

Computational Estimation of Energy Parameters for modified Nucleotides

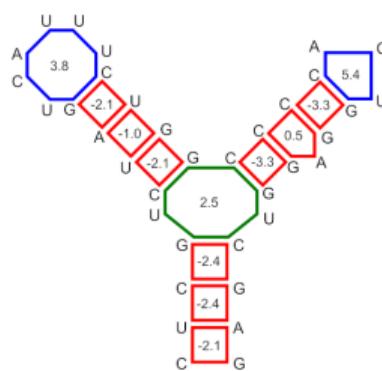
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Nearest Neighbor (NN) Model



$$E(S) = \begin{aligned} & U \square A + C \square G + G \square C + C \square G + \\ & U \square G + A \square U + C \square A + G \square U + A \square C + \\ & C \square G + C \square A + C \square G + C \square G + \\ & = -6.50 \text{ kcal/mol} \end{aligned}$$

- Secondary structures can be decomposed into loops formed by adjacent pairs
- A free energy contribution is assigned to each loop

$$E(S) \approx \sum_{l \in S} E(l)$$

- 294 independent nearest neighbor parameters included in the Turner NNDB¹
 - From 802 UV melting experiments
- **Virtually no energy parameters for modified bases**

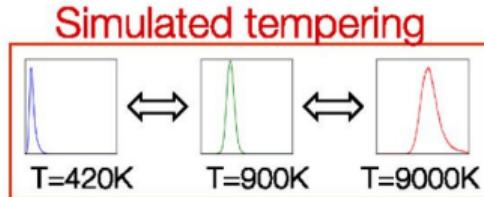
¹ Turner et al., "NNDB: The nearest neighbor parameter database for predicting stability of nucleic acid secondary structure.", 2009, NAR 38, D280–D282

Computational Estimations of Nearest Neighbor Parameters

- Coarse grained models VS Molecular dynamics
 - No solvent
 - Less degrees of freedom
 - Lower computational costs
- Coarse grained models seem to be the best choice
 - Rosetta-RECCES
 - Coarse grained moves
 - Energy calculation at atom level

RECCES² Simulation

- Monte Carlo simulation
- Temperature changes during the simulation
 - To overcome energy barriers and sample the full conformation space
- Energy of each conformation determined by an energy function
 - Combination of Physics- and knowledge-based terms

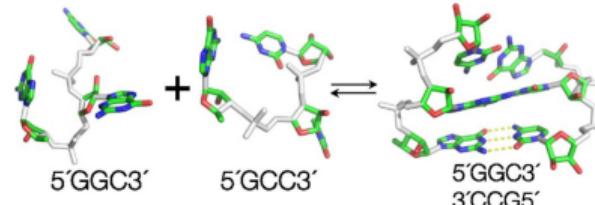


WHAM

Density of states

$$Z = \sum_{s \in \Omega} e^{-\beta E(s)}$$

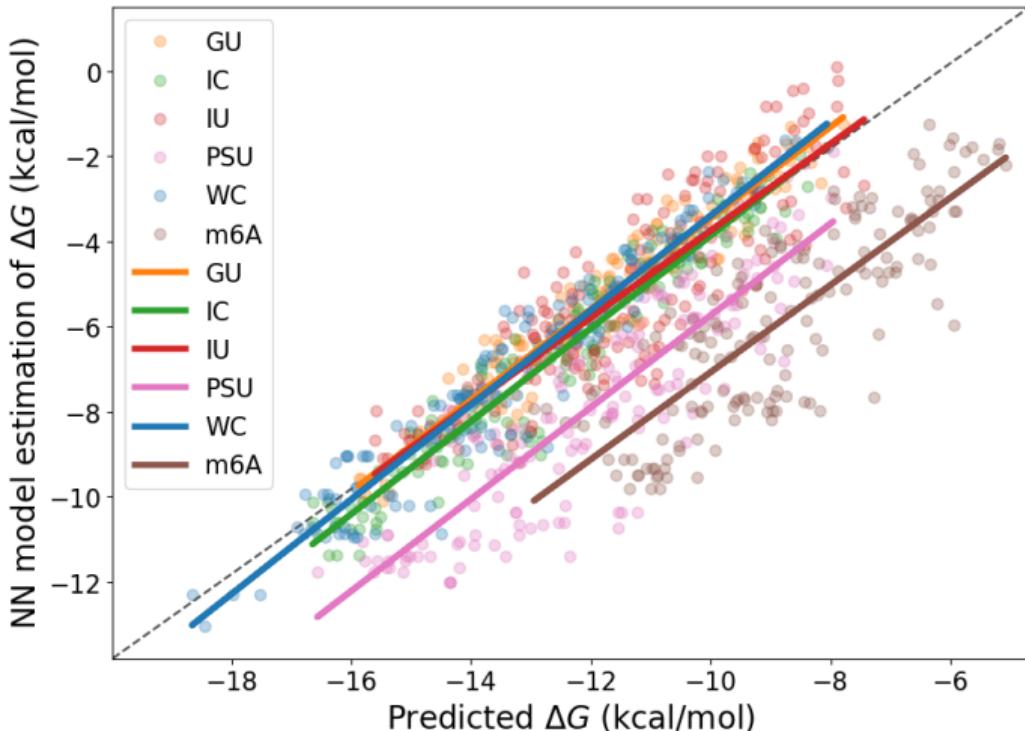
ΔG ΔH



$$\begin{aligned}\Delta G(5'GGC) &= \\ G(5'GGC) - G(GGC) - G(CCG)\end{aligned}$$

² Chou et al., "Blind tests of RNA nearest-neighbor energy prediction", 2016 Proceedings of the National Academy of Sciences, 113(30), 8430–8435.

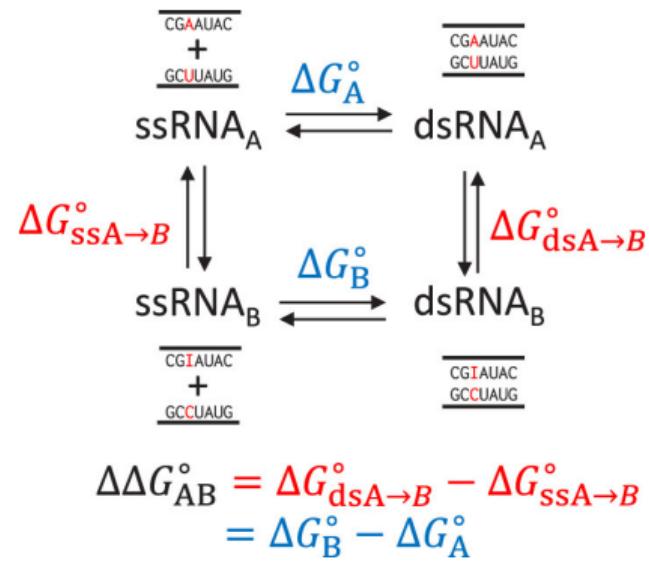
Comparison of free energy estimations



- Comparison to estimations using existing parameters
- Duplex of 3 to 5 base pairs
- "good" correlation
- Systematic error
- Modification dependent
- Rosetta energy function poorly parameterized?
- Water plays a role in the stability of $\Psi \bullet A$ stacks

Molecular dynamics (MD)

- free energies of proteins and their ligands calculated for decades
- studies on nucleic acids rather limited
- including only inosine as modification³
- standard MD based on thermodynamic cycle to calculate the free energies
- impossible to simulate the **chemical path**
- using **alchemical path**
 - insert intermediate state $0 = \lambda_1, \lambda_2, \dots, 1 = \lambda_n$
 - $\Delta G = \sum_i \Delta G_{i,i-1}$

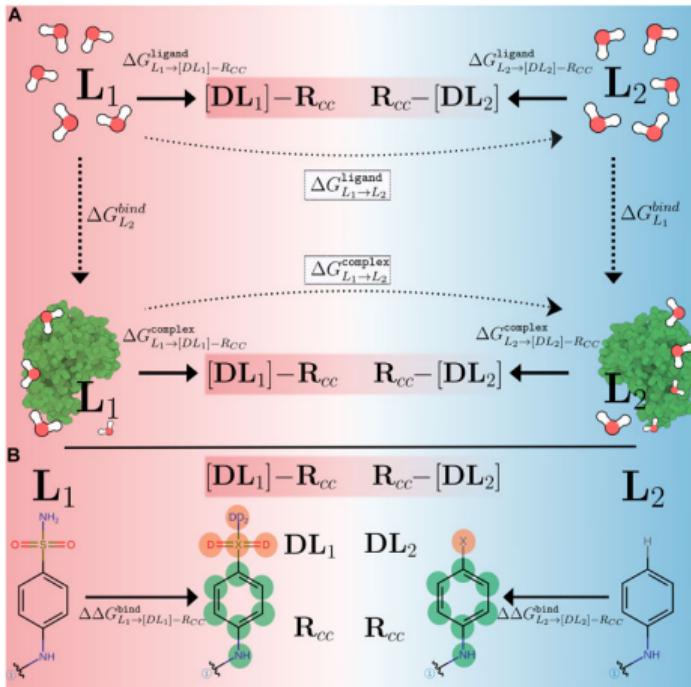


³

Sakuraba et al. "Free-Energy Calculation of Ribonucleic Inosines and Its Application to Nearest-Neighbor Parameters." Journal of Chemical Theory and Computation 16.9 (2020): 5923-5935.

Transformato⁴

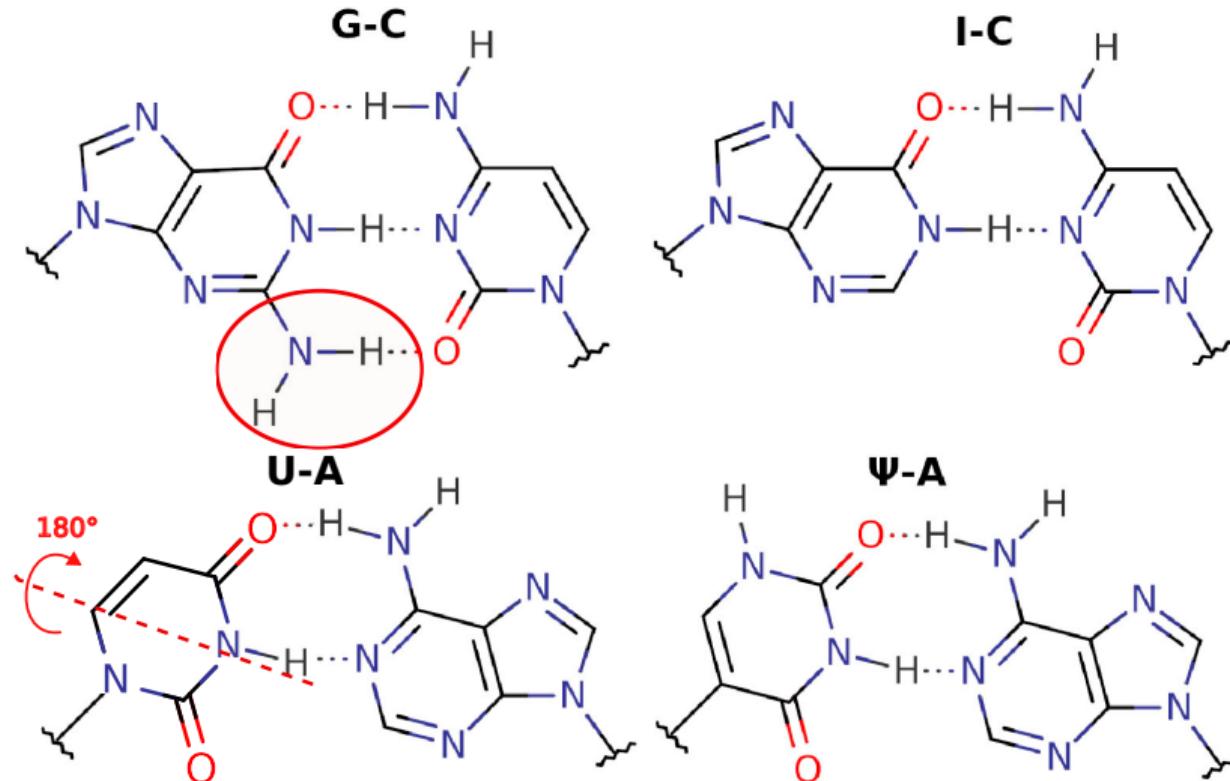
- OpenMM as underlying MD program
- Input files created by CHARMM-GUI
- Identify the maximum common substructure
- Construction of an alchemical path
 - Atoms not belonging to the common substructure are mutated to dummy atoms
 - Over a certain amount of steps the interactions of the dummy atoms are turned off
- Free energy difference to the common core calculated with Multistate Bennett Acceptance Ratio (MBAR)



⁴

Karwounopoulos et al. "Relative binding free energy calculations with transformato: A molecular dynamics engine-independent tool." Frontiers in Molecular Biosciences 9 (2022)

G-C to IC and U-A to Ψ -A at 1M NaCl



$\Delta\Delta G$ MD VS. RECCES

Duplex	MD	std	RECCES	Turner
5'CA I CG 3'GUCGC	2.28	0.21	1.31	2.04
5'GCA I CGC 3'CGUCGCG	2.04	0.27	1.79	2.04
5'GAAGA I GCAA 3'CUUCUCCGUU	1.88	0.44	2.05	1.73
5'GAAGA I ICCAA 3'CUUCUCGGUU	2.03	0.49	1.80	2.04
5'GGGCUCA I GCAGCCG 3'CCCGAGUCCGUCGGC	2.68	0.18		1.54
5'CGGPUCG 3'GCCAAGC	0.87	0.92	0.89	-2.86
5'GGGCAGGPUCGCGCC 3'CCCGUCCAAGCGCGG	-0.28	2.36		-2.86

Conclusion and Outlooks

- Rosetta-RECCES
 - Good predictions for WC, $G \bullet U$, and $I \bullet C$ base pairs
 - Systematic deviation for other modifications ($\Psi \bullet A$, $m^6 A \bullet U$)
- Molecular Dynamics
 - Framework still need some adaptations
 - Promising results with $I \bullet C$ base pairs
 - Use of different salt concentrations
- A combination of methods involving MD, RECCES, and experiments may represent the most suitable way
- Starting computations for a full set of stack pairs containing modifications
- Include larger loops

Acknowledgments

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The whole TBI team

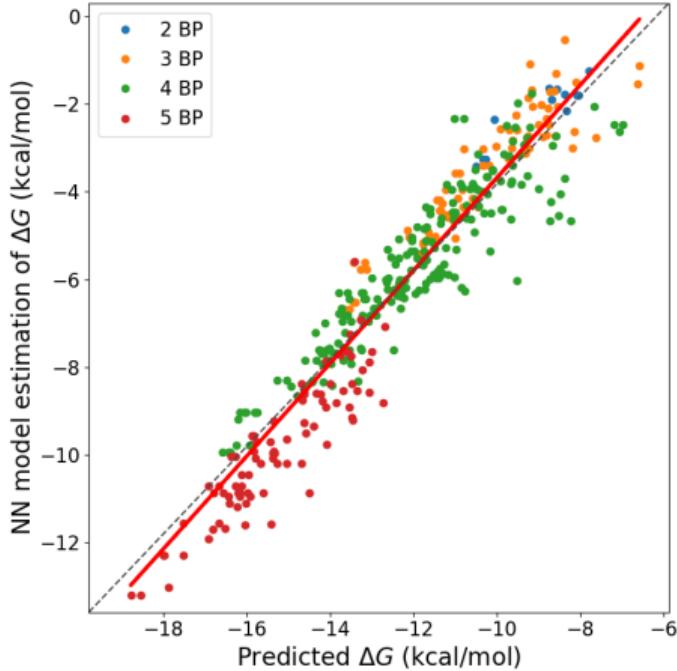
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Comparison of free energy estimations

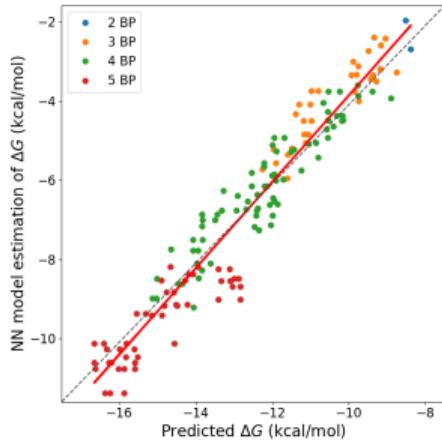
Canonical base pairs



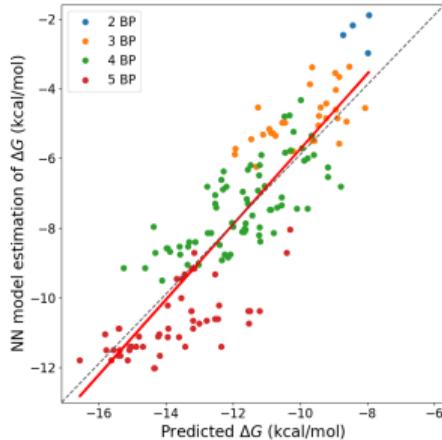
- All simulated duplexes from both methods (2 to 5 BP)
- Average offset = 6.18
- linear fit: $y = 1.06 \cdot x + 6.92$
- Corr. coeff. = 0.95
- High correlation
- Slope of the linear fit near 1
- Both models are separated by \approx only a constant

⇒ This good concordance is reflected in the accurate parameter estimations for canonical base pairs

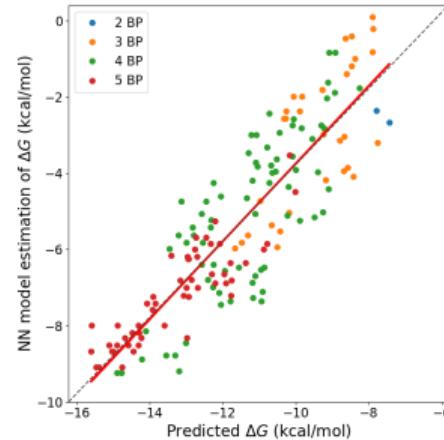
Comparison of free energy estimations



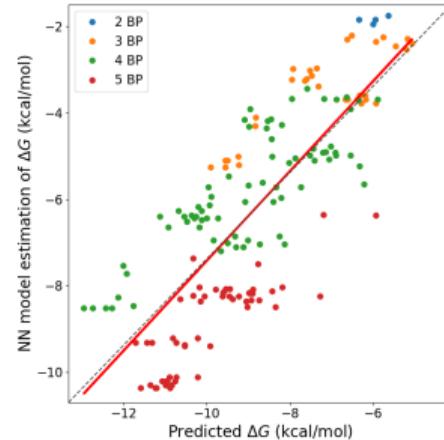
$I \bullet C$
Average offset = 5.90
linear fit:
 $y = 1.09 \cdot x + 7.04$
corr. coeff. = 0.96



$\Psi \bullet A$
Average offset = 4.11
linear fit:
 $y = 1.08 \cdot x + 5.05$
corr. coeff. = 0.87



$I \bullet U$
Average offset = 6.59
linear fit:
 $y = 1.12 \cdot x + 7.97$
corr. coeff. = 0.87



$m^6A \bullet U$
Average offset = 2.62
linear fit:
 $y = 1.04 \cdot x + 2.97$
corr. coeff. = 0.80

High Resolution Melting (HRM) experiments⁵

- Measures fluorescence as a function of temperature
- Derivation of thermodynamic parameters
- Good agreement with UV-melting if $T_m > 40^\circ C$

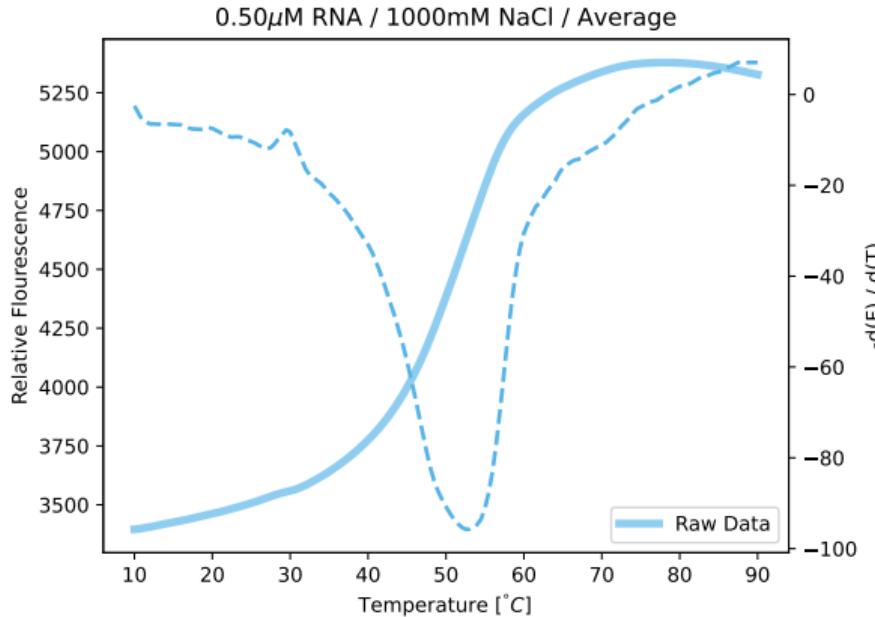
Is this a suitable alternative to UV-melting?

- Measurements for $I - C$ pairs in Jantsch Lab:
 - Oligos of 10 base pairs
 - NaCl concentrations $0mM$, $150mM$, $1000mM$
 - Oligo concentration $0.5\mu M$
 - Melting from $10 - 85^\circ C$ in $0.5^\circ C$ steps
 - 3 replicates

⁵

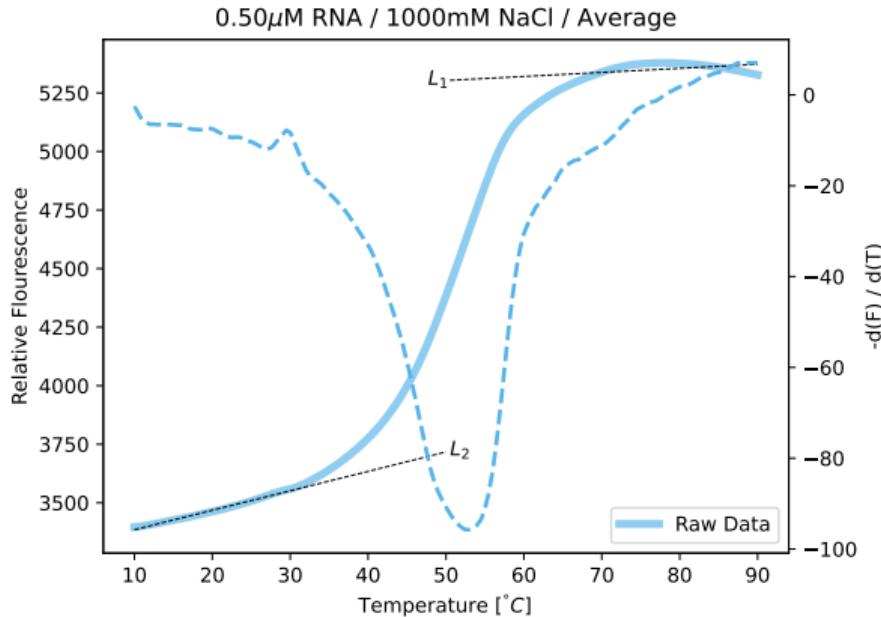
Wang et al. 2016, "Assessment for Melting Temperature Measurement of Nucleic Acid by HRM", Journal of Analytical Methods in Chemistry

Melting Data Analysis



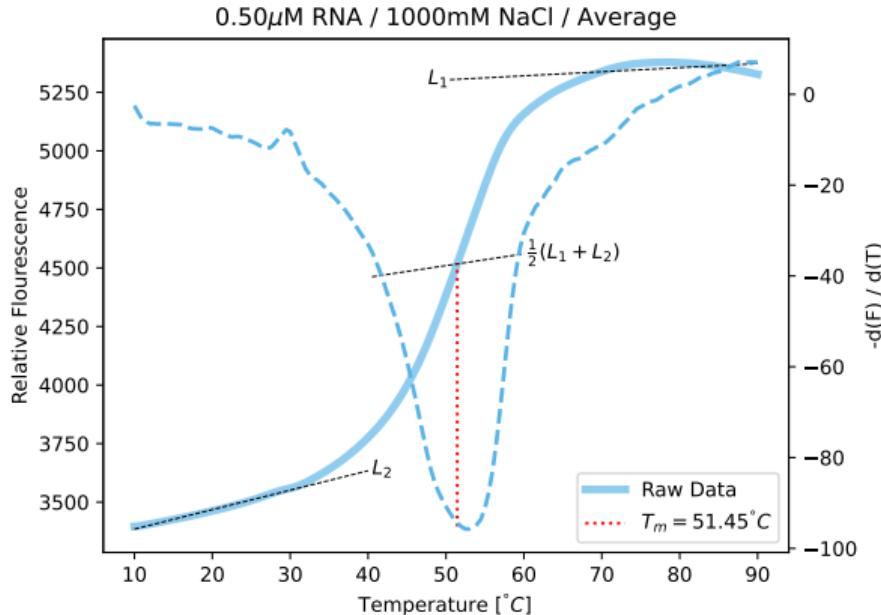
- $F(T)$ is related to the fraction of duplexes $\theta(T)$
- $A + B \rightleftharpoons AB$ with $2[AB] + [A] + [B] = c_o$
- $K_a = \frac{[AB]}{[A][B]} = \frac{2\theta}{c_o(1-\theta)^2}$
- $\theta(T_m) = 0.5$

Melting Data Analysis



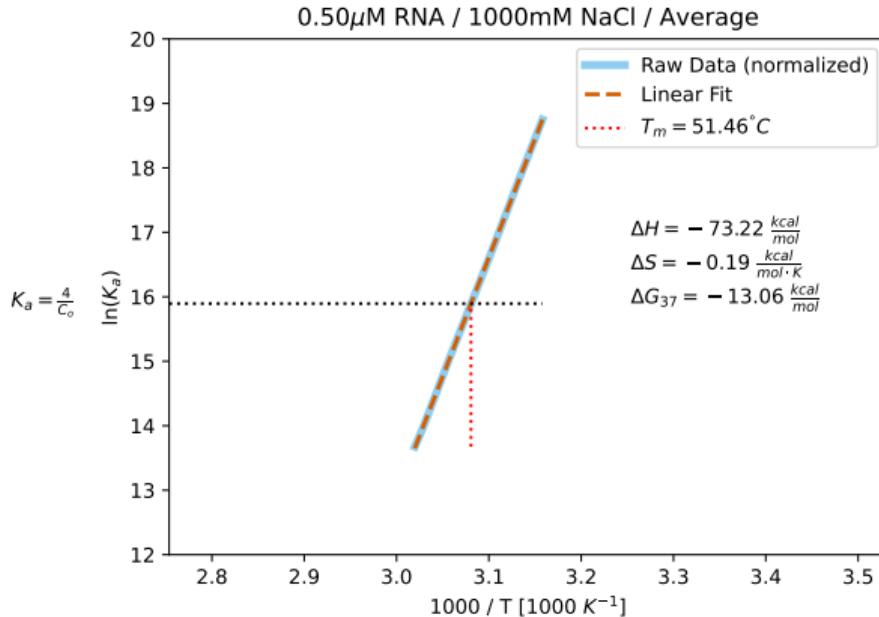
- Use $F(T)$ at low and high temperatures to estimate L_1 and L_2
- $F(T) = L_2(T) \cdot \theta(T) + L_1(T) \cdot (1 - \theta(T))$

Melting Data Analysis



What about ΔG_{37} , ΔH , and ΔS ?

Van't Hoff Analysis



$$\theta(T) = \frac{F(T) - L_1(T)}{L_2(T) - L_1(T)}$$

$$\frac{1}{T_m} = \frac{\Delta S}{\Delta H} - \frac{R \cdot \ln(4/c_o)}{\Delta H}$$

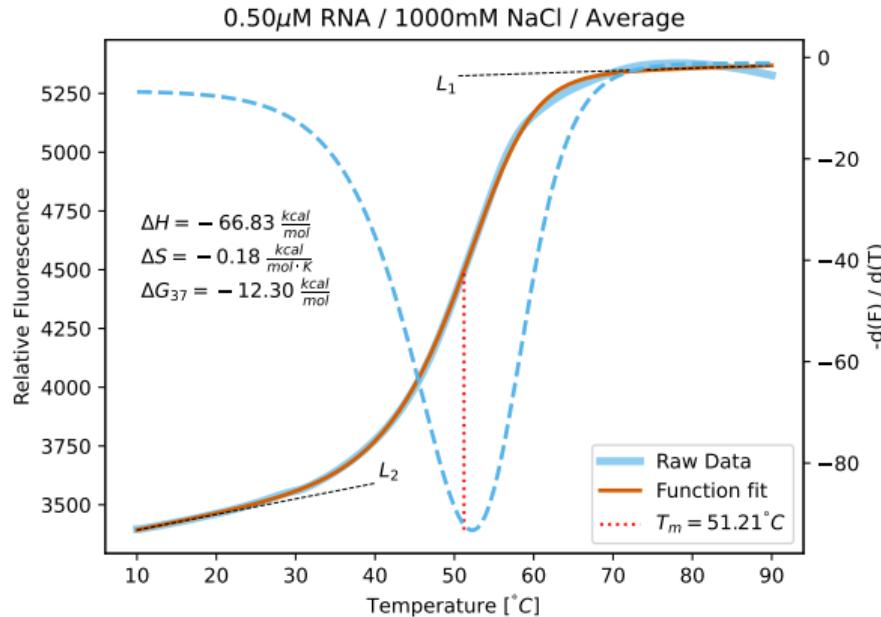
$$0.1 < \theta(T) < 0.9$$

$$\Delta H = -m \cdot R$$

$$K_a = \frac{[AB]}{[A][B]} = \frac{2\theta}{c_o(1-\theta)^2}$$

$$\Delta S = \frac{\Delta H}{T_m} + R \cdot \ln(4/c_o)$$

Curve fitting



- formulate $\theta(T)$ as function of T , ΔH and ΔS using
$$\Delta G = \Delta H - T\Delta S = -RT \ln K_a$$
 and $K_a = \frac{2\theta}{c_o(1-\theta)^2}$
- fit full function $F(T) = L_2(T) \cdot \theta(T) + L_1(T) \cdot (1 - \theta(T))$