

Artifacts of Sequences Under SELEX Selection Constraints

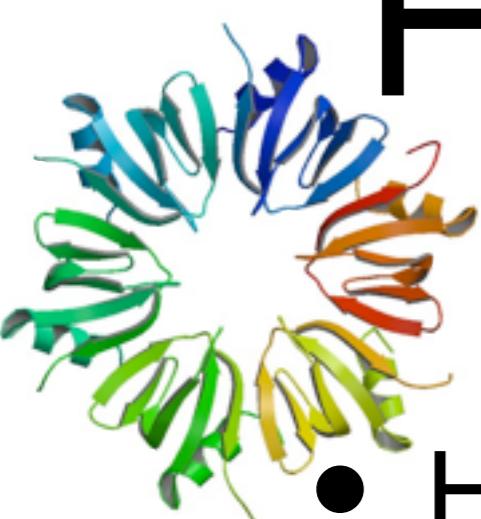
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Christina Lorenz and Renée Schroeder
Bled TBI Winterseminar
20 February 2009

Strategies for Detecting Novel ncRNAs

- *in silico*
 - Requires significant homology/structure
- *in culture/vivo* (RNomics)
 - Requires transcription conditions
- *in vitro* (Genomic SELEX)
 - Independent of expression levels
 - May present biases

Evaluation of Genomic SELEX Screen

1. Genomic SELEX for Hfq-binding RNAs
2. *in silico* analysis of sequenced cDNAs
3. Neutral SELEX as control for SELEX
4. 454 sequencing of Neutral and Genomic SELEX cDNAs
5. *in silico* analysis of SELEX pools

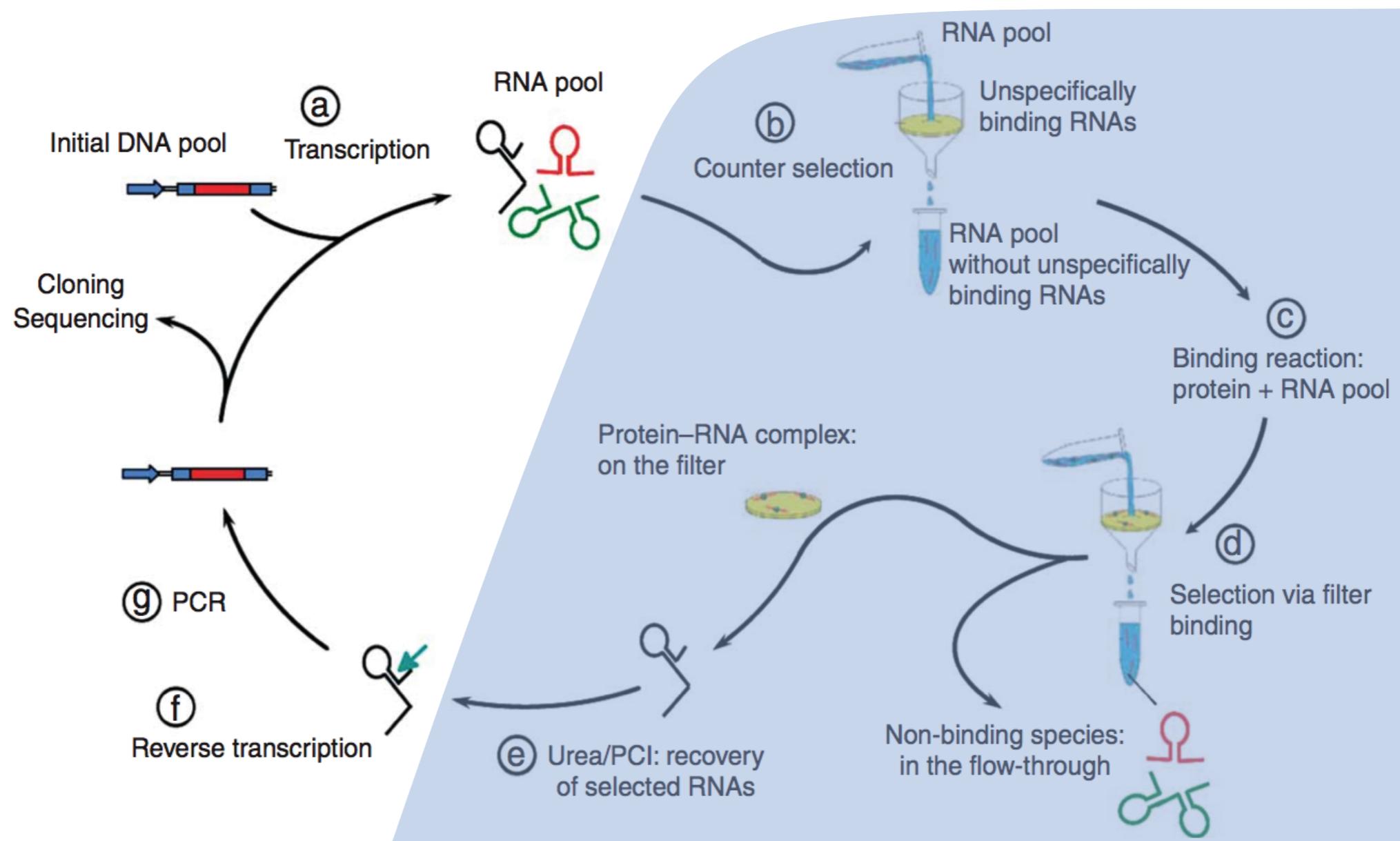


Hfq is a RNA-binding microbial protein

- Highly abundant, pleiotropic regulator involved in ncRNA/mRNA interactions
 - OxyS RNA O₂-stress regulation
 - Spot42 RNA kinase down-regulation
- Interested in a broader view of Hfq targets
- Develop a method irrespective of expression level: Genomic SELEX w/ *E.coli*

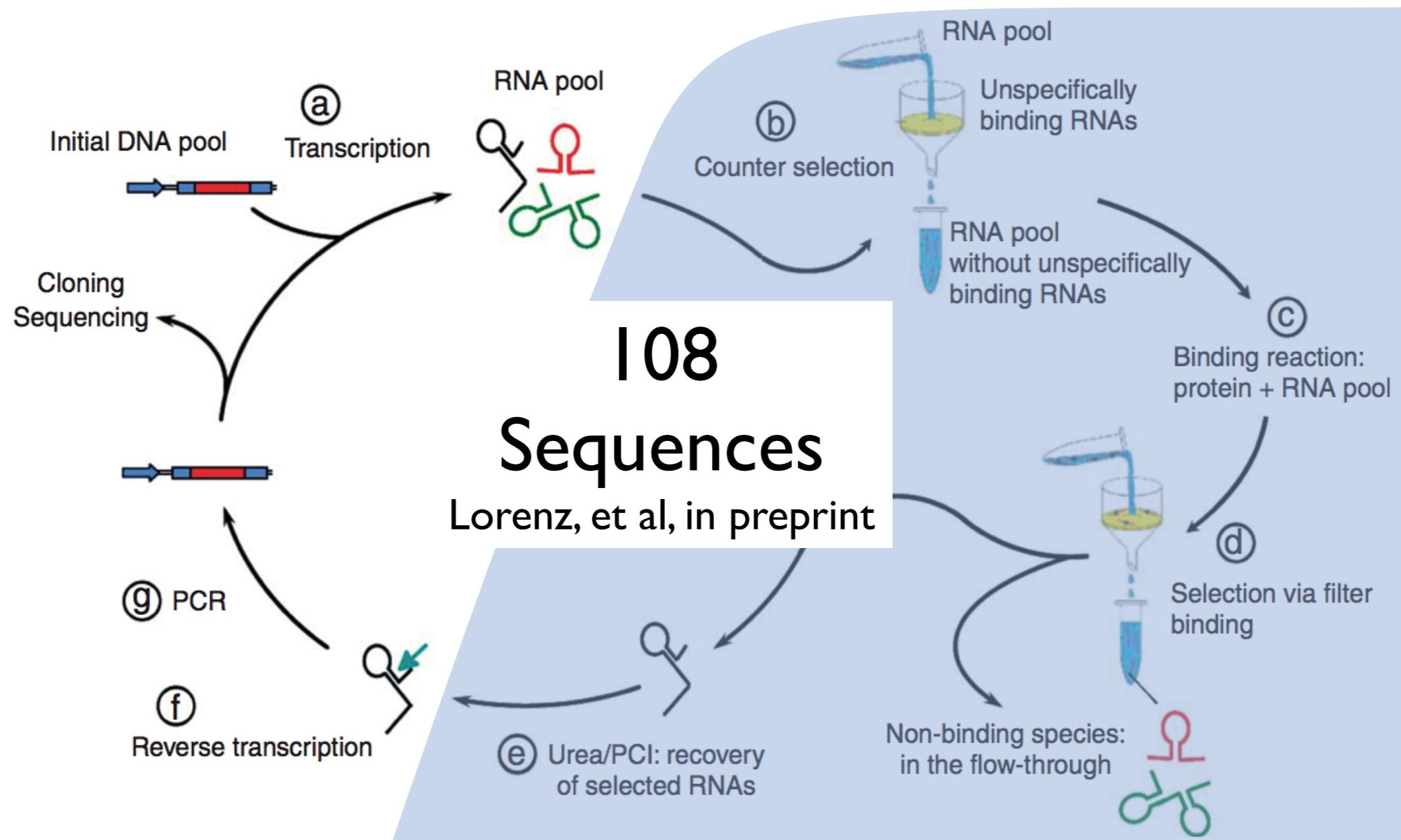
Lorenz, et al, in preprint

I. Genomic SELEX for Hfq-binding RNAs



Lorenz et al, 2006

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Lorenz et al, 2006

2. *in silico* analysis of sequenced cDNAs

- Measure structural stability: MFE Z-score
- Compares predicted secondary structures of
 - same length
 - same nucleotide content

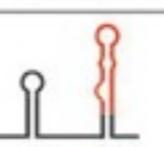
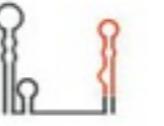
$$Z = \frac{E_x - \mu(\mathbf{E}_{x_{\text{shuff}}})}{\sigma(\mathbf{E}_{x_{\text{shuff}}})}$$

Le, et al 1989

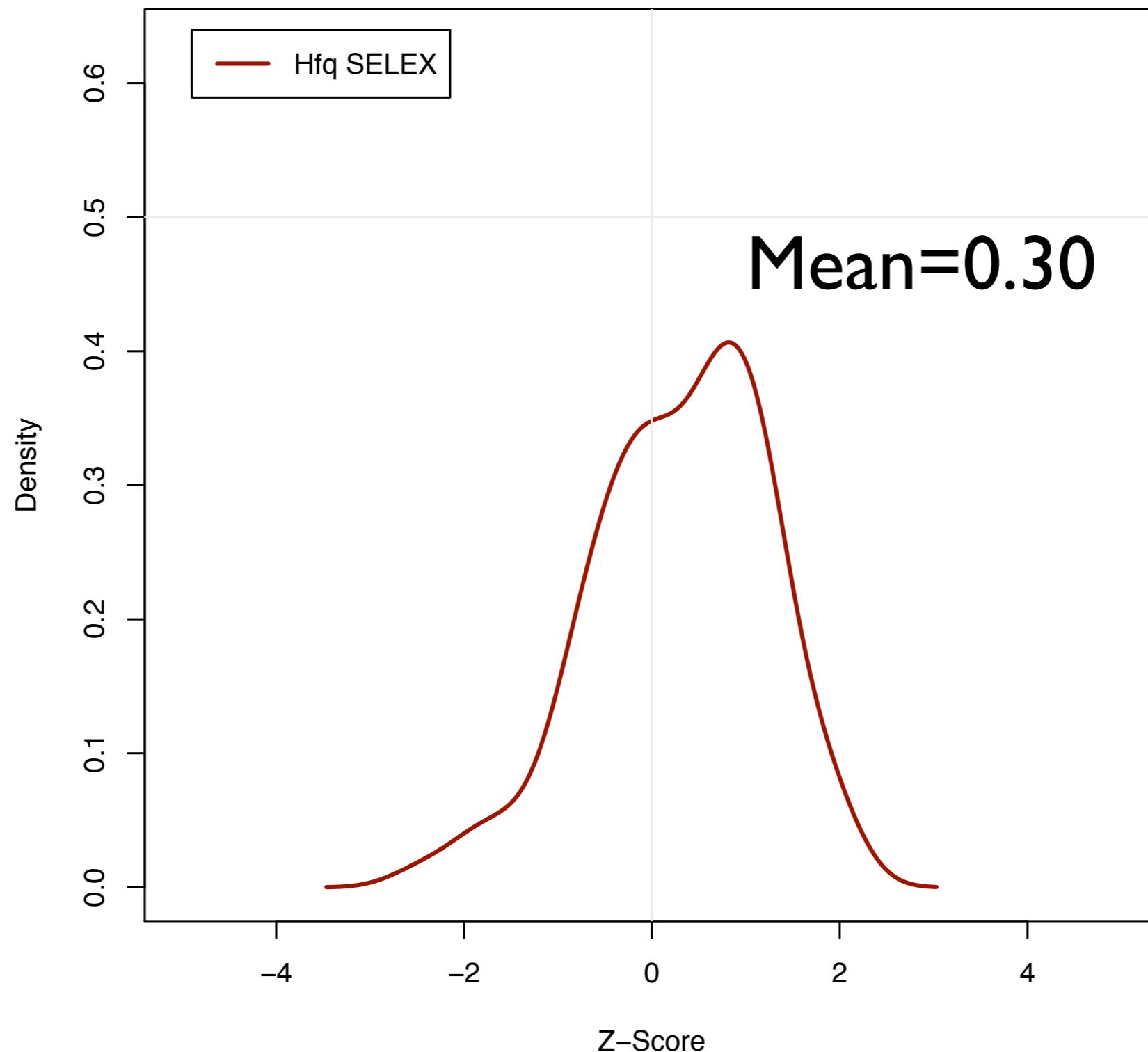
2. *in silico* analysis of sequenced cDNAs

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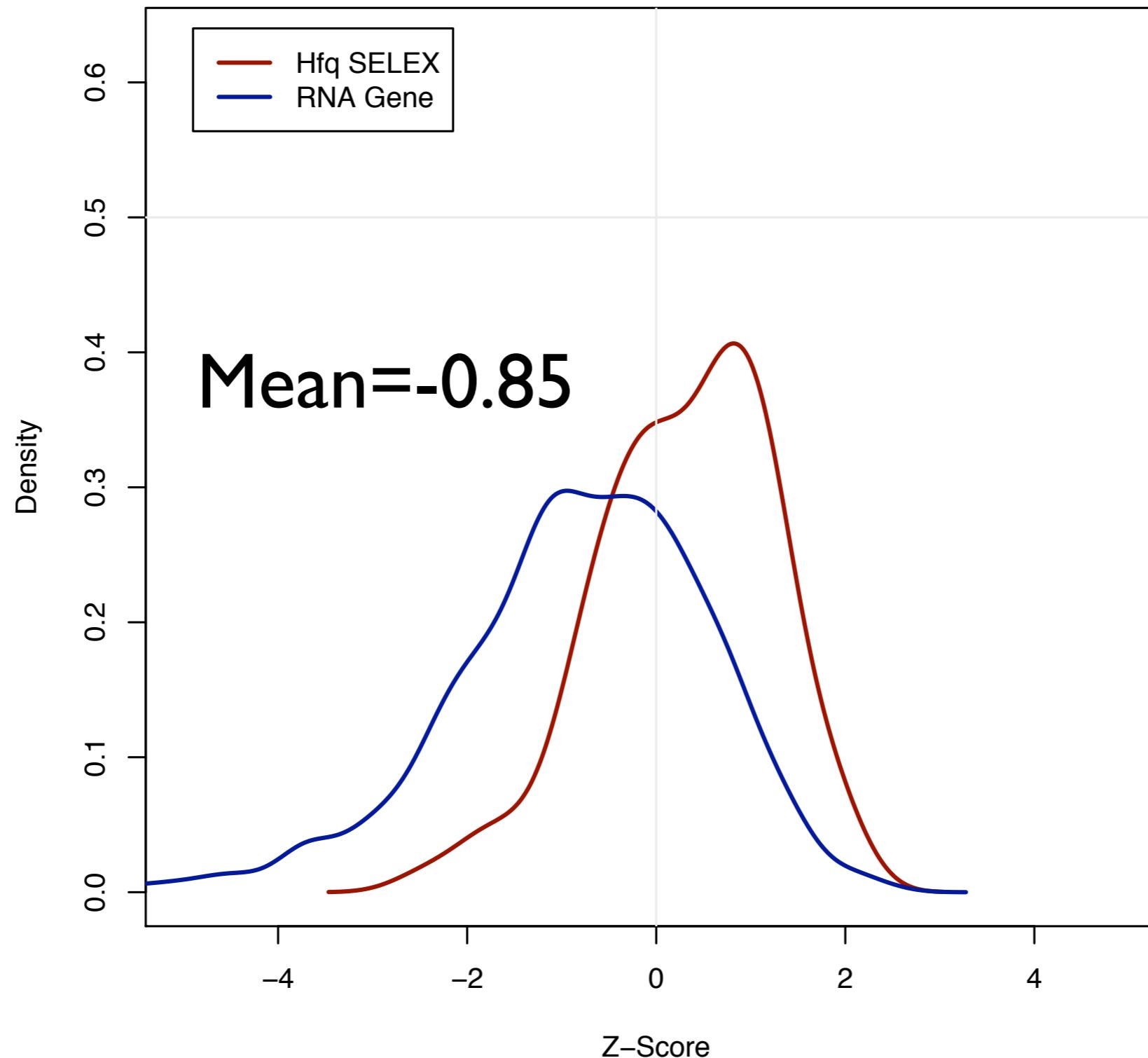
Z-scores of known binders

RNA	Secondary Structure	
6S A		-5.52420
DicF A		-1.60733
DsrA A		-2.09376
MicF A		-1.95915
OxyS A		-3.03948
RhyB A		0.40742
RprA A		-0.75068
Spot42		-0.95073

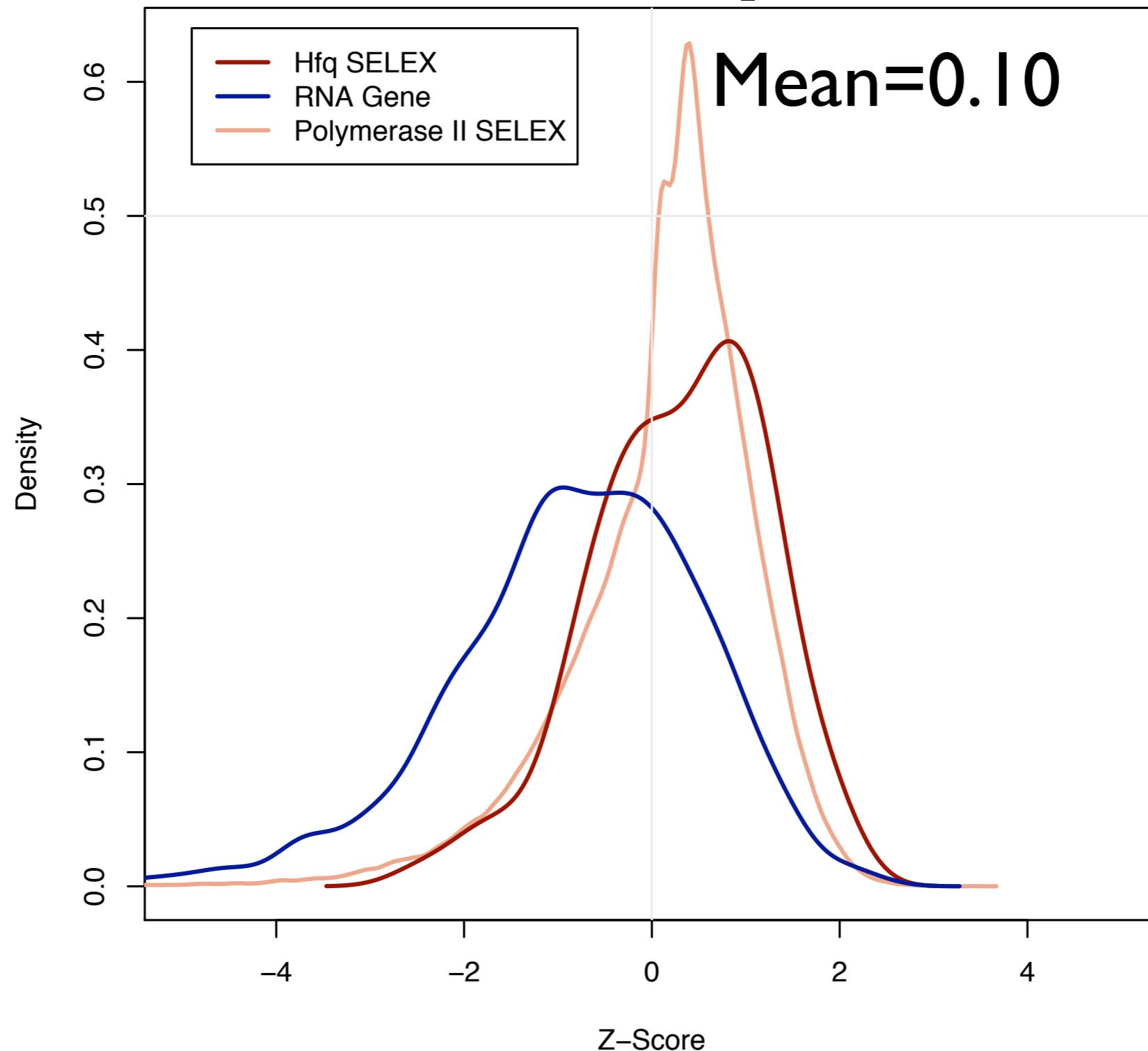
Hfq Sequence Stability



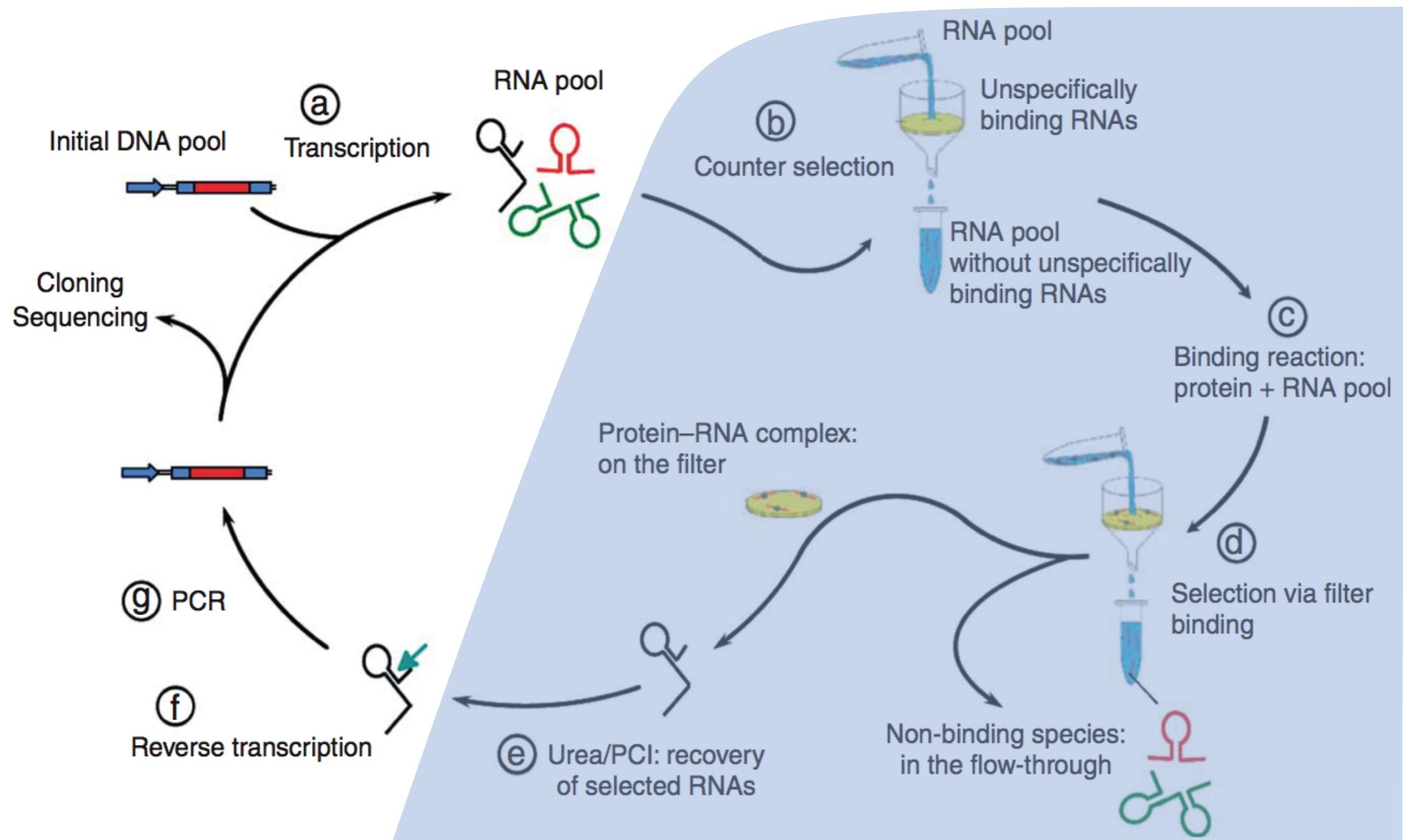
Does Hfq favor binding unstable RNAs?



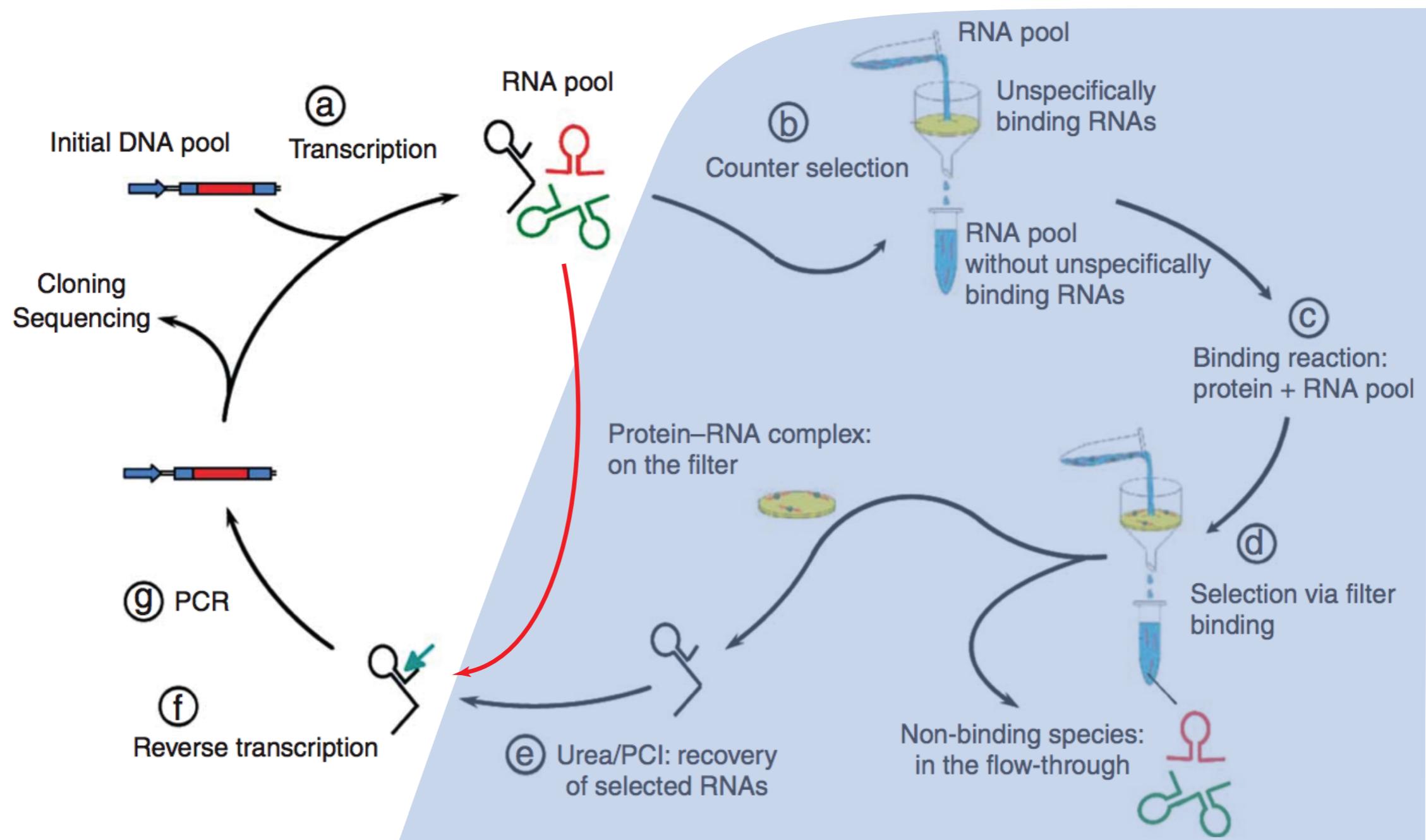
Does SELEX produce unstable sequences?



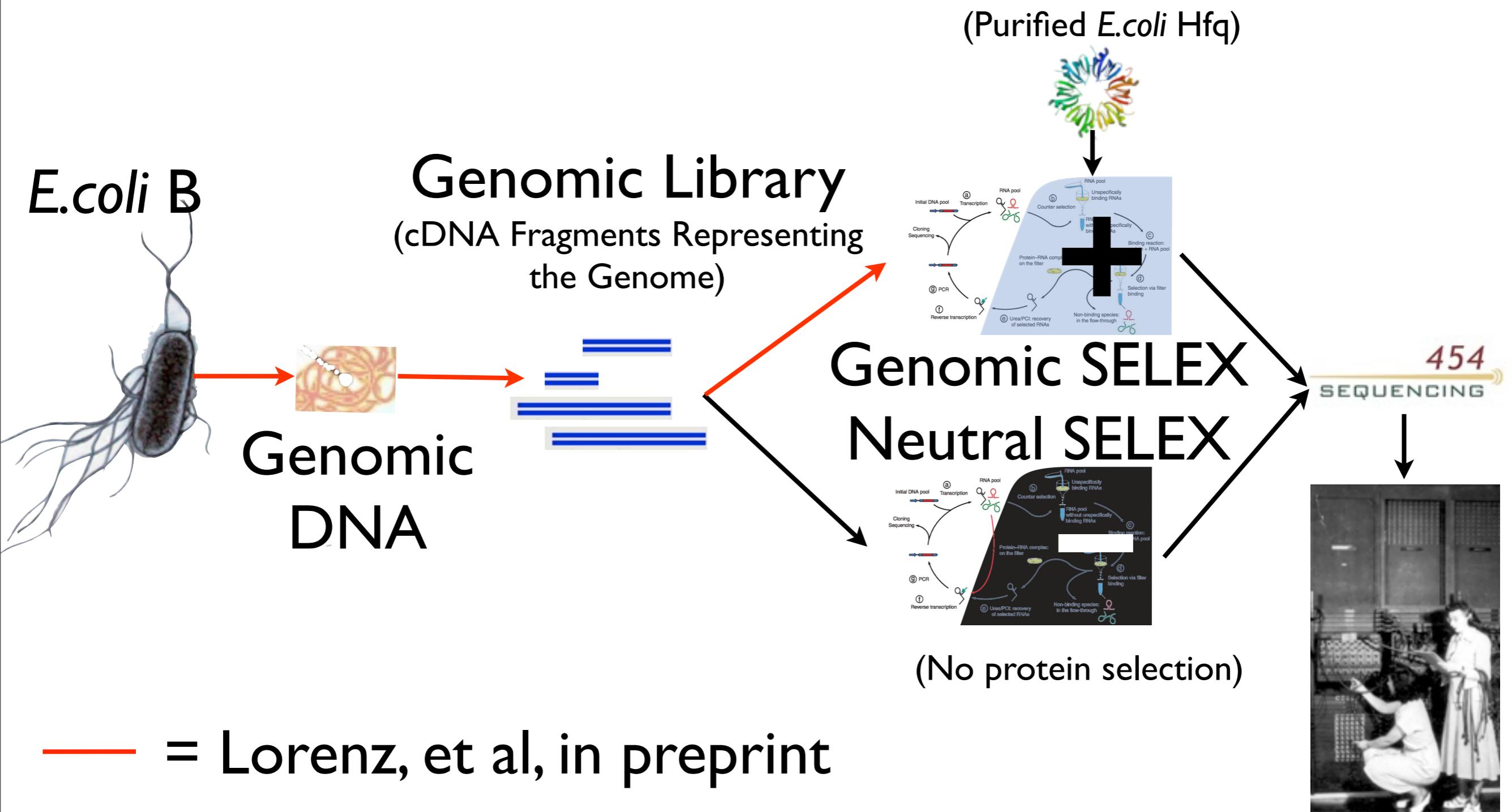
SELEX



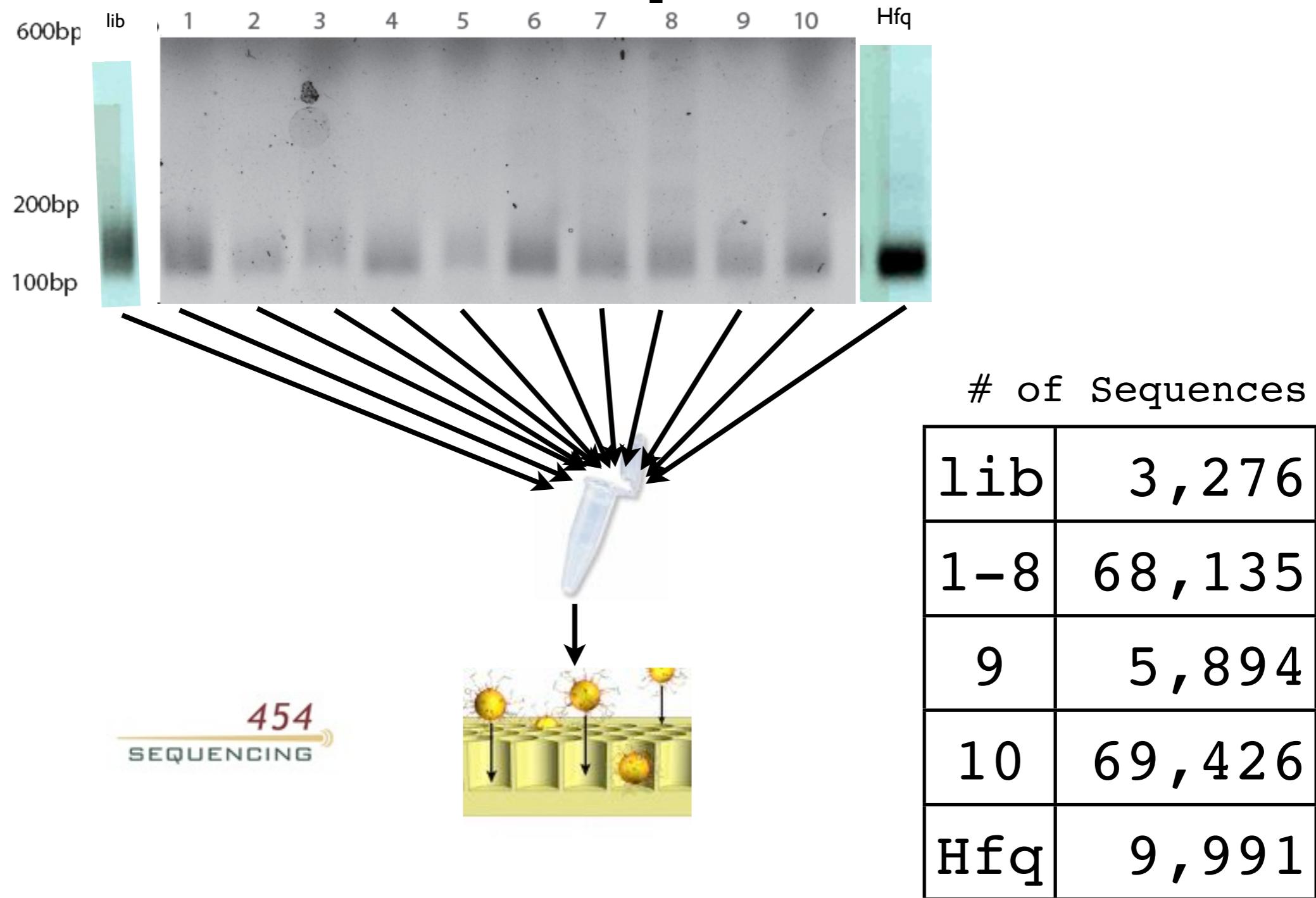
3. Neutral SELEX



Schematic of SELEX Experiments

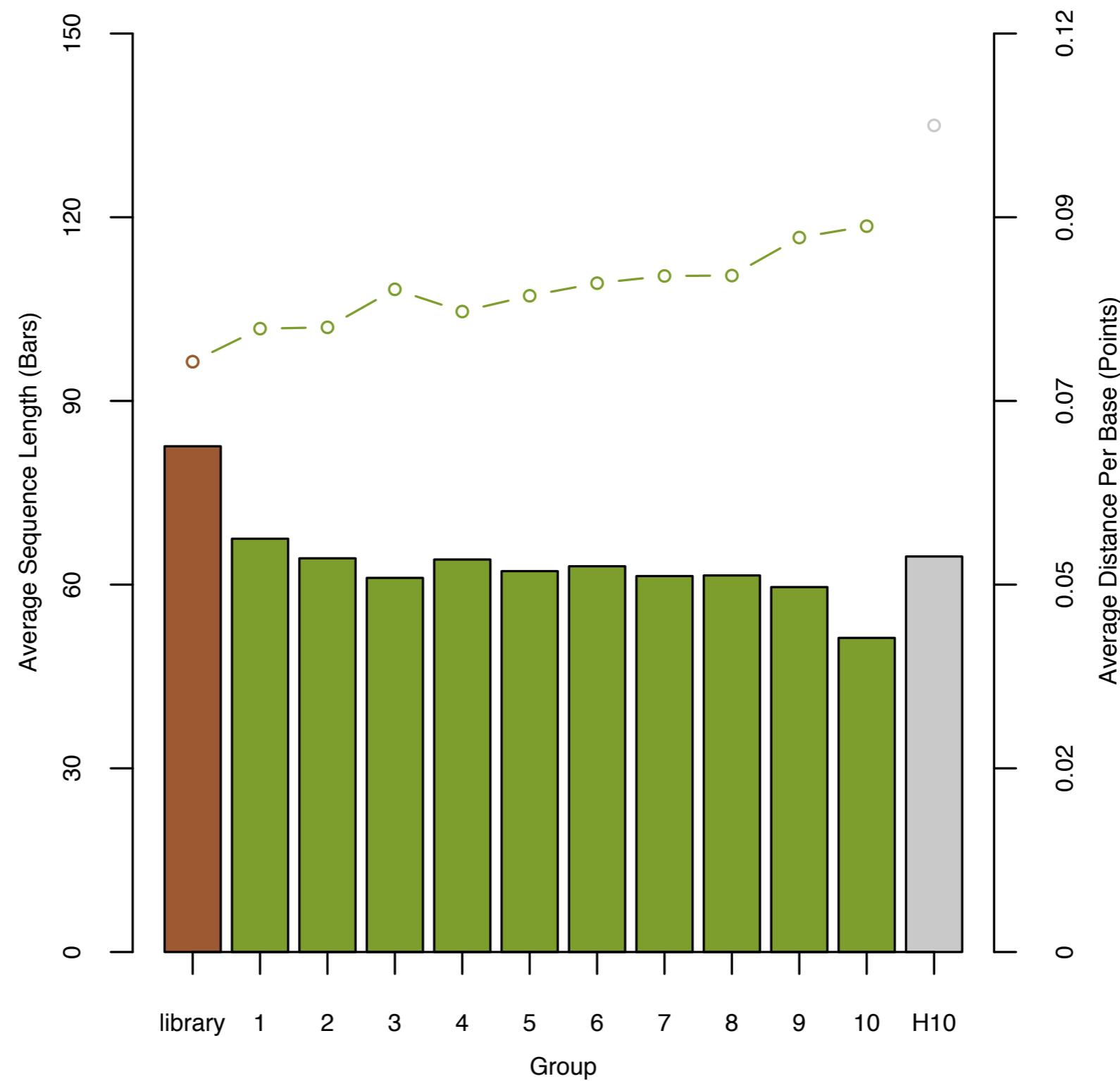


4. 454 Sequencing of SELEX pools



5. *In silico* analysis of SELEX pools

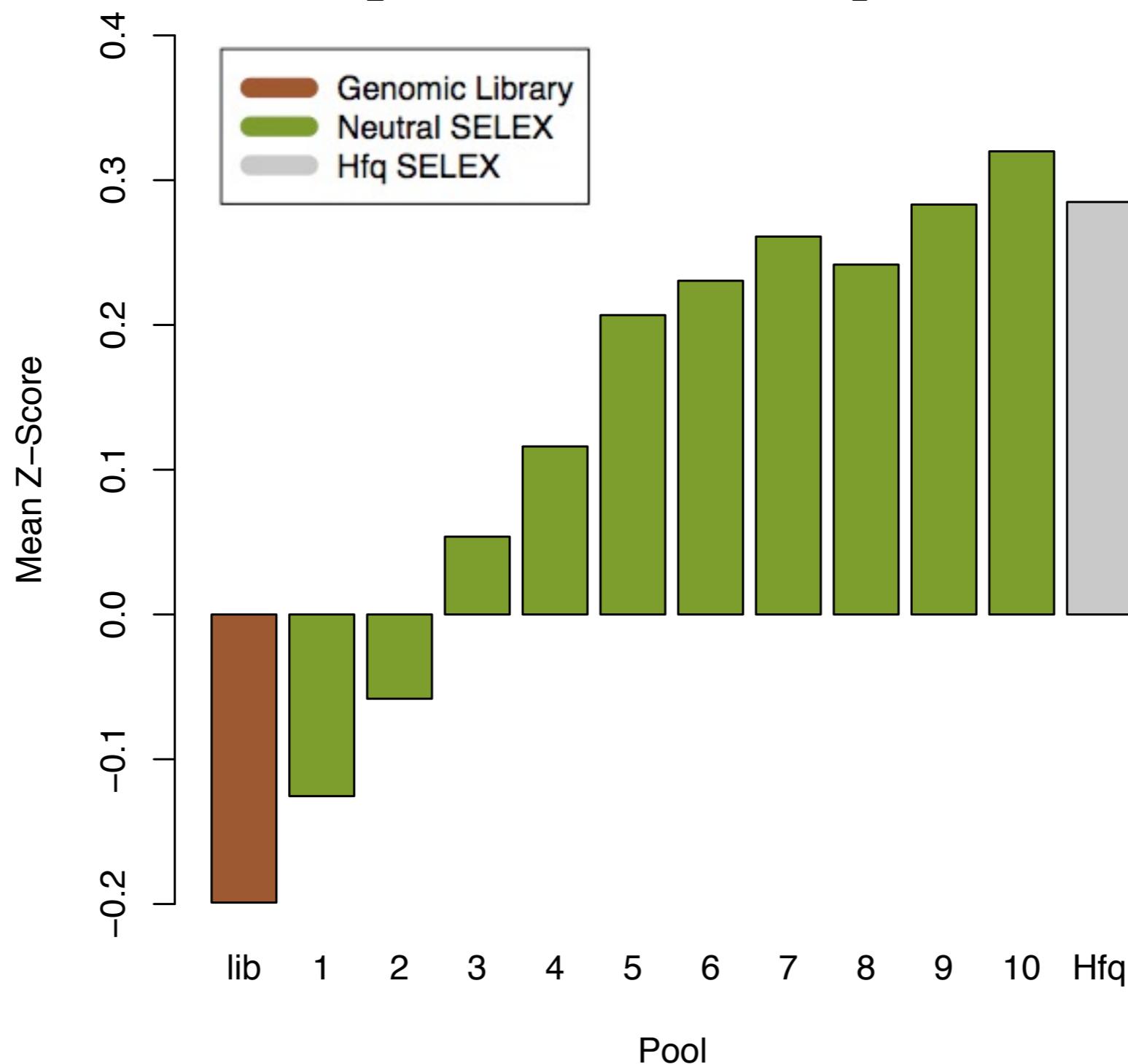
Lengths and Distances of Null SELEX Sequences



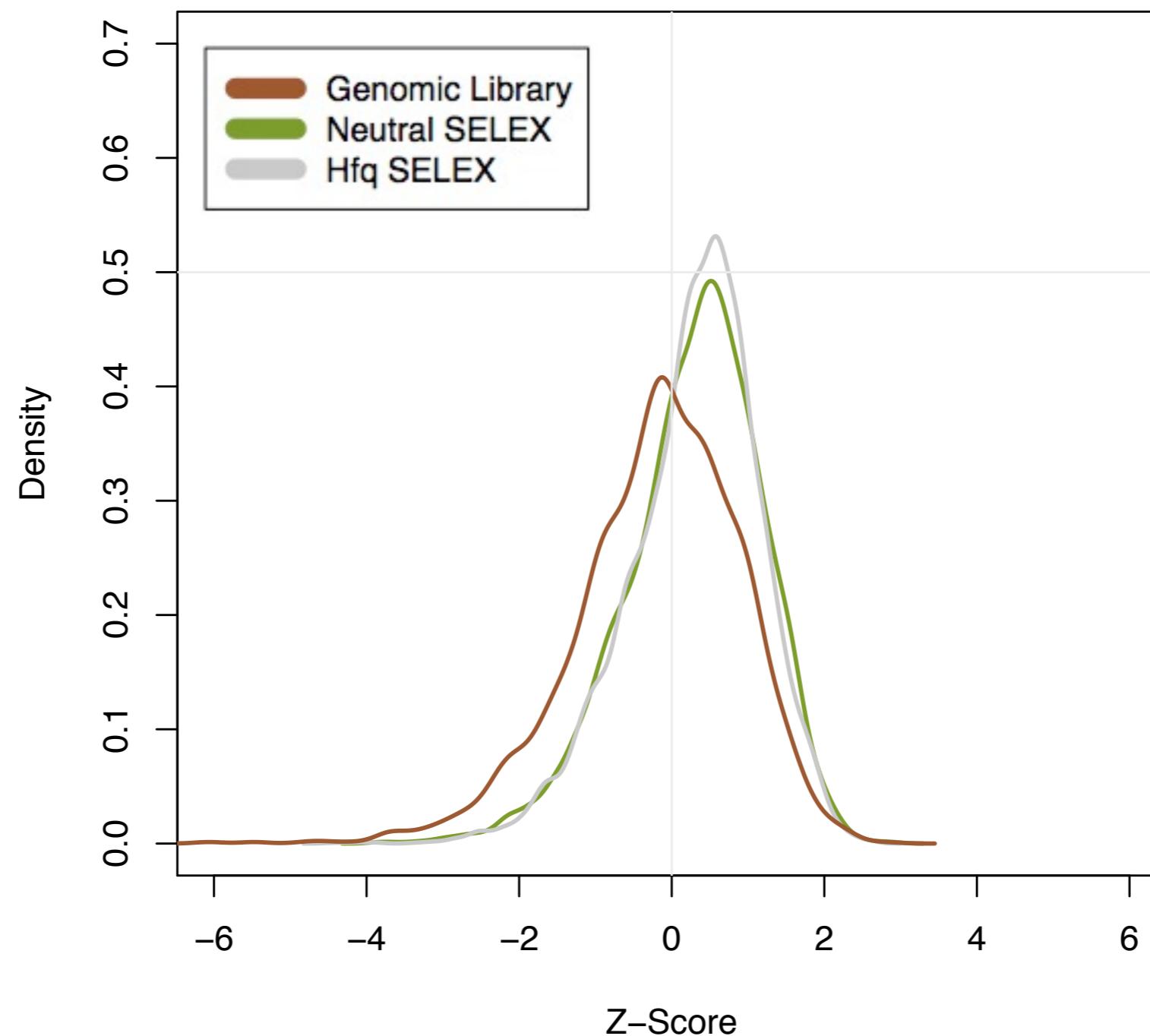
What is the effect of SELEX on the stability of sequences?

- Measure Z-score sequences in each pool
- Take the average of each pool
- Study progression of the pools' averages

SELEX decreases stability of sequences



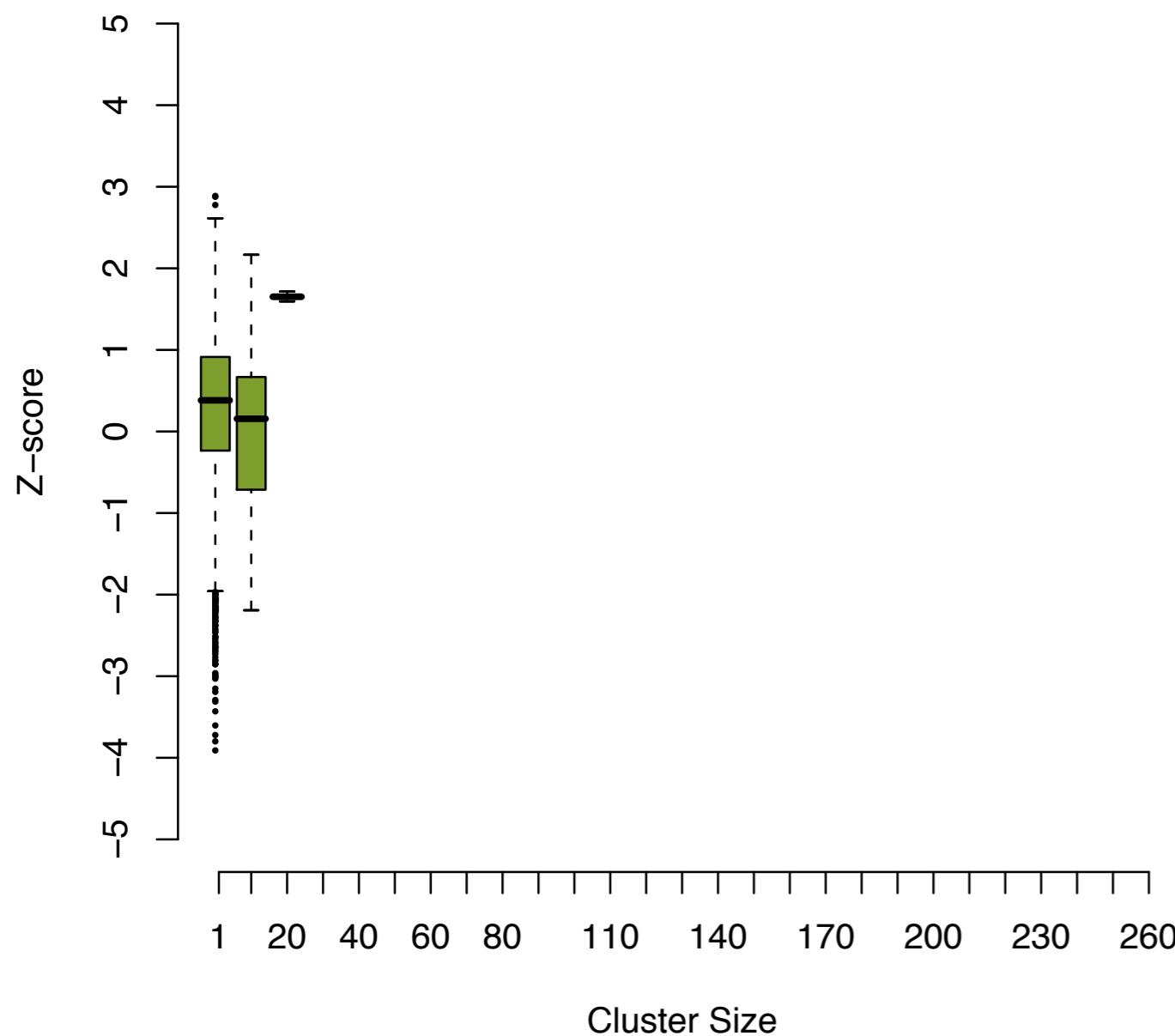
Hfq sequences are unstable from SELEX



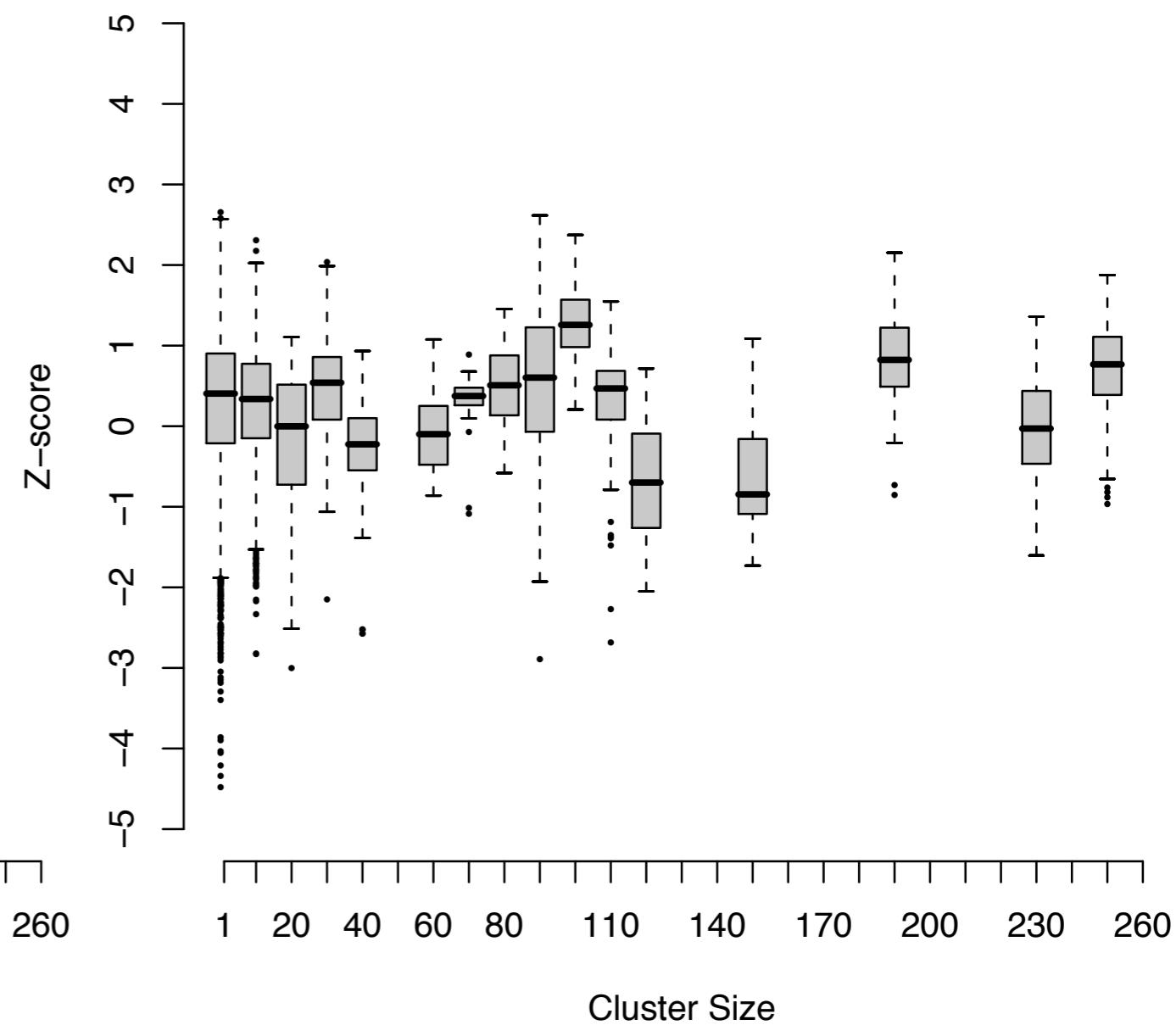
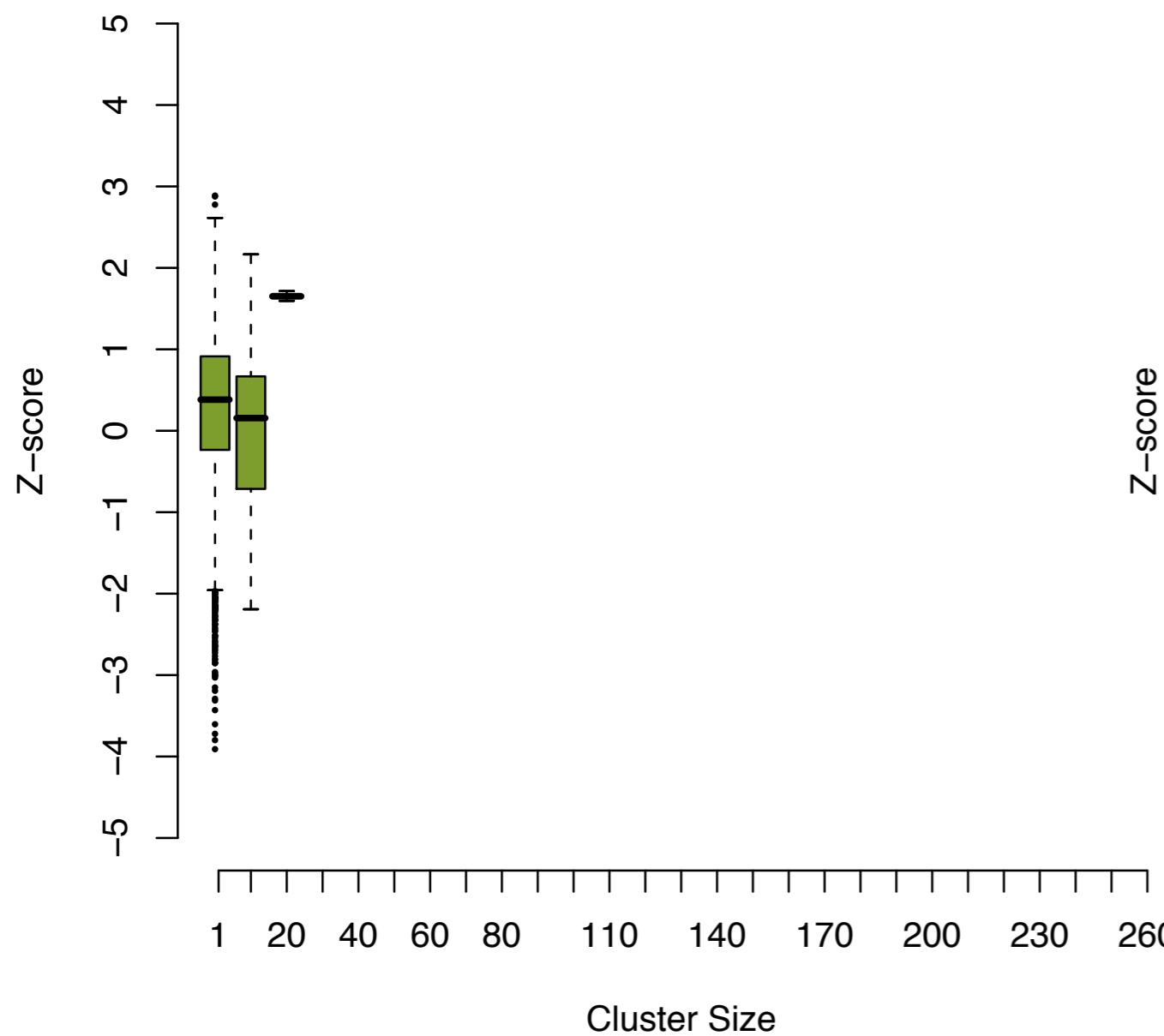
Must enriched sequences be unstable?

- Cluster sequences through pairwise alignments
 - If the sequences are 85% identical, group them in a cluster
 - Size of the cluster is a measure of the level of enrichment
- Plot the enrichment against Z-score

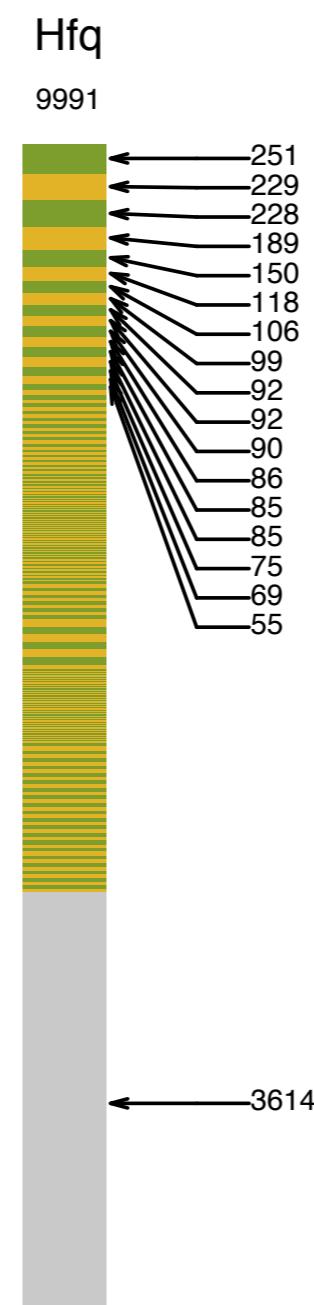
Enrichment does not imply instability



Enrichment does not imply instability

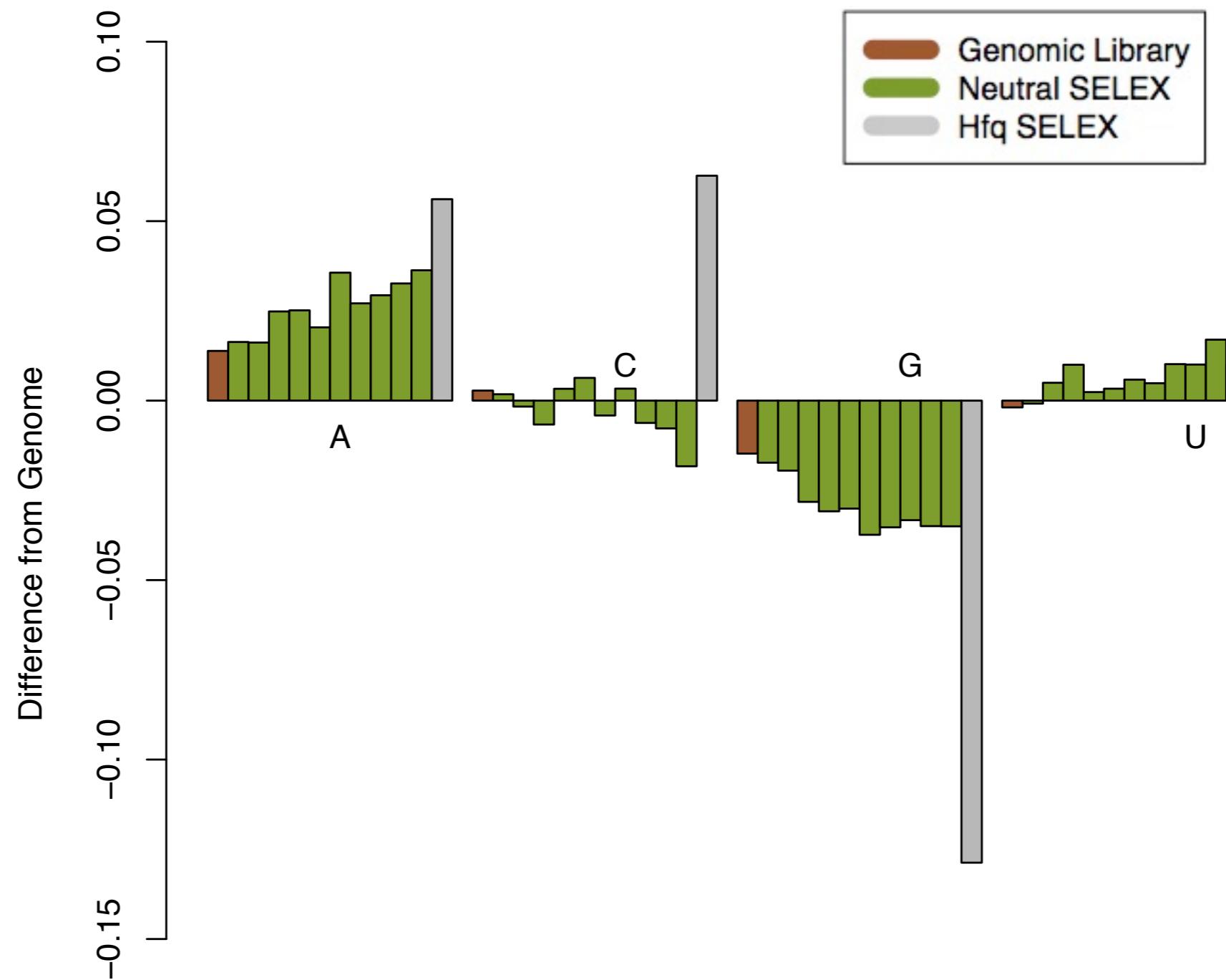


Enrichment is a Strong Signal



Enrichment is a Strong Signal

Nucleotide Content

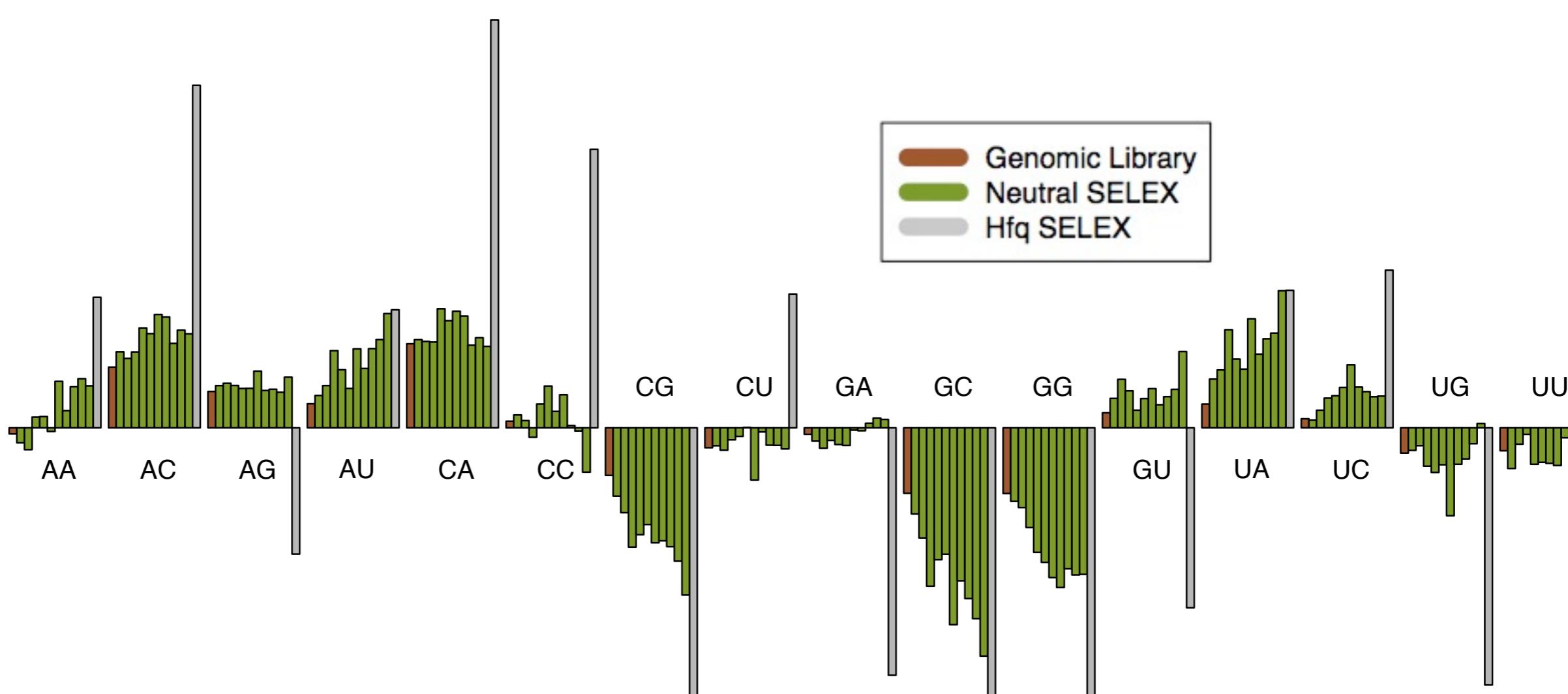


Dinucleotide Content

Difference from Genome

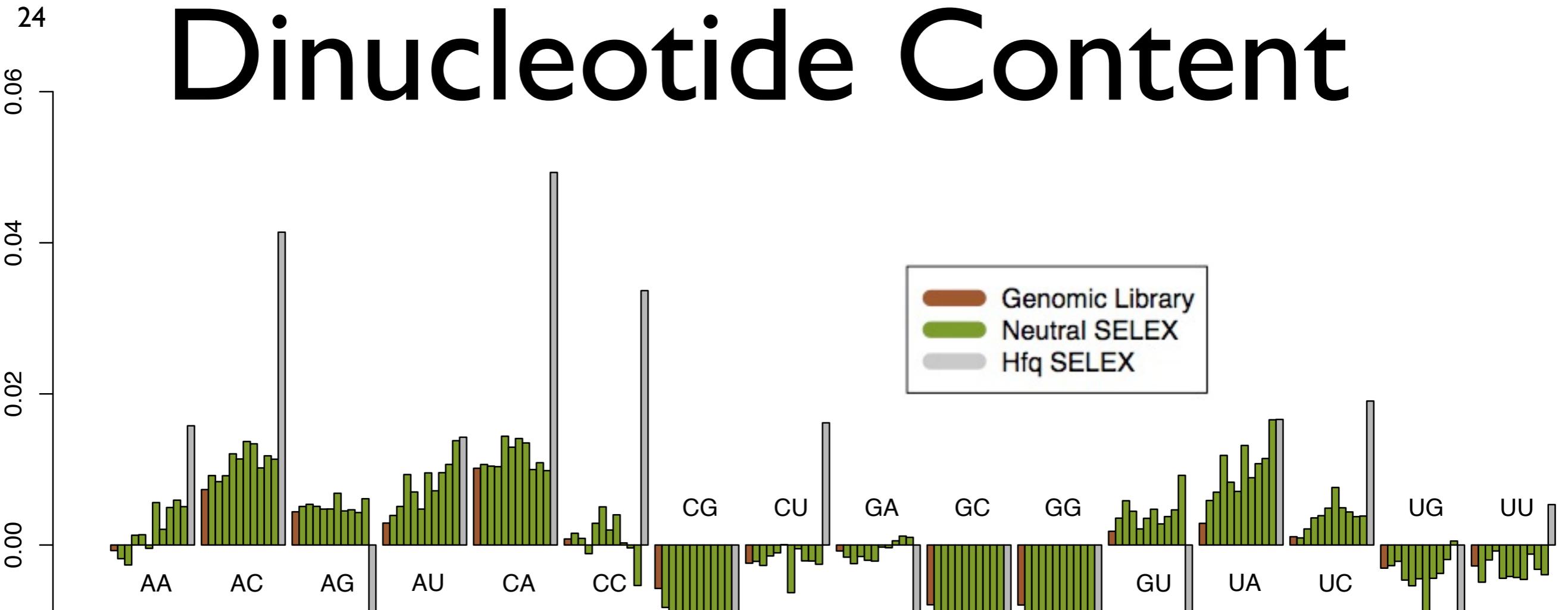
24
0.06
0.04
0.02
0.00
-0.02
-0.04
-0.06

- Genomic Library
- Neutral SELEX
- Hfq SELEX



Dinucleotide Content

Difference from Genome



Winning Sequences: (of 999 !)

1. CCCACG-TTAATCCAATAAGAACAGAACAGATTATCCGTTAACATCACTCCTTCAGTG (251x)
2. ACCA--ATAACTACAACATCAGCCCCAGTTCTTCTGCCAACTCATTGACTT (229x)
3. CCATAACA-ATAATCCATAGCTTCCCAGCCCACAAGGTACGAAACAAACCCACGAATATCCAGTTCATTTCAGCGT (228x)
4. CTACACCATTGAAAATAAGAACATTCTACTCACCCCTTGCACCTCACTCCTGCTTACCAACGTTTC (189x)
5. ACTTCATCACTAACAAACAATAATACGGGAGACATTGGTTCTT (150x)
6. AACAA-CAATAAA-GCGGCACACCCCTACCCAAAACGCTGCTCGCATCTCTTCTCTGATCTTCAAGC (118x)
7. AATTCCACCATTATATCCATCTCCTCCCCAACACTTCCATTATTATTAGCGTAAAGTTTCACCGTA (106x)
8. CAAAAACCTACAAGACCATAAGAACCGCCAGCCTTCGCCTTTTCAACACTCATTGCCACATTCCCTGTGTATA (99x)
9. GTCTTACGAACGGTATCCGCAGCCACTTCATCACTAACAAACAATAATACGGGAGACATTGGTTCTT (92x)
9. CGCCCAGACCAATAATAATAGTGACGGAACCAAGCACACCGCCCATTCCGGCGATCATCACTTACCCACTT (92x)
10. CATAAGAACAAACAAGACAATAATTGCCATTACAGCCACAATAGCCAGCGTGC (86x)

Conclusions

- Genomic SELEX presents artifacts
 - Unstable sequences are favored
 - Certain dinucleotides are enriched
- Genomic SELEX: an *in vitro* alternative
 - Can screen genome for binding motifs independent of expression level
 - Sequence enrichment is a strong signal
 - It is possible to enrich stable RNAs

Future Plans

- Can Neutral SELEX aid in the bioinformatic analyses of SELEX pools?
- How does SELEX compare to Co-IP
 - Polymerase II: Genomic SELEX and Co-IP
 - What bioinformatic analyses could lead up to the discovery of these RNAs *ab initio*?

Acknowledgements

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Marina, Andreas, Elli, Silke,
Katharina, Peter**



Renée Schroeder



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Laboratories**

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Christina Waldsich**

**Committee:
Eric Westhof
Ivo Hofacker**



Polishing Up

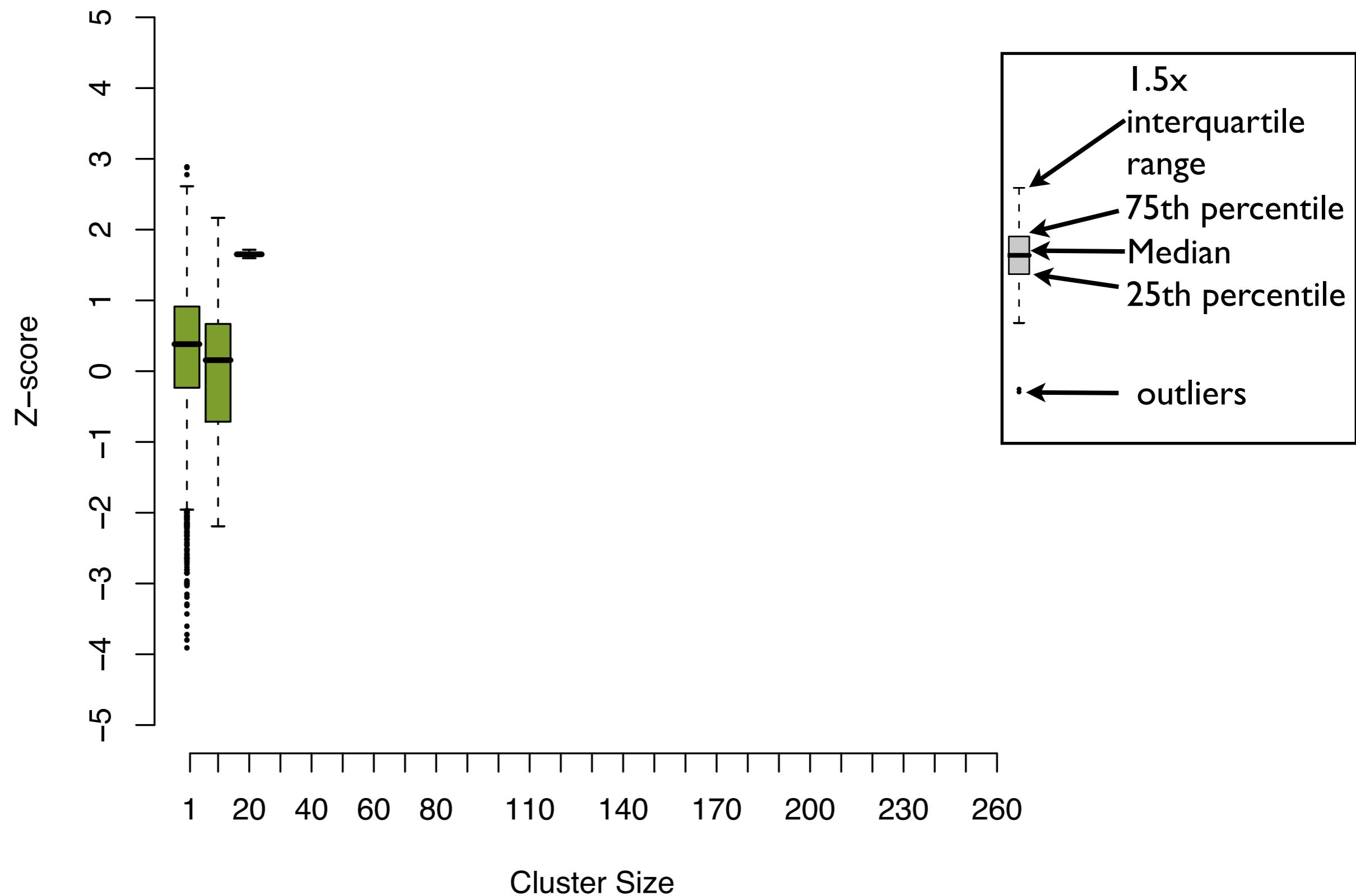
- To prove the worthiness of Genomic SELEX, we need the positive Hfq results
- We believe (are almost sure) the reverse transcriptase is at fault, but some biochemical grounding for the biases needs to be established. Could it be better?
- Previous work....

How is the nucleotide content affected by Hfq and Neutral selection?

- Count occurrence of each nucleotide of each pool
- Normalize to frequencies ($f_A + f_C + f_G + f_T = 1$)
- Compare SELEX pools' frequencies to genomic frequency

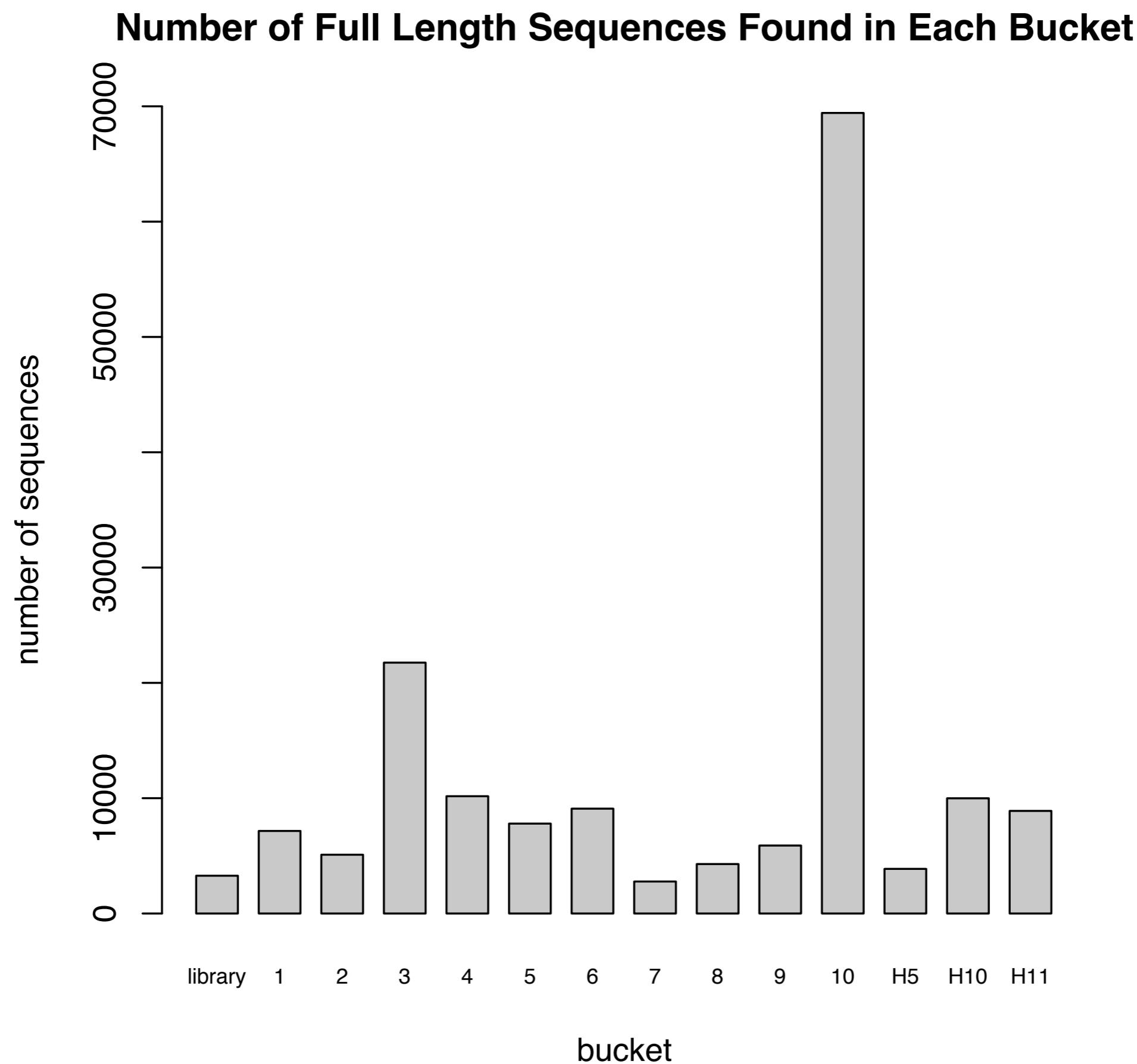
How stable are the Hfq-selected RNAs?

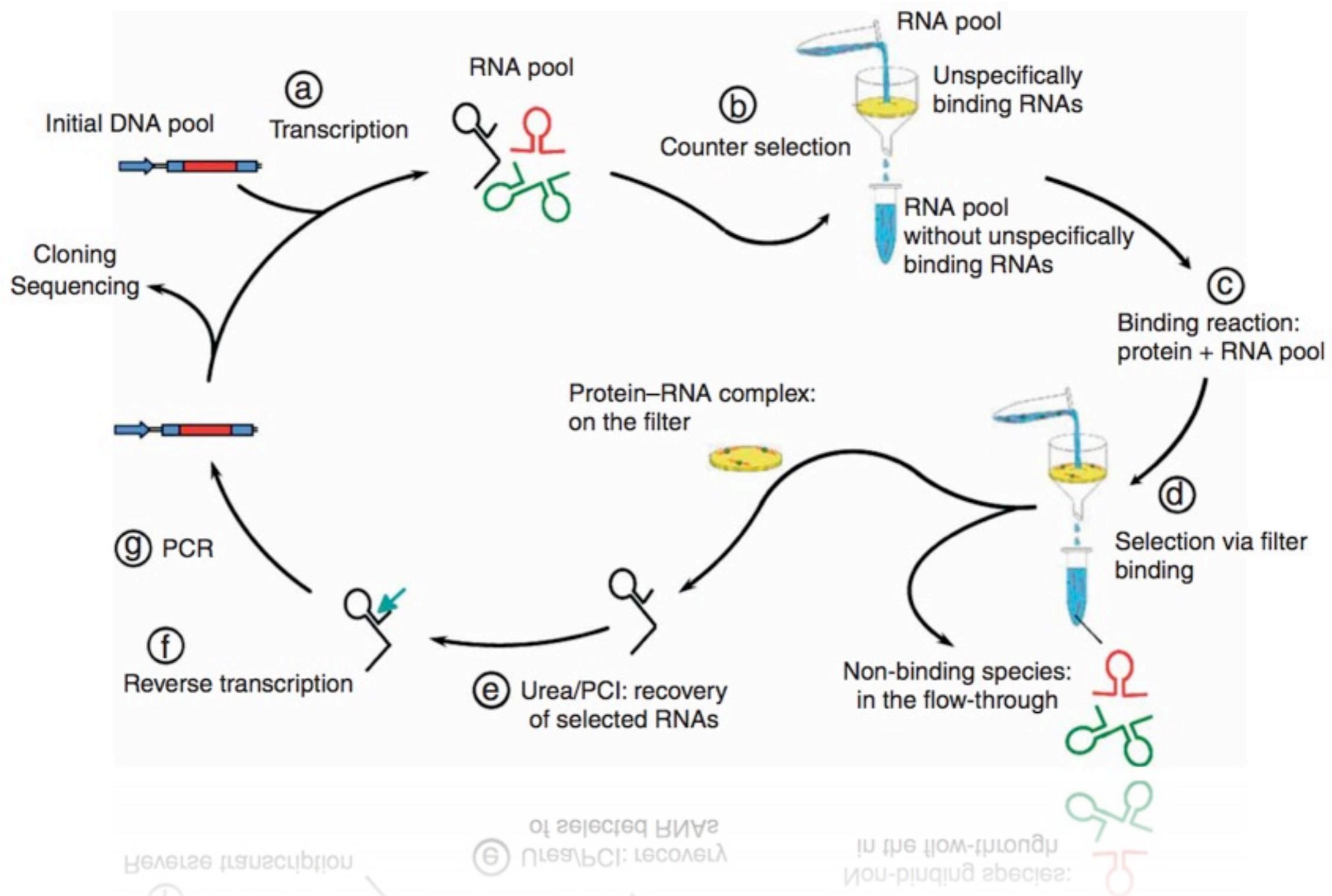
- Measure Z-score of each sequence in pool
- Look at the resulting distribution

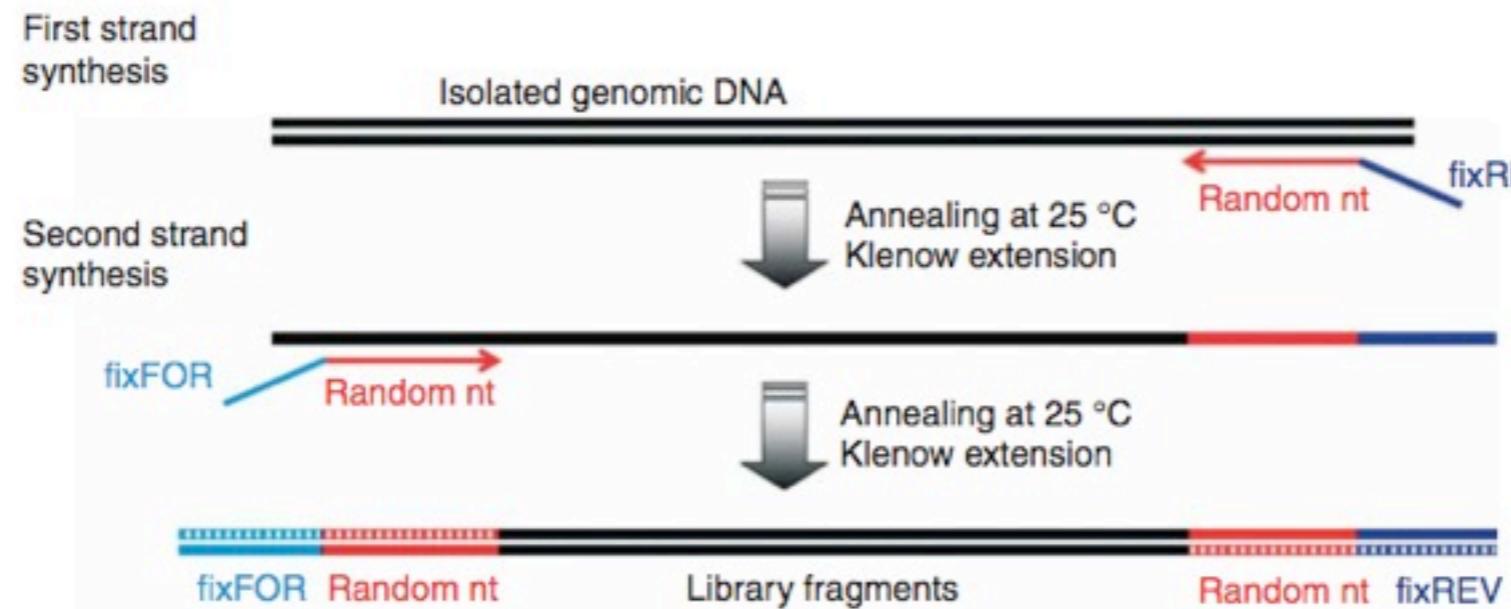


Future Directions

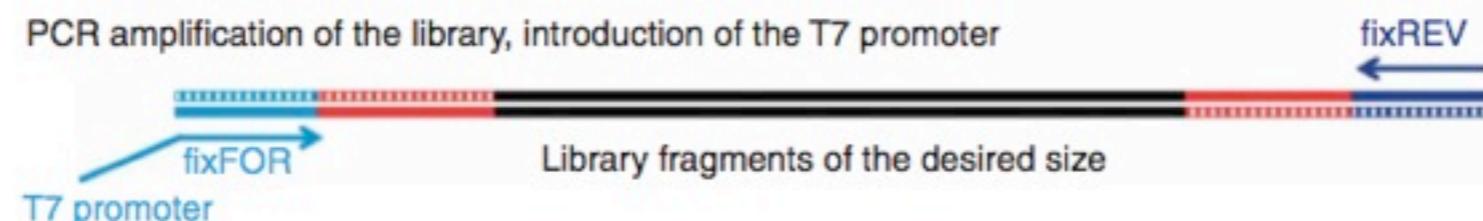
- Look into the enriched Hfq sequences
 - What are the motifs?
 - In what ways do they differ from the background?
 - Can Neutral SELEX aid in the bioinformatic analyses of SELEX pools?
 - Why are certain dinucleotides enriched?







Selection of fragments with the desired size by denaturing gel electrophoresis



Initial DNA pool

