATTAAGAATGAGGCCTGGGCGCTTTATGATGA
END_EMBEDDED
```

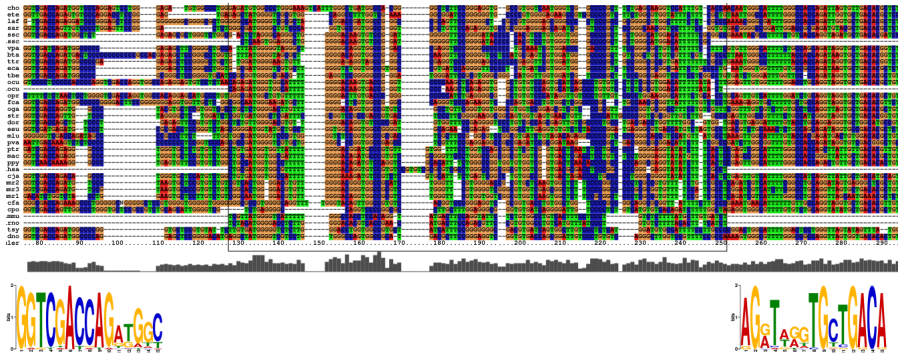
```
motif: 2
name: promoter
mindistance: 1000
maxdistance: 10000
method: Sequence
seq: TATAAT
strand: + only
mutations: 2
```

```
motif: 3
name: ncRNA
mindistance: 10
maxdistance: 100
method: Sequence
seq: TGAGCCTTTAGTTGCT
strand: + only
mutations: 5
```

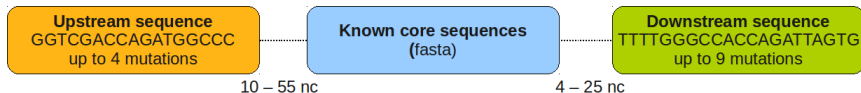
Does it work? - A test with paRNAs

paRNA

- promoter associated RNAs
- complementary in sequence to the rDNA promoter
- needed for rDNA methylation and silencing



Does it work? - A test with paRNAs



Run with genome of *Bos taurus*

M4		124452425	124452607	MATCH_1 .	+	
#MATCH_1	M4	124452425	124452441	up	.	[GGTCGACCAGATGACTC]
#MATCH_1	M4	124452471	124452577	paRNA	9e-50	+
#MATCH_1	M4	124452587	124452607	down	.	[TTTTCTACCACCAGATAAGCA]
MUn.004.5853		2895	3083	MATCH_2 .	+	
#MATCH_2	MUn.004.5853	2895	2911	up	.	[GGTCGACCAGATGACTC]
#MATCH_2	MUn.004.5853	2945	3043	paRNA	1e-48	+
#MATCH_2	MUn.004.5853	3063	3083	down	.	[TTTTTTACCACCAGATAAGTG]
M21		52061934	52062120	MATCH_3 .	-	
#MATCH_3	M21	52061934	52061954	down	.	[TTTTTTACCACCAGGTAAGTG]
#MATCH_3	M21	52061964	52062070	paRNA	9e-50	-
#MATCH_3	M21	52062104	52062120	up	.	[GGTCGACCAGATGACTC]
MUn.004.7994		2594	2789	MATCH_4 .	-	
#MATCH_4	MUn.004.7994	2594	2614	down	.	[TTTTTTACCACCAGGTAAGTG]
#MATCH_4	MUn.004.7994	2624	2730	paRNA	4e-52	-
#MATCH_4	MUn.004.7994	2773	2789	up	.	[GGTCGACCAGATGACTC]

Does it work? - A test with paRNAs

bta	4	mmr	13
cfa	1	mmu	1
cho	6	ocu	37
cja	7	oga	1
cpo	3	opr	30
dno	6	pca	2
dor	33	ppy	1
eca	8	ptr	1
eeu	1	pva	4
ete	10	rno	2
fca	7	sar	10
hsa	0	ssc	1
laf	27	str	6
mac	15	tbe	20
mlu	4	tsy	1
		ttr	12

Have

- all-in-one tool
- web interface
- command line interface
- export and import function
- replicability
- extendable
- parallelization
- .bed output

Outlook

- add more tools
- some applications
- add chose from a list of genomes instead of upload
- predefined search patterns
- flexible scoring scheme
- .bed input
- more fancy output
- output of necessary citations
- small tutorial scientists with limited background in computer science

Thank you