

The majority of long non-coding RNAs is conserved

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joint work with Anne Nitsche, Dominic Rose, Mario Fasold

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mRNA-like ncRNAs

over the last few years ncRNAs that otherwise look quite similar to mRNAs have become a major research topic
(using, as usual, a variety of acronyms) mlncRNA, lincRNAs,

- How well conserved are lncRNAs?

Two answers:

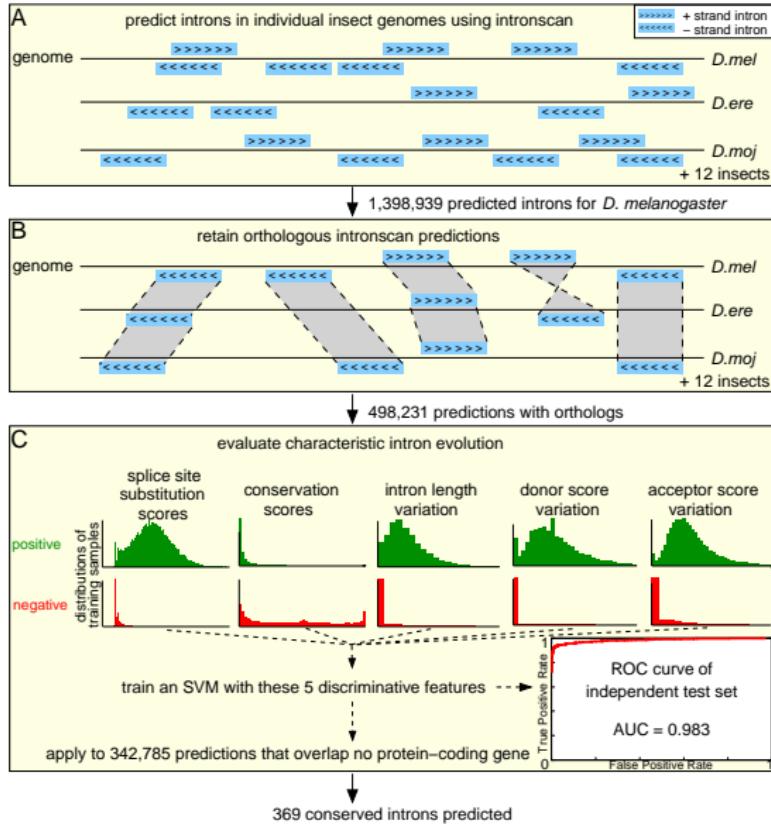
- ① “relatively low degree of sequence constraint”
(Marques & Ponting 2009)
- ② but ... some very well-conserved examples
(Chodroff *et al.* 2010, ...)

One problem: sequence conservation does not necessarily imply conservation of the ncRNA!

De novo Prediction of mRNA-like ncRNAs

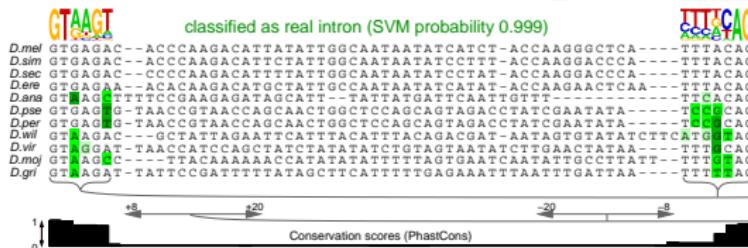
- long ncRNA = contains at least one intron
- predict non-coding transcripts by predicting **conserved short** introns
- Why introns?
 - intron evolution is slow and essentially independent of the evolution of the mature sequence
 - splice sites are often conserved
 - disruption of correct splicing usually destroys function
 - ! non-coding transcripts do not have randomly placed large in/dels.
- Why short introns?
 - Most *Drosophila* introns are short.
 - Can be accurately predicted (94% with both splice sites correct)
- **Intron prediction** (Lim & Burge 1999): machine learning using patterns of donor, acceptor, intron length, branch point, intron composition

Intron-prediction pipeline

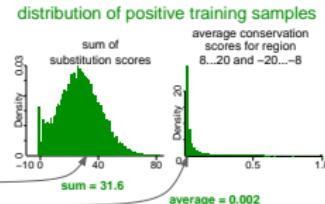


Evaluation of Introns: Species-Specific Patterns

A

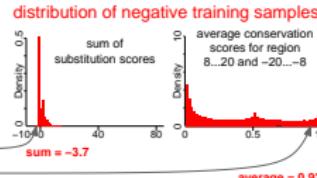
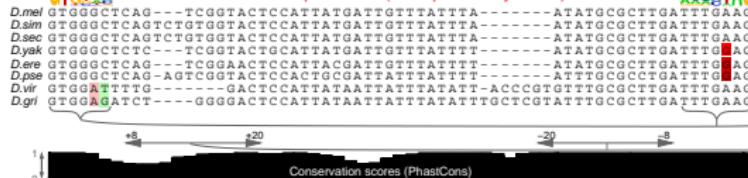


B

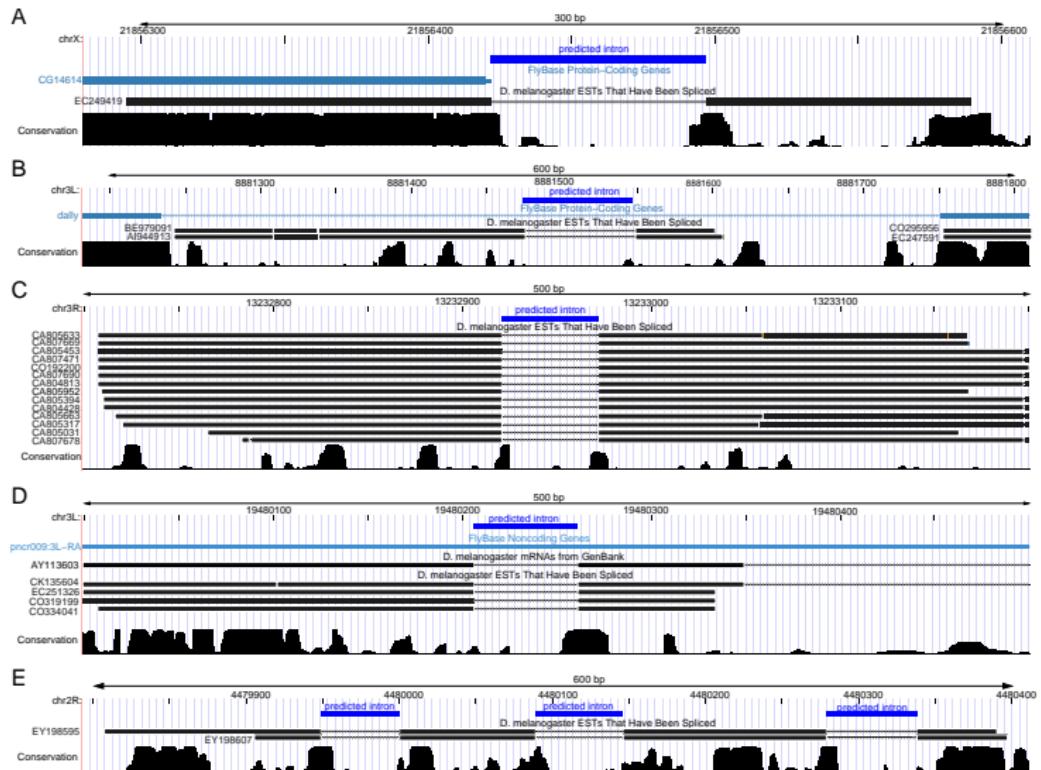


GTAACT

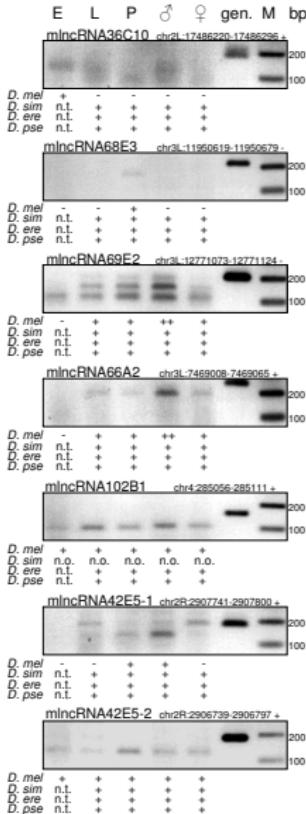
classified as false prediction (SVM probability 0.001)



Validation with un-annotated ESTs



Novel conserved ncRNAs in *Drosophila*



11 out of 17 predictions
verified by PCR and
sequencing

Expression of transcripts
and existence of introns
also verified in 3 other fly species

Embryo

Larva

Pupa

male

female

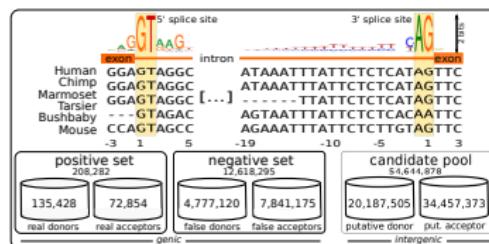
Functional mIncRNA in Vertebrates

- vertebrate introns \neq insect introns
(2% vs 54% short introns)
- predict single individual splice-sites
(instead of introns)
- splice-site prediction ! = intron prediction
vertebrate exons are still short, so let's predict exons
(novel pipeline, new SVM features, re-implementation)
downside: single-intron genes are not visible

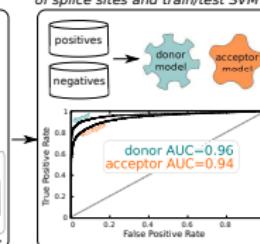
Conserved mlncRNA in Vertebrates

A) Splice site prediction

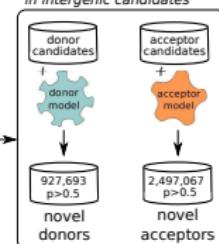
1. Scan alignments for splice sites, prepare and partition data



2. Compute evolutionary signatures of splice sites and train/test SVM

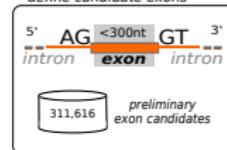


3. Predict novel splice sites in intergenic candidates

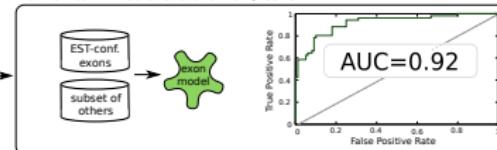


B) Exon prediction

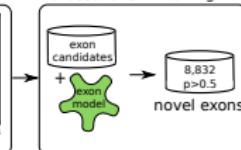
1. AG/GT splice sites pairs define candidate exons



2. Compute evolutionary signatures of EST-confirmed exons and train/test SVM

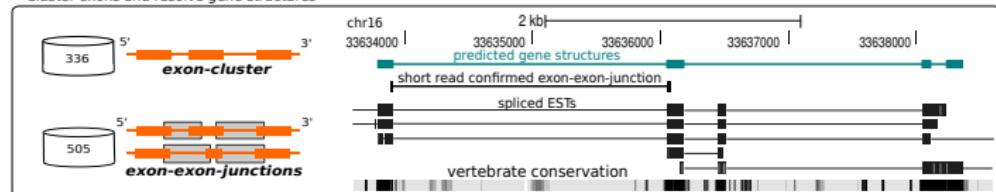


3. Classify exons which were not used for SVM training



C) Transcript prediction

Cluster exons and resolve gene structures



Conserved mlncRNA in Vertebrates

A) Alignment

evaluate substitution patterns
example: column -5

intron acceptor exon

19 5 1 :::

```

TCCCTGCACCTCCTCTCTAGA Human
TCCCTGCACCTCCTCTCTAGA Marmoset
CCCGTGTGTTCTCCCCAGA Lemur
GCCTCGTGTTCCTCCCCAGA Bushbaby
CTGCTGCACTTCTCCGGAGA Tree Shrew
TTTTTTATTTCTGCCAGA Mouse
CTCTTCTATTCTCTGCTAGA Rat
TCCCTGTACTTCTTCCCAGA Squirrel
TCCCCGGCTCTCTCCGGAGA Alpaca
TCCCCGGCTCTCTCCGGAGA Dolphin
TCTCTGCTGCTTCTCCCCAGA Horse
TCTTGGCGCTTCTCCCCAGA Cat
TCTTGTCGCTTCTCTCAGA Dog
CTCCCCGGCTTCTCCCCAGA Megabat
CCCTTGCTGCTTCTCCCCAGA Hedgehog
CCCTTGCGCTGCTTCTCCCCAGA Elephant
TCTTTGTCGCTGCTTCCCCAGA Rock Hyrax
GCCTCCCTGGCTTCTCCAGG Armadillo
TCCCTGCGCTTCTACCTAGA Sloth
GTTTTGTCGCTTCTCTAGA Opossum
TTCCTGGGTTCTCTCTAGA Platypus
CCCCCTCCCCGTCTCGAGA Medaka

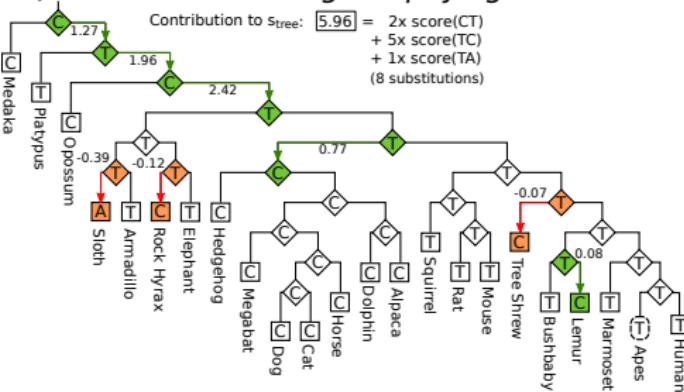
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B) Pairwise substitutions

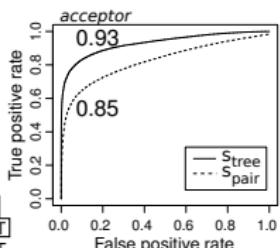
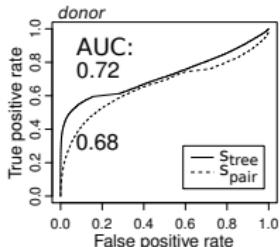
"human versus other species"

Contribution to s_{pair} : $8.08 = 12 \times \text{score(TC)} + 1 \times \text{score(TA)}$
 $= (12 \times 0.69) + (-0.20)$

C) Substitutions along the phylogenetic tree

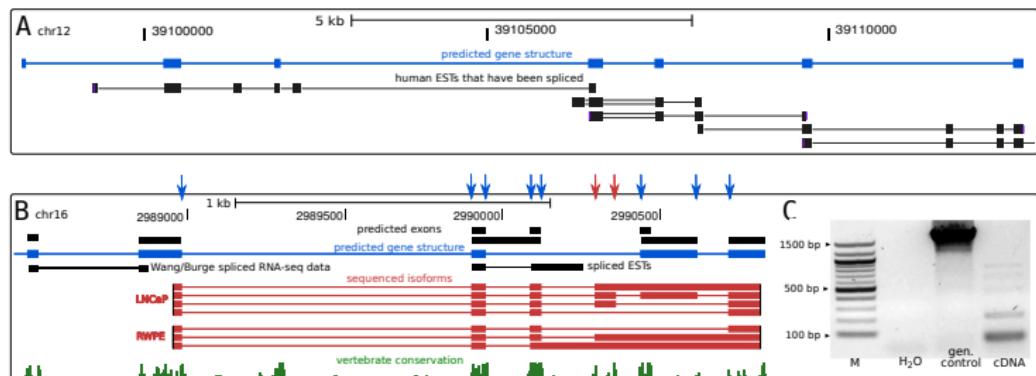


D) ROC analyses



Conserved mlncRNA in Vertebrates

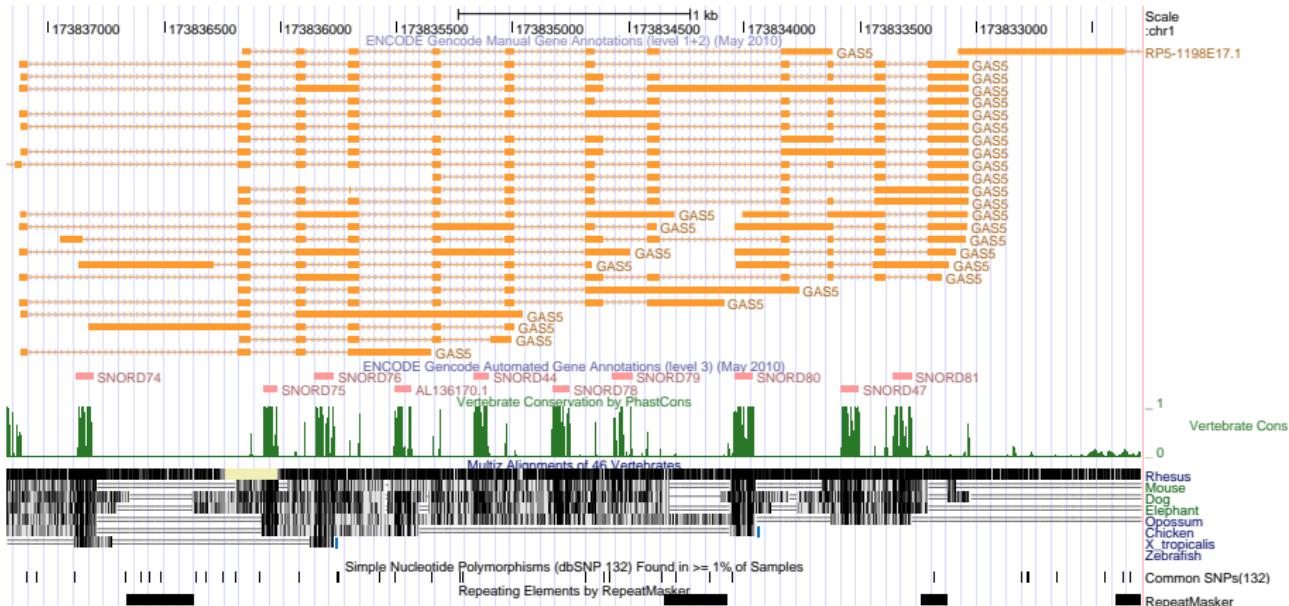
two validated examples



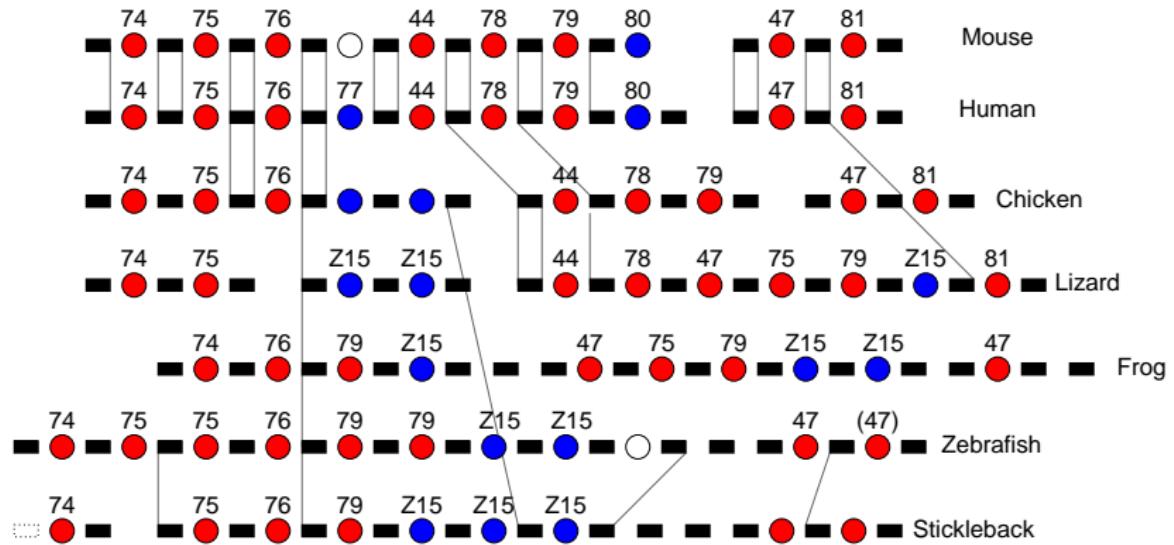
GAS5: A highly conserved host gene

- most famous snoRNA host gene, 10 different snoRNAs are the payload in its introns
- The exonic part (“mRNA”) sequesters and inhibits the glucocorticoid receptor (Kino, Sci. Signaling 2010)
- conserved at least in gnathostomes
- very rapid evolution of exonic sequence, snoRNAs well conserved
- major changes in gene structure

Human GAS5 – a complex locus



Evolution of GAS5



Two superimposed effects

- changes in the structure of the host gene itself
gain & loss of splice sites
- snoRNAs can be behave like mobile elements

Evolution of mlncRNAs: HOTAIR

- transcribed from the HOXC cluster in antisense direction from the HoxC12-HoxC11 intergenic region
- directs PRC2 to the HOXD locus, silencing HoxD11-HoxD8. [Rinn et al 2007, Tsai et al 2010]
- however, the mouse homolog does not have this function [Schorderet & Duboule 2011]

Evolution of m^lncRNAs: HOTAIR



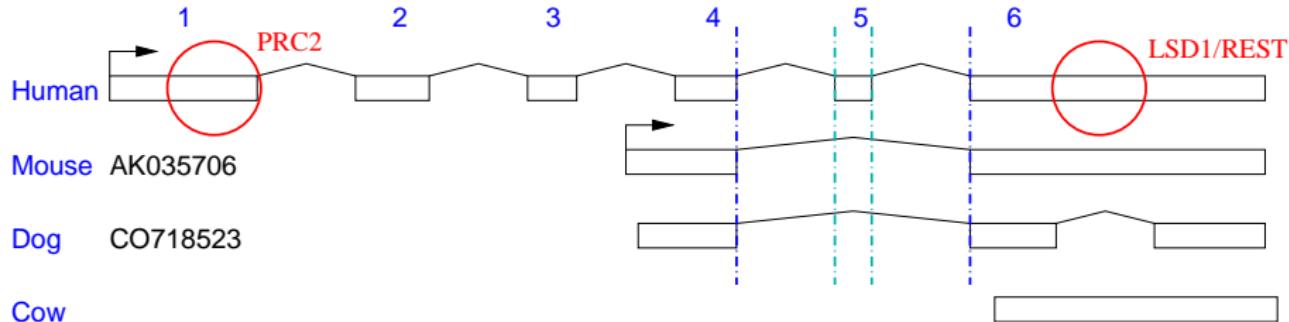
	Exon 1	Exon 2	Exon 3	Exon 4	Exon 5	Exon 6
Mouse	74%	48%	75%	86%	92%	49%
Dog	73%	64%	85%	91%	94%	55%
Cow	76%	71%	84%	91%	79%	68%
Elephant	75%	75%	77%	92%	-	62%
Armadillo	76%	66%	-	-	94%	61%
Opossum	-	-	-	83%	-	-
Platypus	27%	-	47%	55%	-	-
Latimeria	-	-	-	57%	-	-

Jan Engelhart

Problem: conservation of sequence does not imply conservation of the transcript.

Sequence conservation could be caused e.g. by cis-acting DNA elements

Evolution of mlncRNAs: HOTAIR



Schorderet P, Duboule D. (2011): Mouse HOTAIR has a different structure, presumably lacks PRC2 binding domain

Comparative Map of Splice Sites

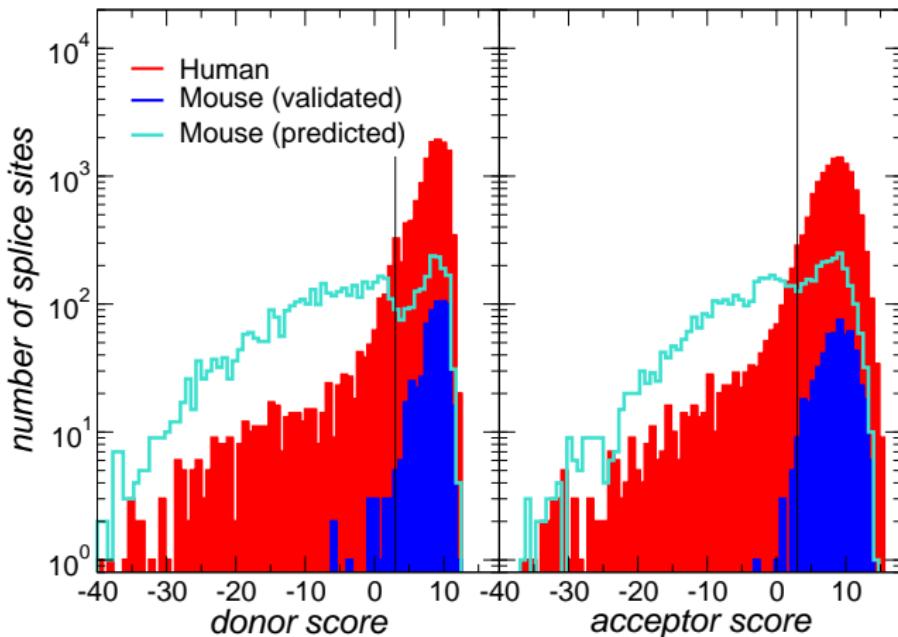
Simple idea:

- ① use a genome-wide multiple sequence alignment
 - ① UCSC 46-way multiz alignment
 - ② ENSEMBL 12-way EPO alignment
- ② map all splice sites that are experimentally known to the alignment
RefSeq plus all ESTs

Prediction of functional splice sites

Limited coverage of transcript data limit sensitivity.

Use splice site scoring scheme (here maxentscan scores) to estimate whether a splice site is conserved.



Score distribution of all alignable position is bi-modal:

UCSC 46-way map of splicesites

Splice site set: all ESTs and all RefSeq genes

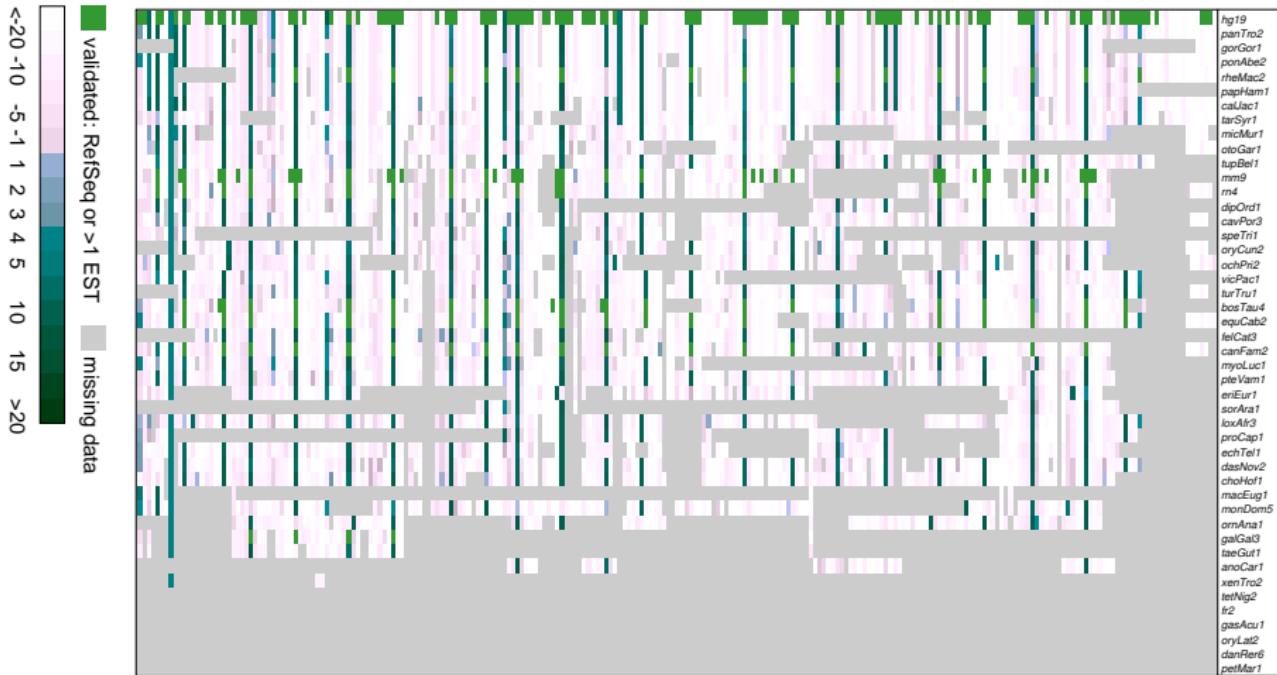
1257773 SS of which 387318 are contained in RefSeq genes

Conservation of RefSeq splice sites between human and mouse

	all	coding		3' UTR		5' UTR		non-coding	
hg19	387318	353836		1103		15200		17179	
aligned	362258	93.5	340617	96.2	834	76	11910	78.3	8888
predicted	338510	87.3	324504	91.7	670	61	7933	52.1	5403
validated	336913	86.9	325458	91.9	599	54	6734	44.3	4122
									31.4

... ncRNAs appear to be much less alignable than UTRs
hints at a problem with the genome-wide alignments

Splice Site Map for GAS5



we know GAS5 is conserved throughout vertebrates, but we have very little aligned sequence already in chicken & frog and nothing in teleosts.

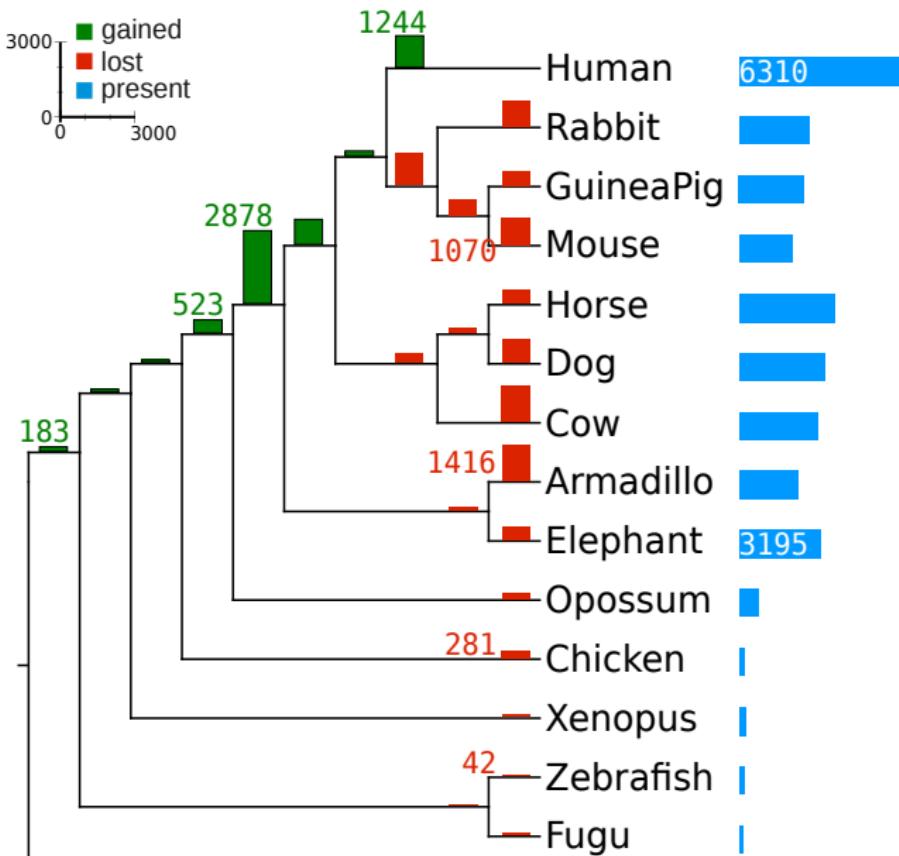
Conservation of lncRNAs

GENCODE set of 25644 splice sites in 6310 lncRNA transcripts

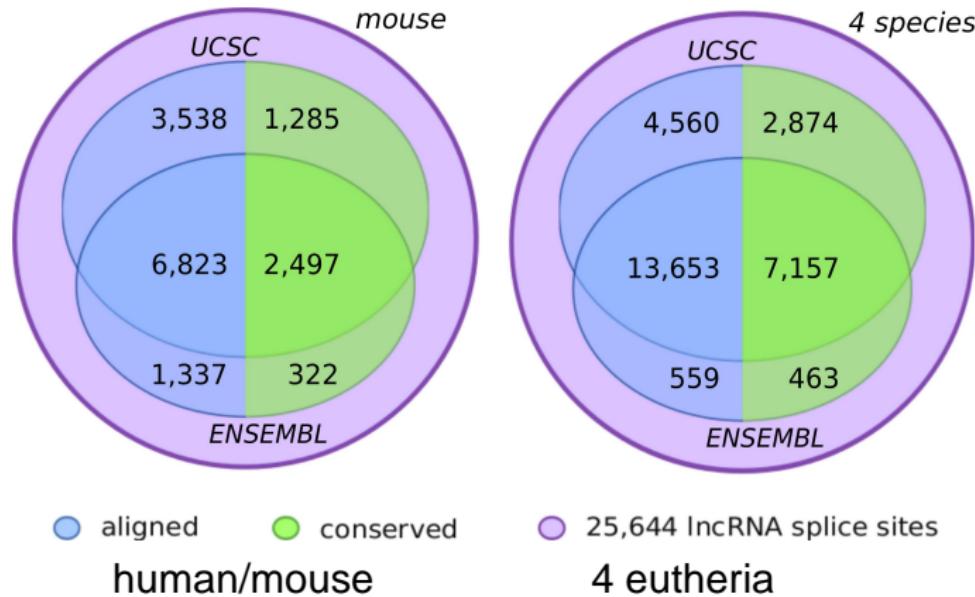
(removed all overlaps with coding regions and RNACode hits)

Species	splice sites		transcripts	
	cons.	val.	cons.	val.
mouse	3,809	1,155	2,098	362
rat	3,508	752	1,968	224
cow	6,214	1,047	3,096	351
dog	6,828	694	3,382	238
<i>union 5</i>	10,057	1,876	4,292	585
<i>union 15</i>	13,720	2,098	5,072	635

Gains and Losses



Comparison of UCSC and ENSEMBL Alignments



Conservation of special mlncRNA sets

	aligned	predicted	known
213 human transcripts hosting microRNAs			
mouse	151	91	15
dog	185	144	5
5 Eutheria	191	164	23
94 human transcripts hosting snoRNAs			
mouse	72	57	46
dog	81	70	40
5 Eutheria	84	74	51
2,076 mouse lncRNAs from [1]			
human	1,770	1,113	446
dog	1,628	944	185
4 Eutheria	1,776	1,237	472
1,508 zebrafish lncRNAs [2,3]			
Teleostei	953	513	112
Tetrapoda	476	170	56

[1] Guttmann *et al.* Nature 477: 295-300 (2011); [2] Pauli *et al.* Genome Res. 10.1101/gr.133009.111 (2011); [3] Ulitsky *et al.*

Cell 147: 1537-1550(2011)

P.F. Stadler (Leipzig)

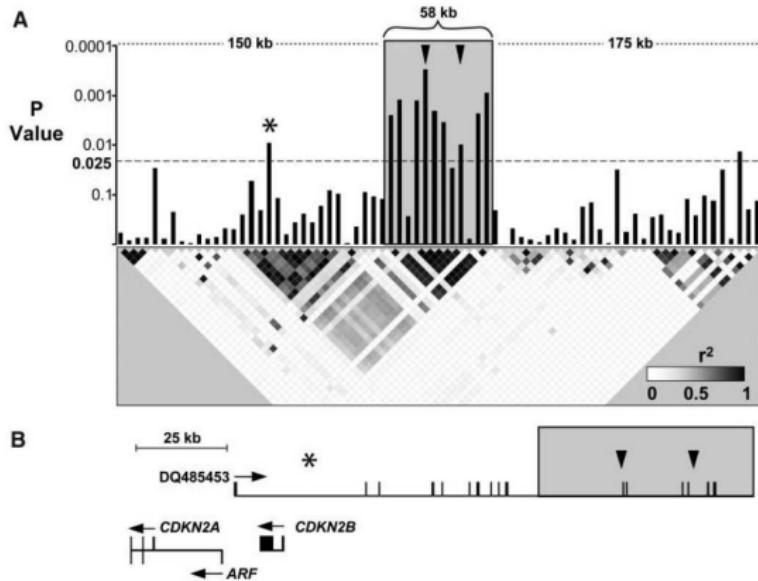
mlncRNAs

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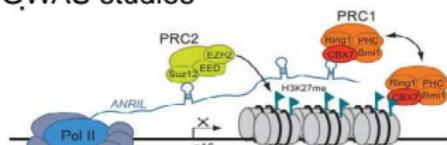
Most QTLs for complex multi-genic diseases hit noncoding regions

Association of coronary heart disease (CHD) with a 58kb region on chr. 9p21



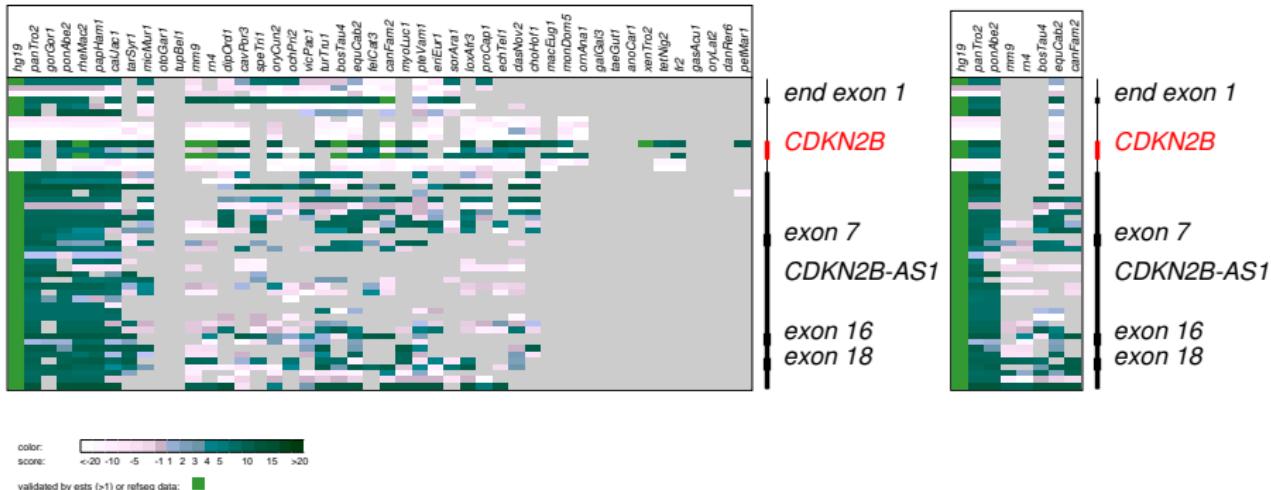
McPherson et al., Science (2007)

ANRIL transcript(s) in many isoforms
associated with the atherosclerosis risk
Holdt et al. (2010)
and it appears in many other GWAS studies

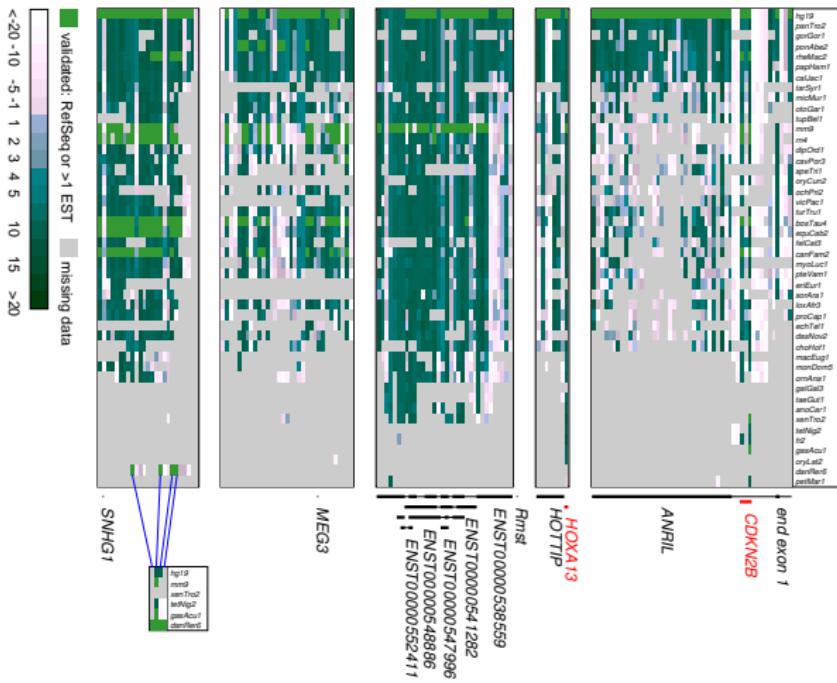


Yap et al (2010)

Conservation of ANRIL



Some more examples



Summary

- Conservation and substitution patterns of splice sites can be used to infer novel non-coding genes even in the absence of RNA secondary structure and
- As entities, mlnRNAs are evolutionarily much older and better conserved than their sequence
- Most mlnRNAs are conserved only in parts with rapid changes in gene structure
- Large repertoire of unspliced lncRNAs whose evolutionary patterns we do not yet understand

Many, many thanks ...

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FH RNomics group: Jörg Hackermüller, **Kristin Reiche**, Kathy Schutt, Kerstin Ullmann, ...
FG ncRNAs: **Friedemann Horn**, Thomas Arendt, **Kurt Engeland**, **Peter Ahnert**, ...
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- Dresden: **Michael Hiller**
- Marburg: Manja Marz and her group
- Freiburg: Rolf Backofen, **Dominic Rose**
- Copenhagen: Jan Gorodkin, Stefan Seemann, Peter Menzel, and the RTH
- Barcelona: Roderic Guigó, Andrea Tanzer
- Strassbourg: Catherine Florentz, Joern Pütz, Frank Jühling
- MIT: Stefan Washietl, Sebastian Will
- Affymetrix: Tom Gingeras, Phil Kapranov, *et al.*
- PICB Shanghai: Axel Mosig and Phil Khaitovich and their students (PICB/SIBS)
- ASU Tempe: Julian L. Chen and his lab
- ENCODE: Ewan Birney and $10^{2.5}$ coauthors
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