

# Automated gating of flow cytometry data

Carmen Bruckmann

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# Introduction to flow cytometry

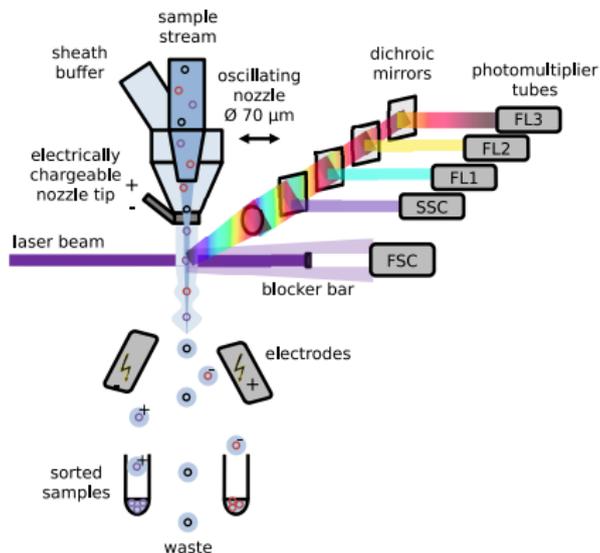


Figure: Adapted from: J. Lambrecht, *The community sensor - Monitoring and control of microbiome dynamics in anaerobic processes*, PhD Dissertation, 2020

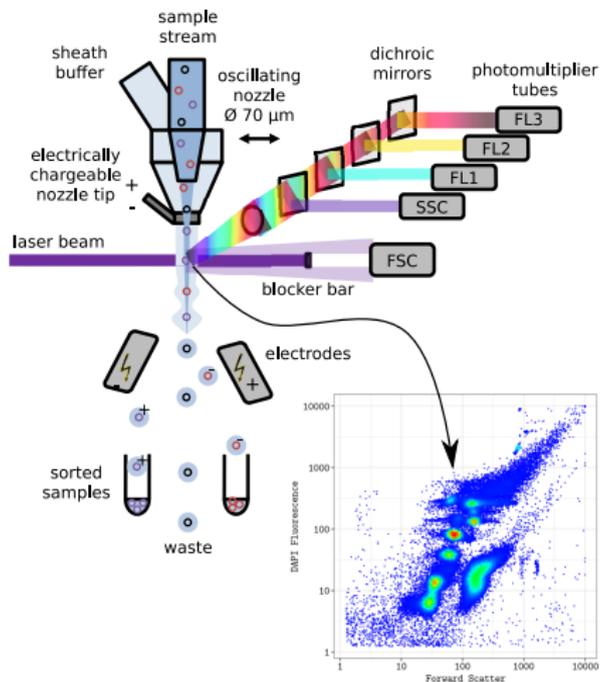
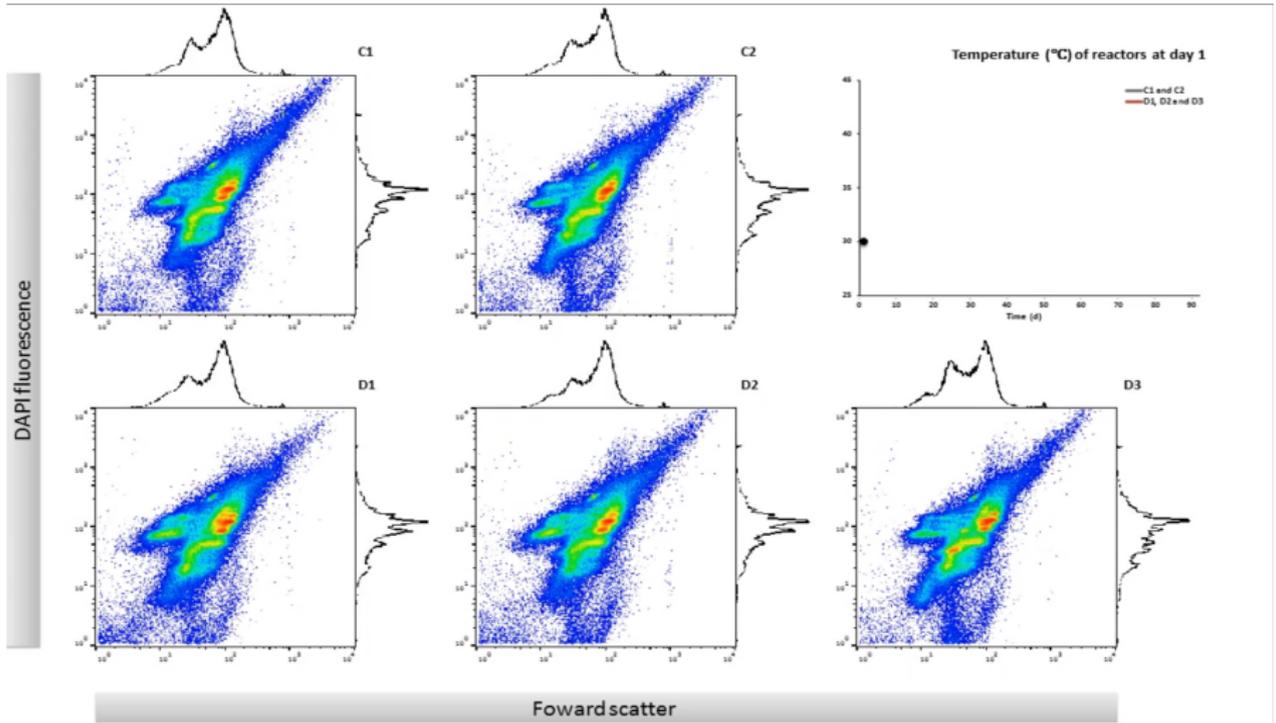


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# Introduction to flow cytometry



Liu et al., Neutral mechanisms and niche differentiation in steady-state insular microbial communities revealed by single cell analysis, *Environmental Microbiology*, 2019

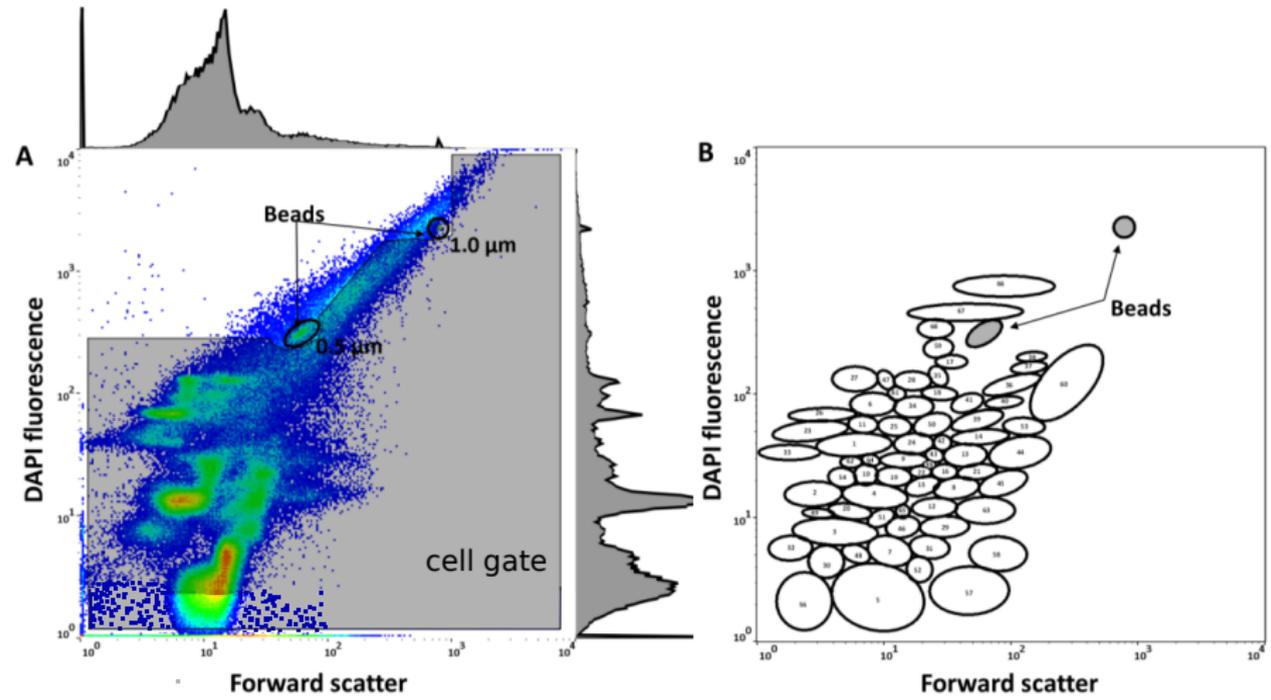


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# Underlying ecological questions

- How diverse is the community?

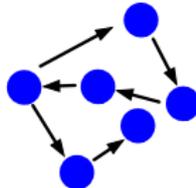
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- How does the microbial community develop over time?



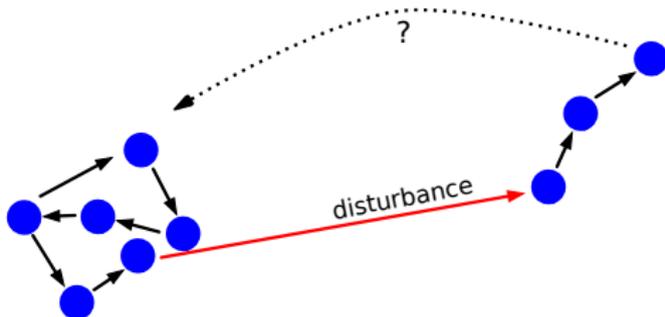
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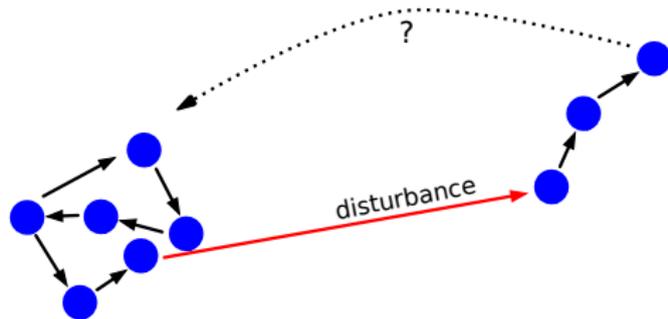
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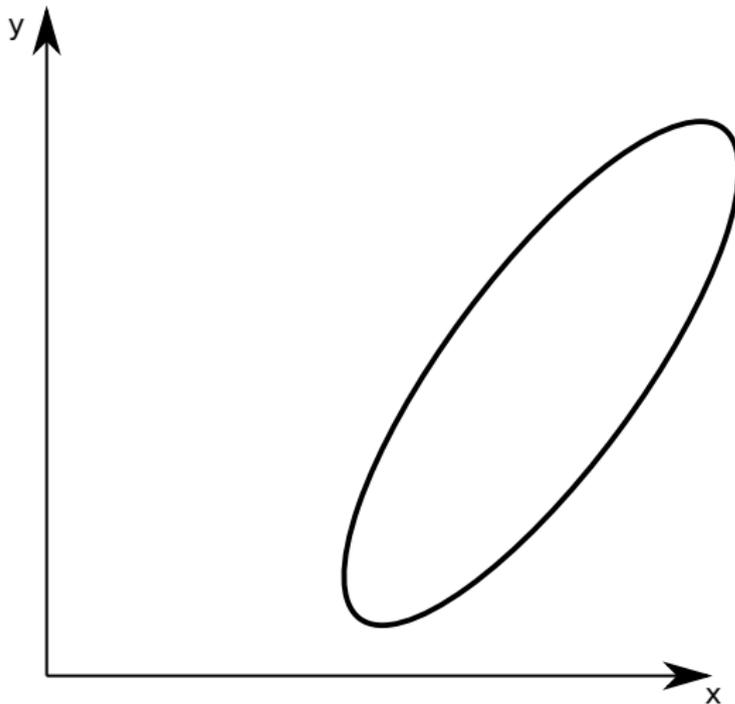
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⇒ **Need of automated gating!**

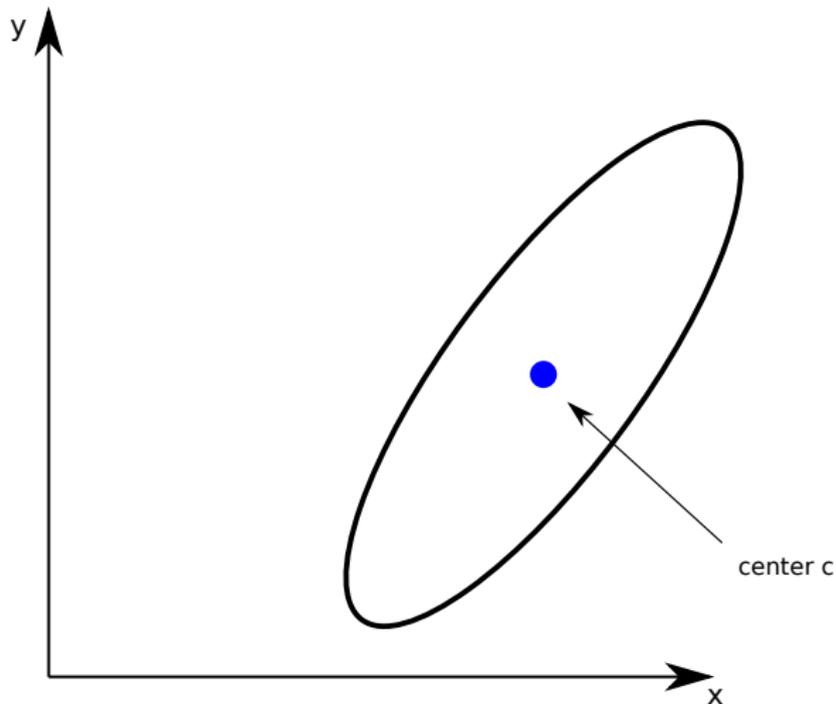
# Introduction

What is an ellipse?



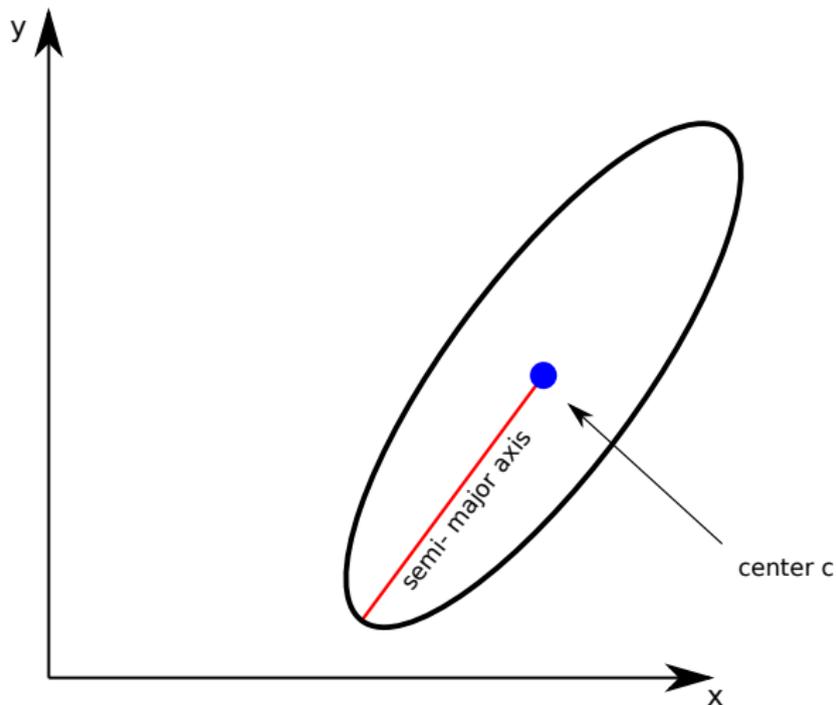
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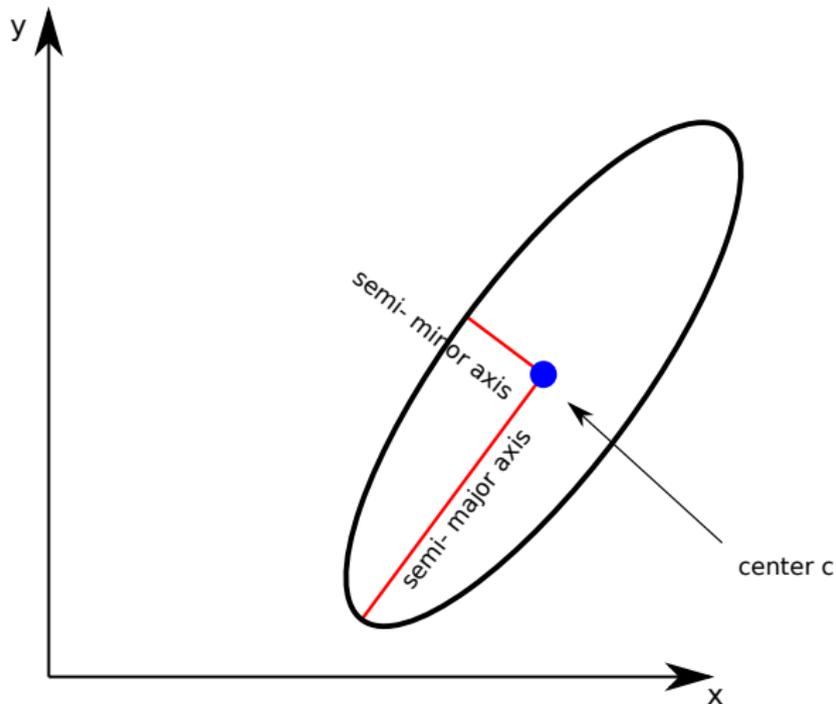
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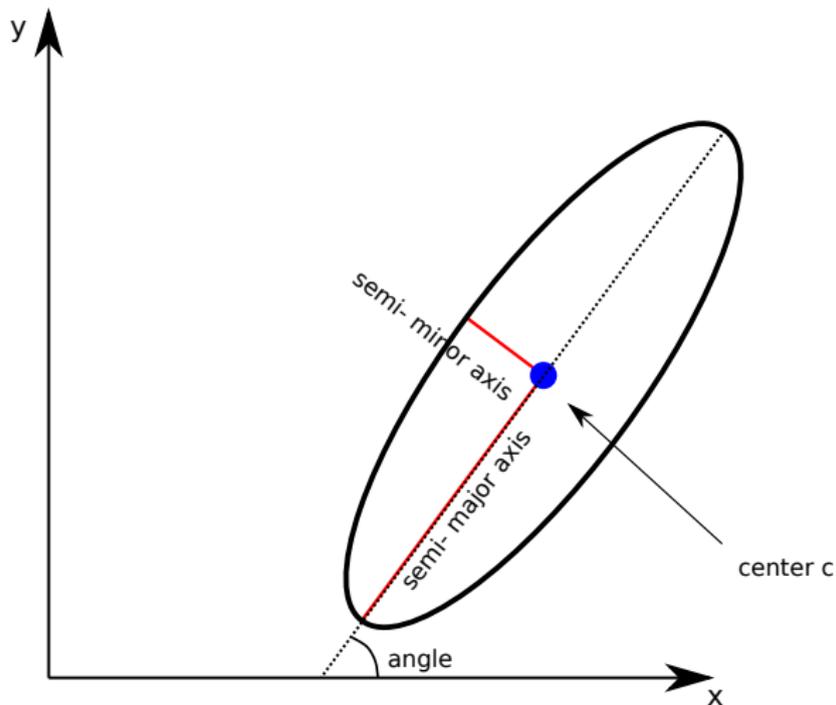
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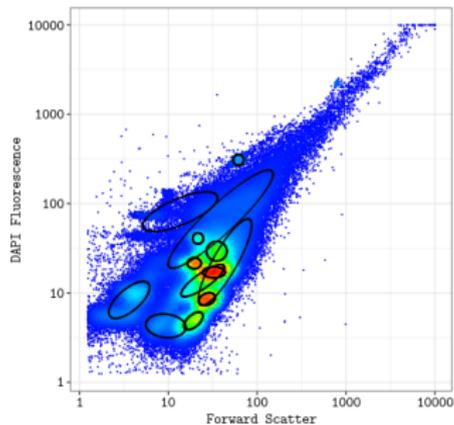
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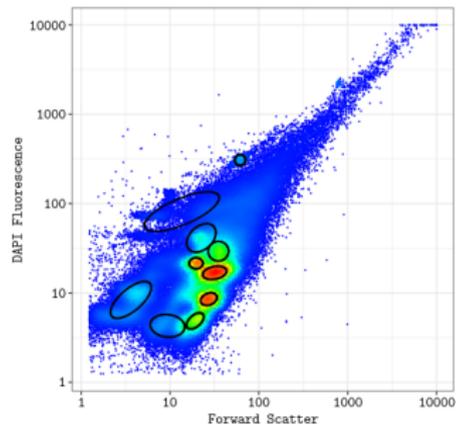
# Introduction

What is an ellipse?





flowEMMi v1



flowEMMi v2

# But how to remove the overlaps?

## Merging

- Let  $E_1$  and  $E_2$  be two ellipses that overlap more than a given threshold,  
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- Let  $\Sigma_1$  and  $\Sigma_2$  be the covariance matrices and let  $\mu_1$  and  $\mu_2$  be the centers of  $E_1$  and  $E_2$ , respectively.

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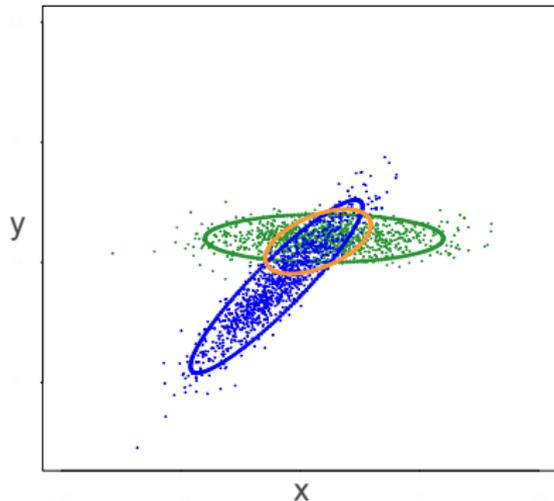
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- In the end, the original ellipses  $E_1$  and  $E_2$  will be deleted and replaced by  $E_c$ , i.e. by  $\mu_c$  and  $\Sigma_c$ .

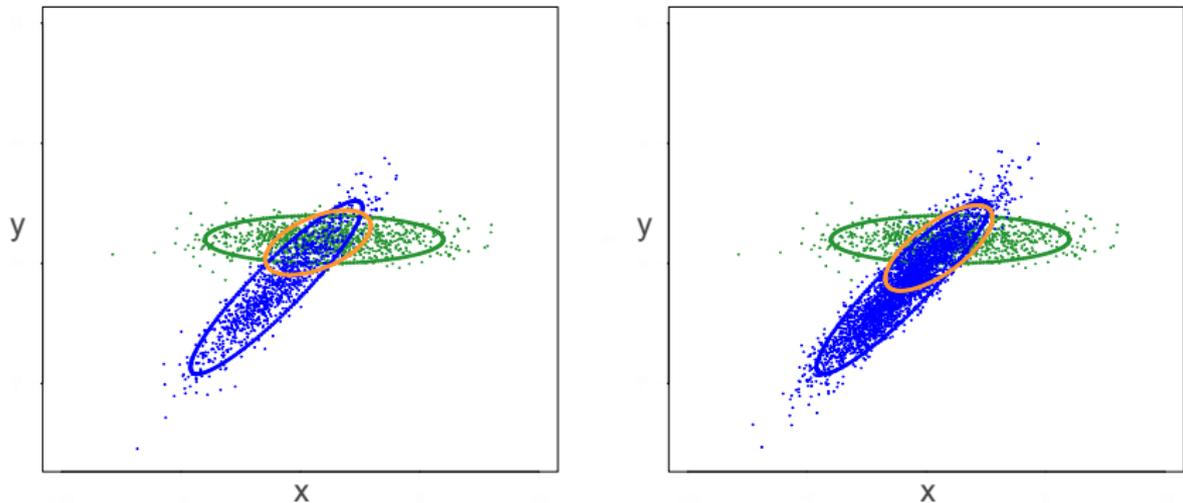
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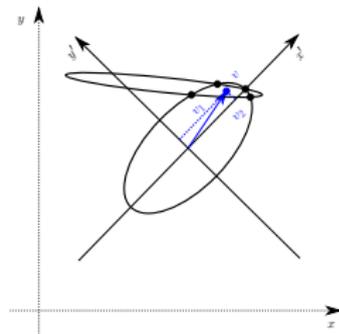


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## Shrinking

Let  $E_1$  and  $E_2$  be two ellipses that overlap with  $o = \text{size of intersection area}$ .

Let  $w_1$  and  $w_2$  be the weights of  $E_1$  and  $E_2$  in the mixture model, respectively. Then:



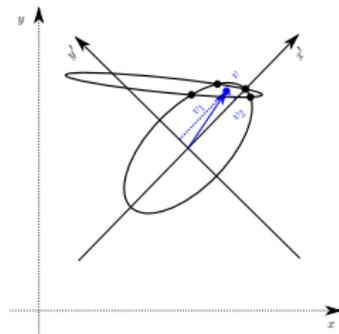
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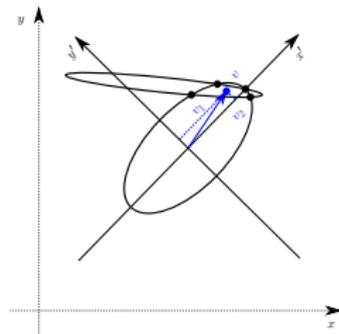
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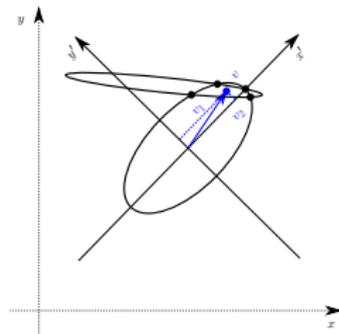
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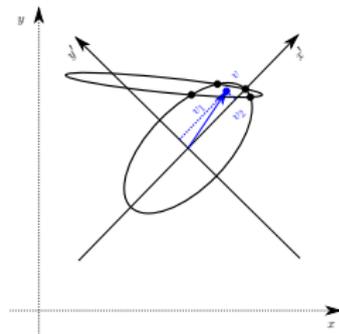
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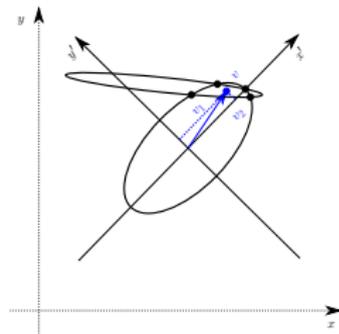
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5. Recalculate  $E_1$ :

$$\text{major}_{\text{new}} = \text{major} \cdot \left( 1 - \sqrt{\frac{o}{|E_1|} \cdot \frac{|v_1|}{|v_1| + |v_2|} \cdot \frac{w_2}{w_1 + w_2}} \right)$$

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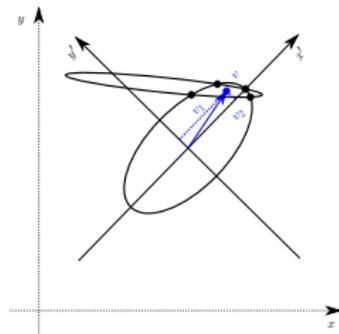
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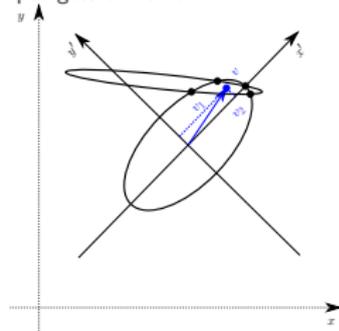
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6. Recalculate  $E_2$  in the same way.
7. Repeat steps 1. to 6. until  $E_1$  and  $E_2$  do not overlap anymore or until one ellipse gets too small.



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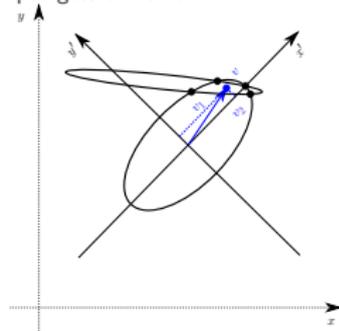
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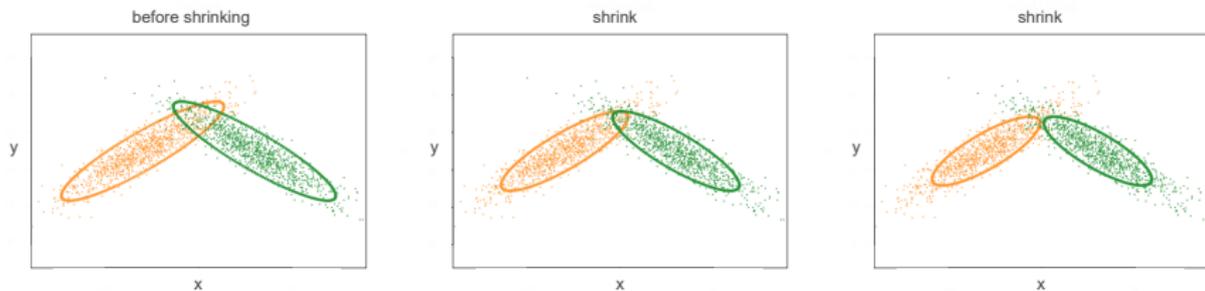
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6. Recalculate  $E_2$  in the same way.
7. Repeat steps 1. to 6. until  $E_1$  and  $E_2$  do not overlap anymore or until one ellipse gets too small.
8. If one ellipse gets too small, then merge the original  $E_1$  and  $E_2$  instead.



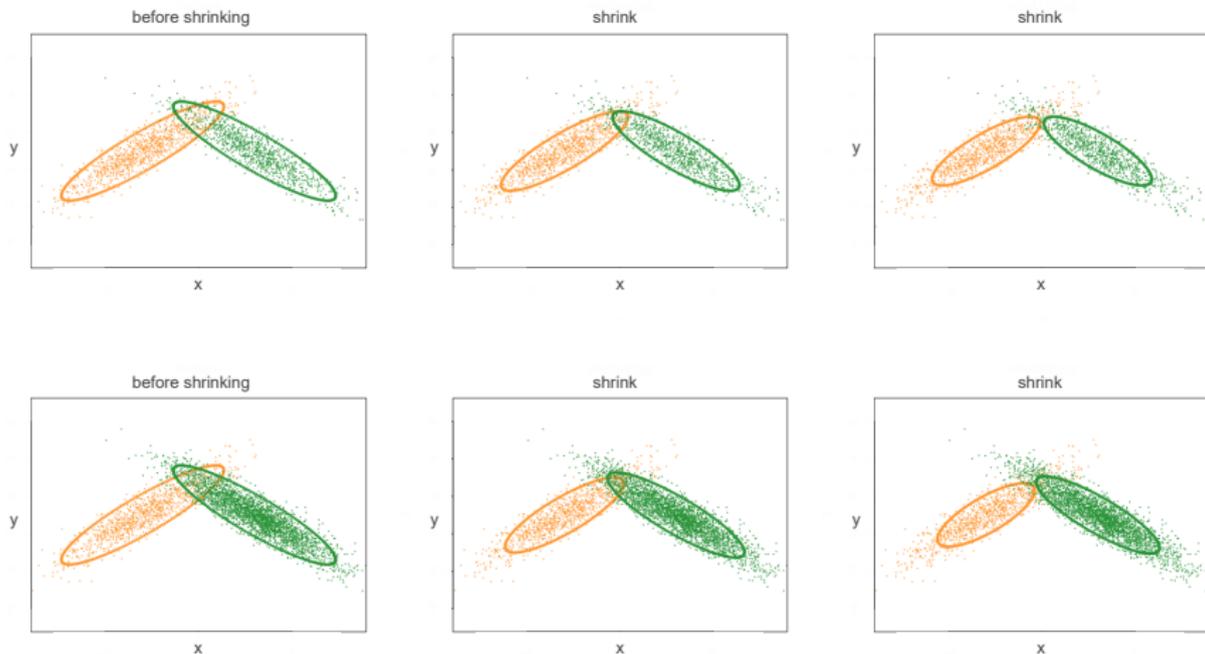
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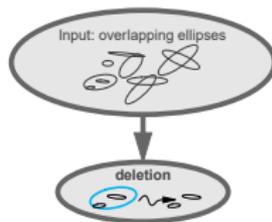


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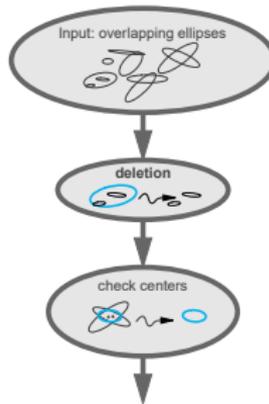
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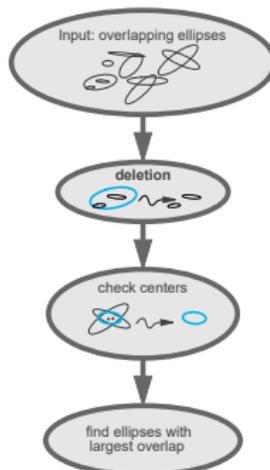


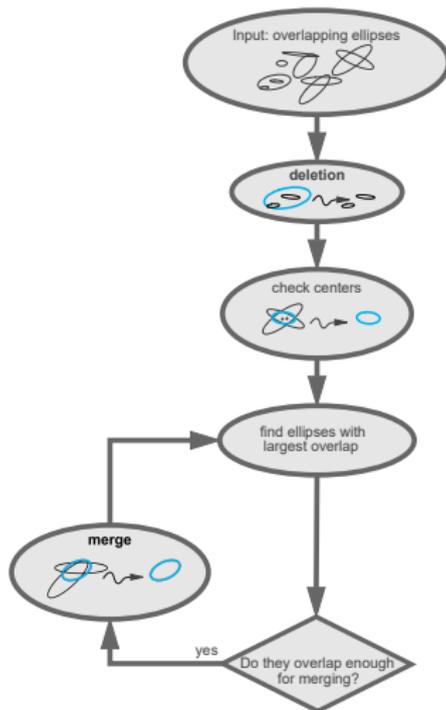


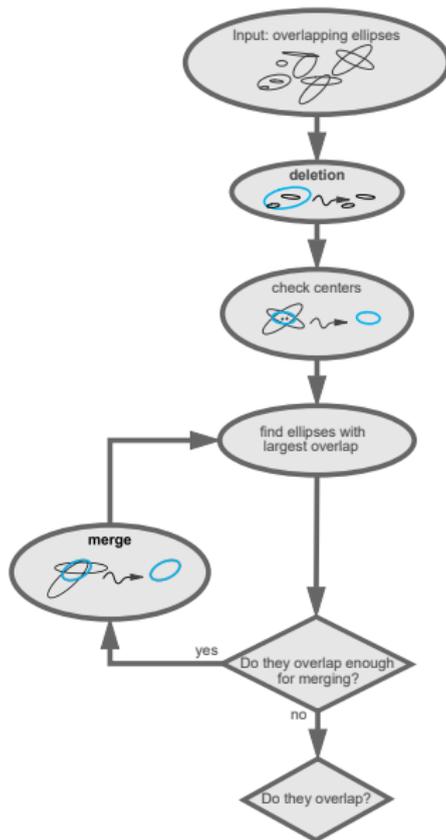
# Workflow

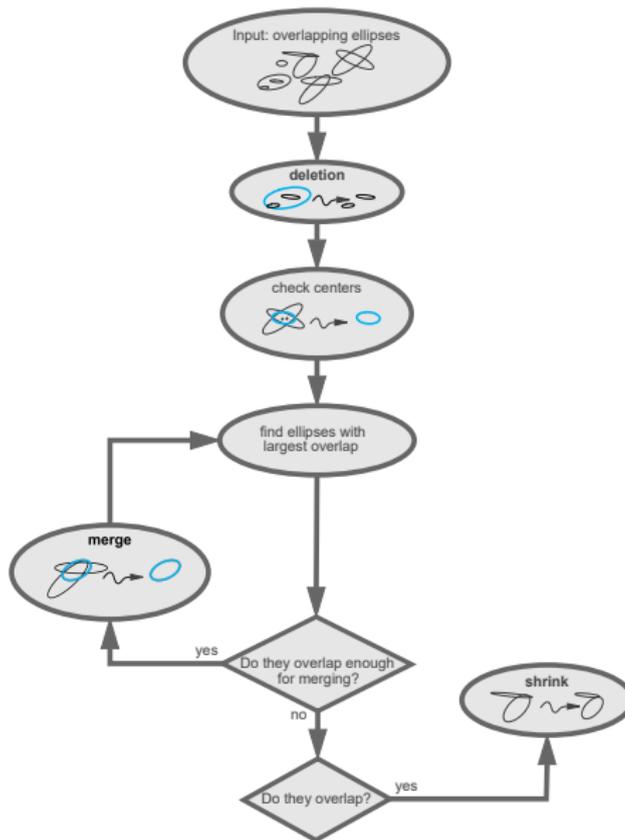


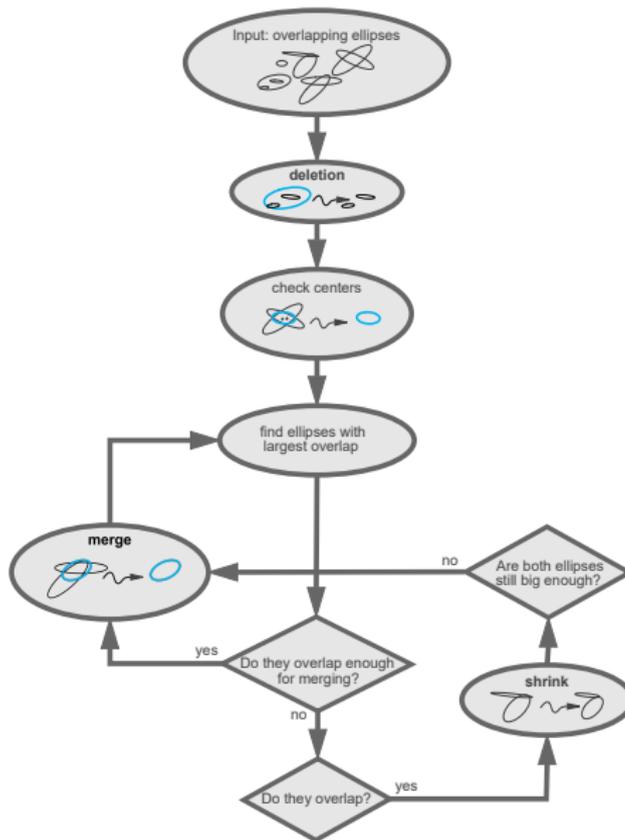
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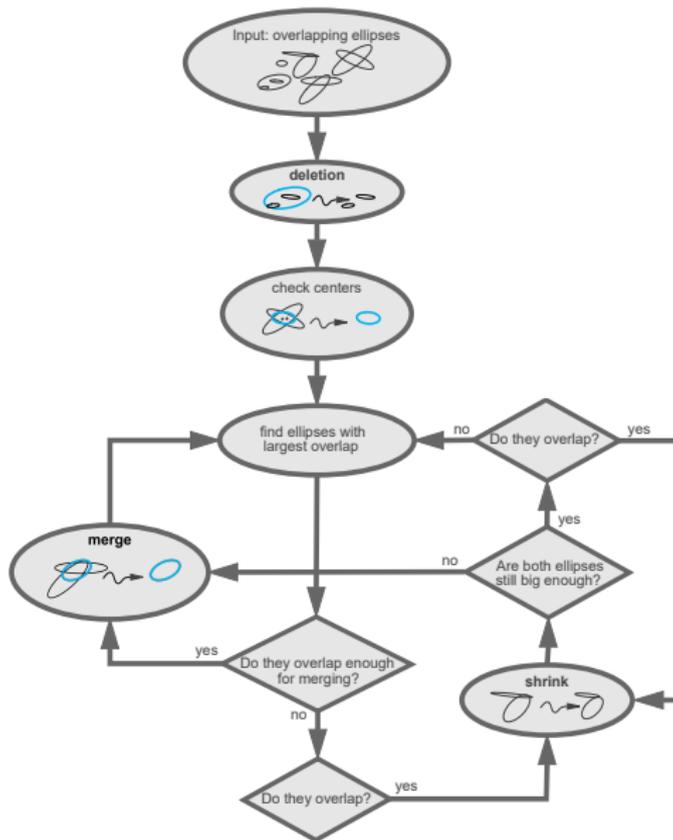


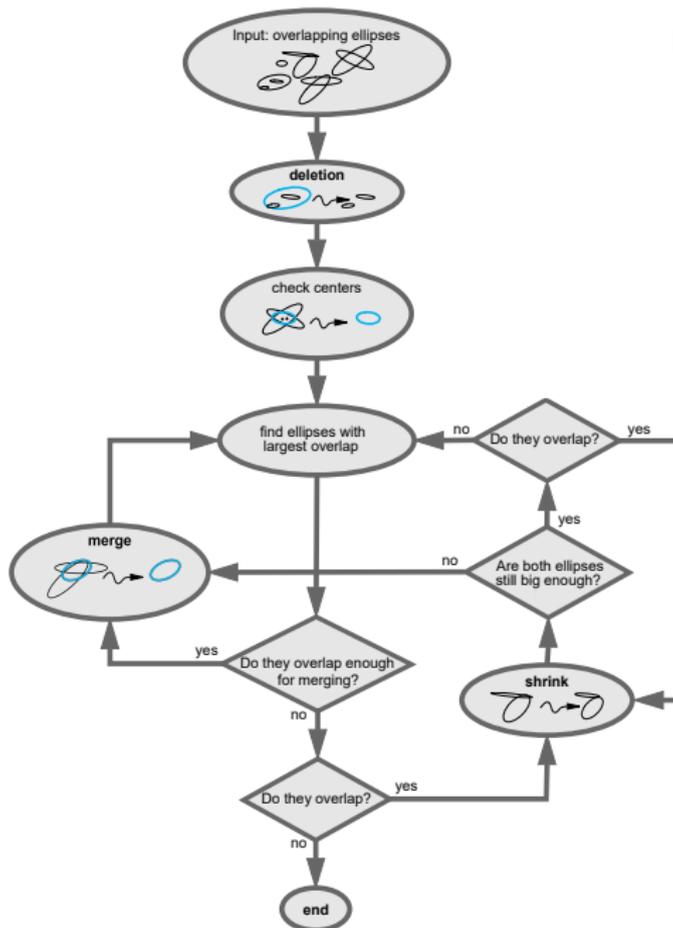




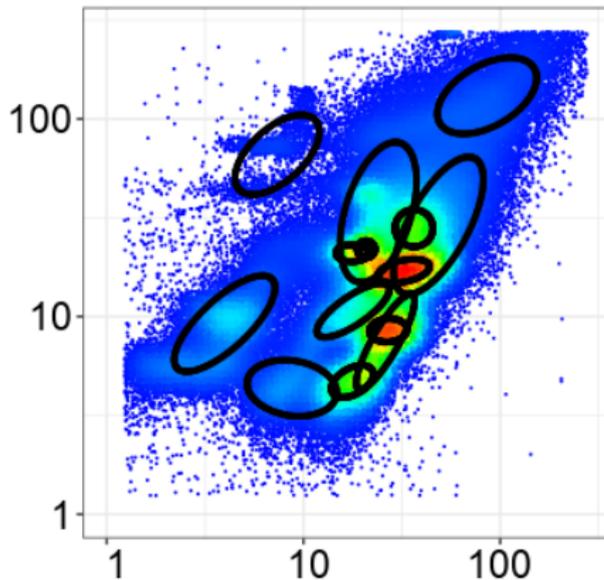




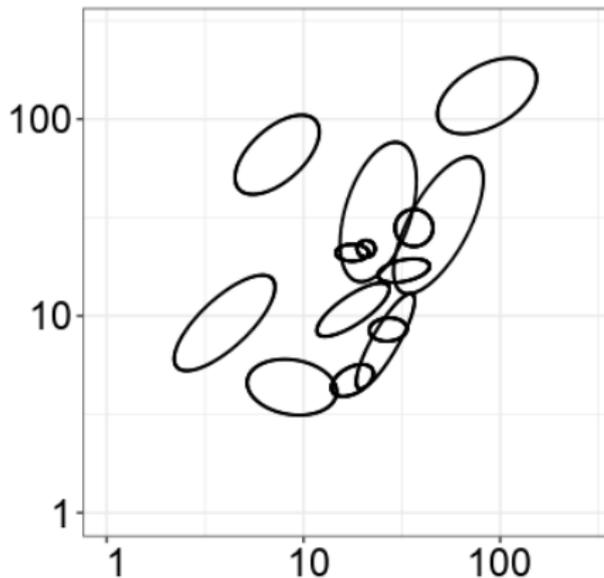




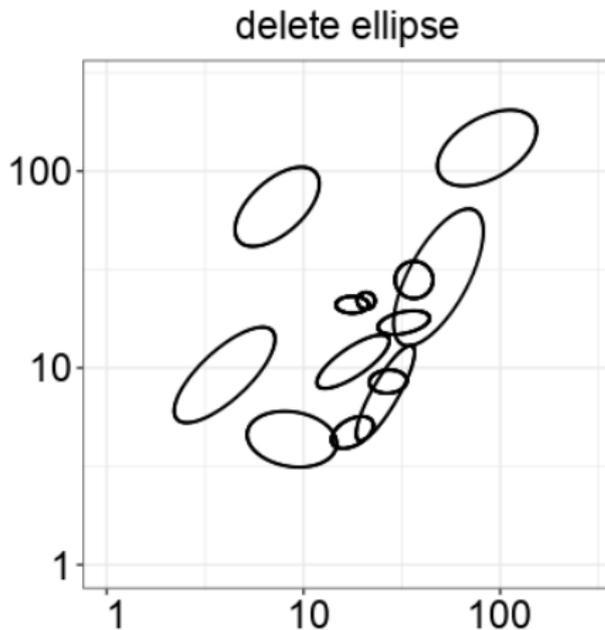
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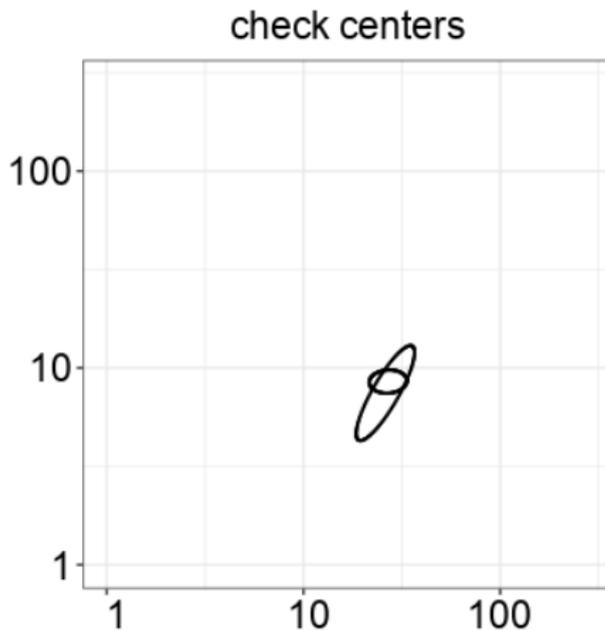
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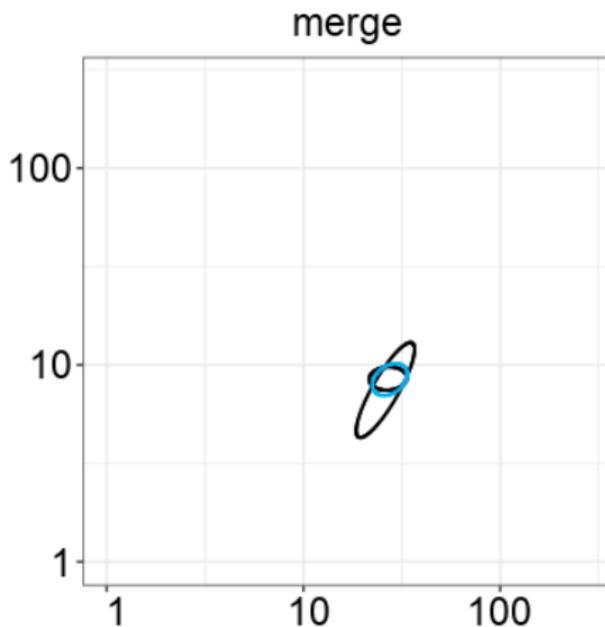
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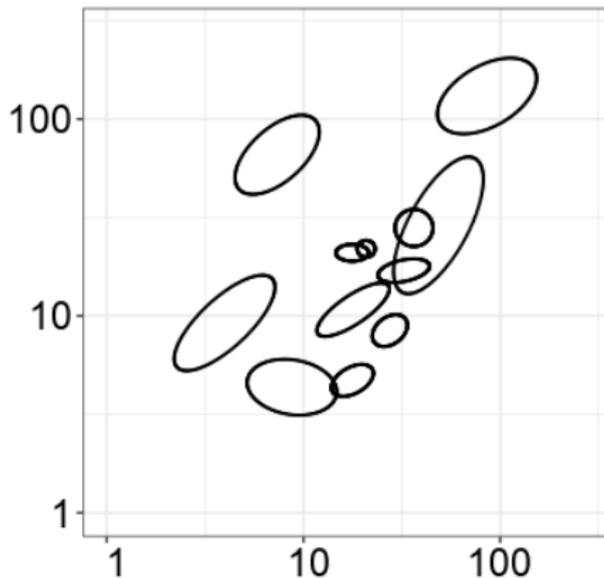
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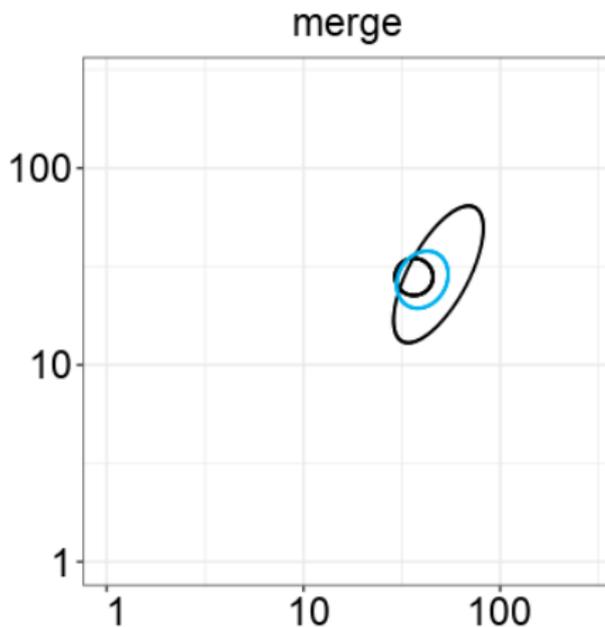
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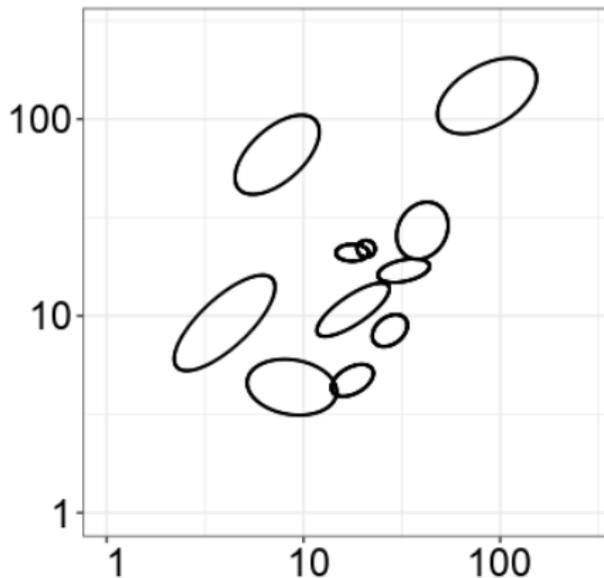
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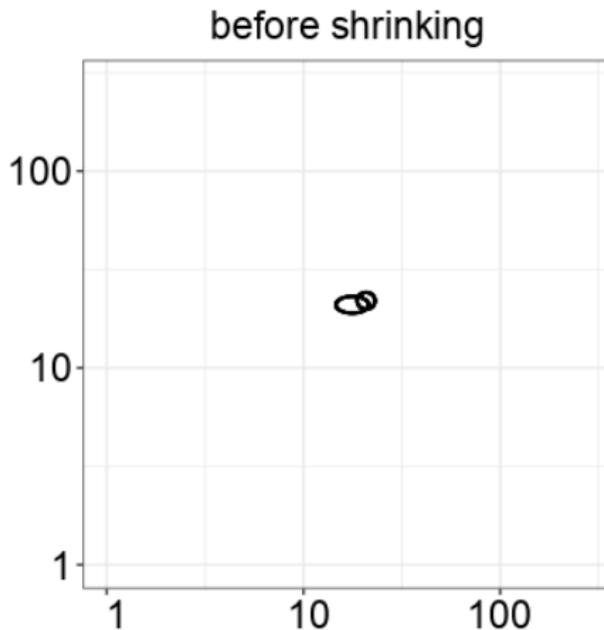
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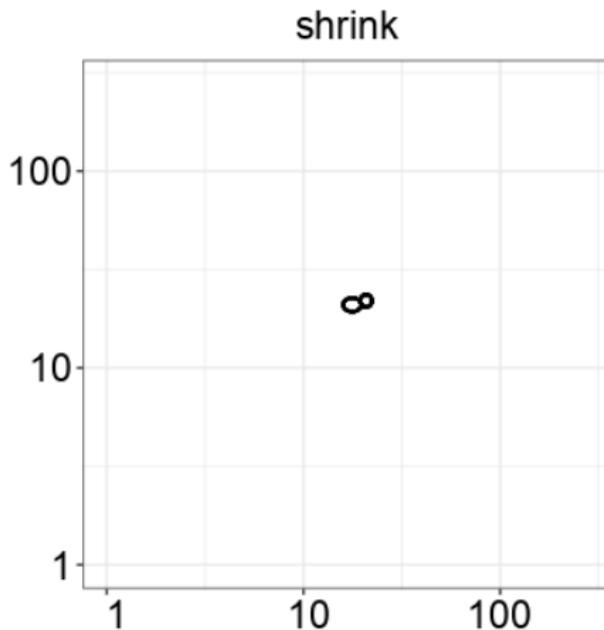
# Example



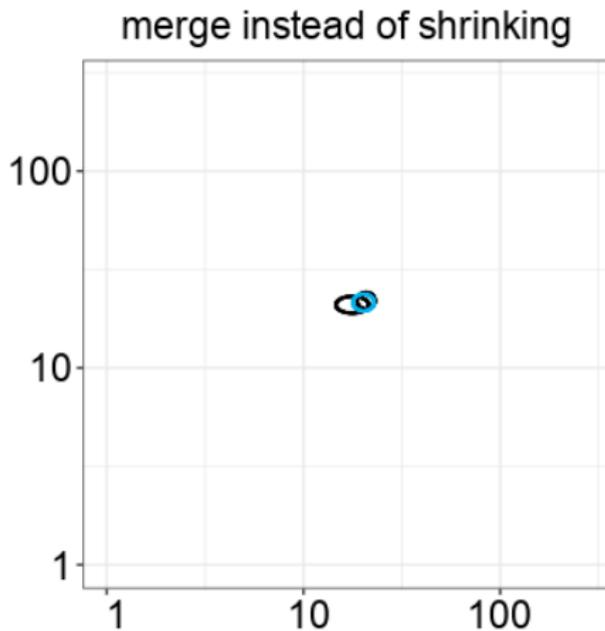
# Example



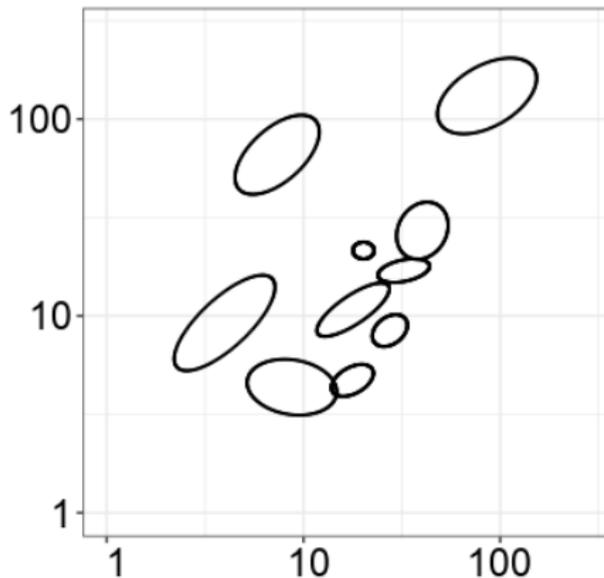
# Example



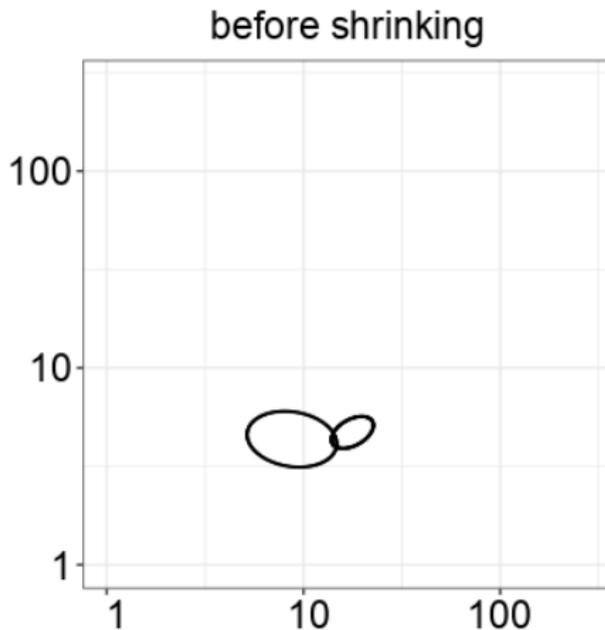
# Example



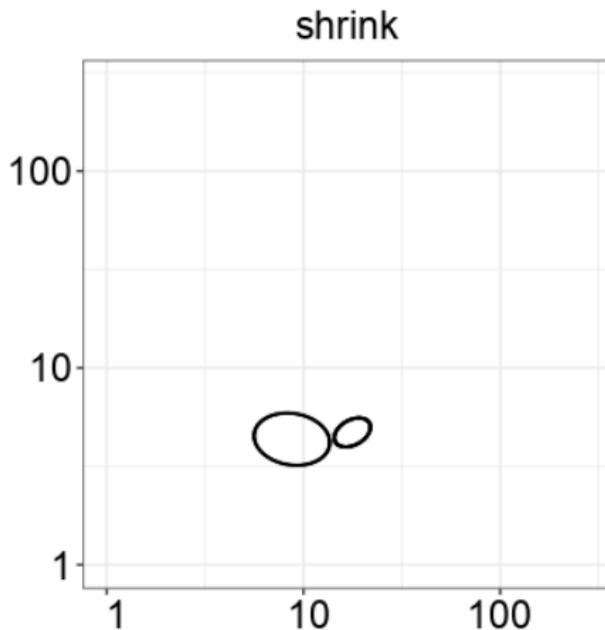
# Example



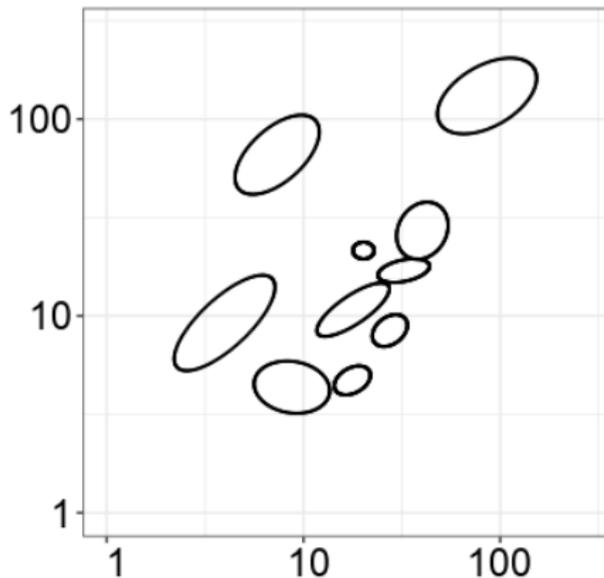
# Example



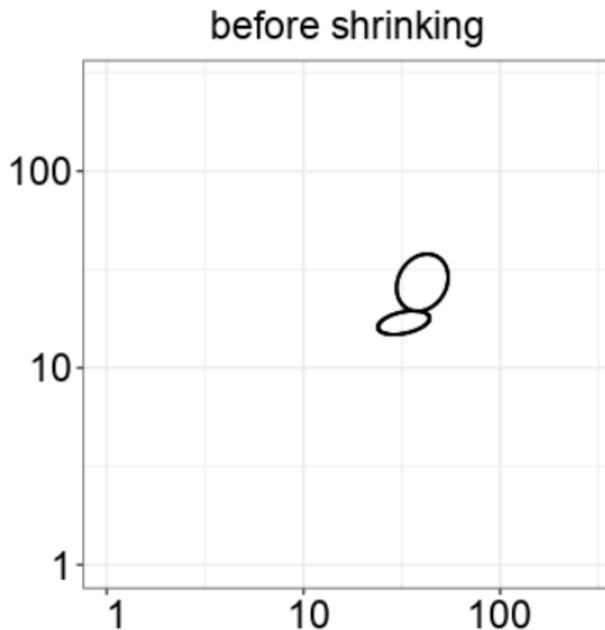
# Example



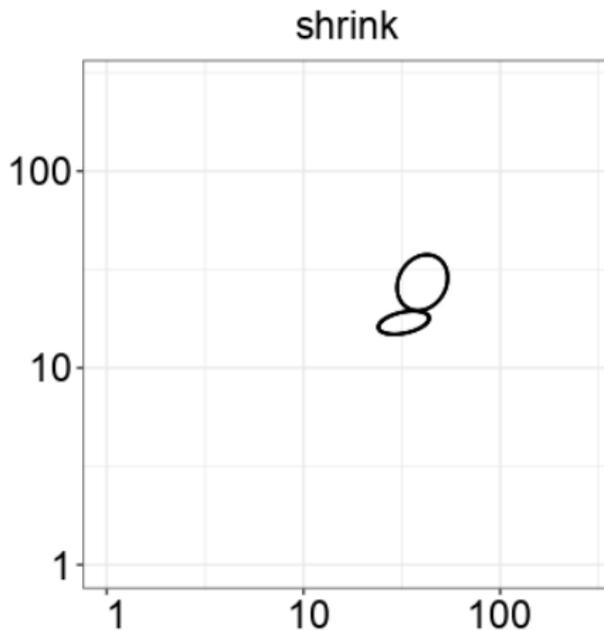
# Example



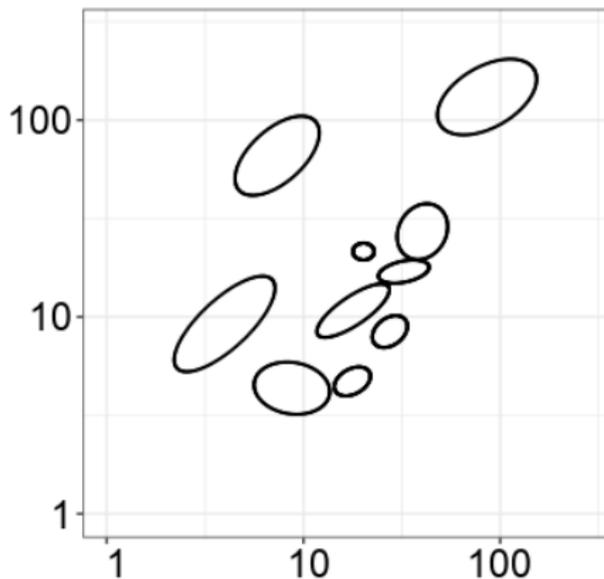
# Example



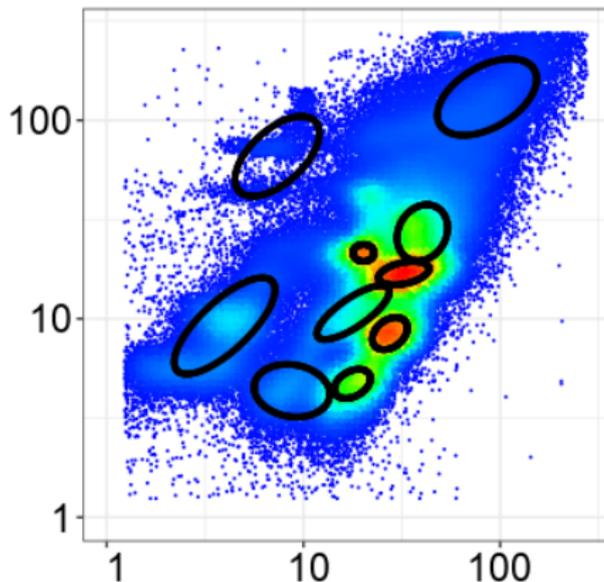
# Example



# Example

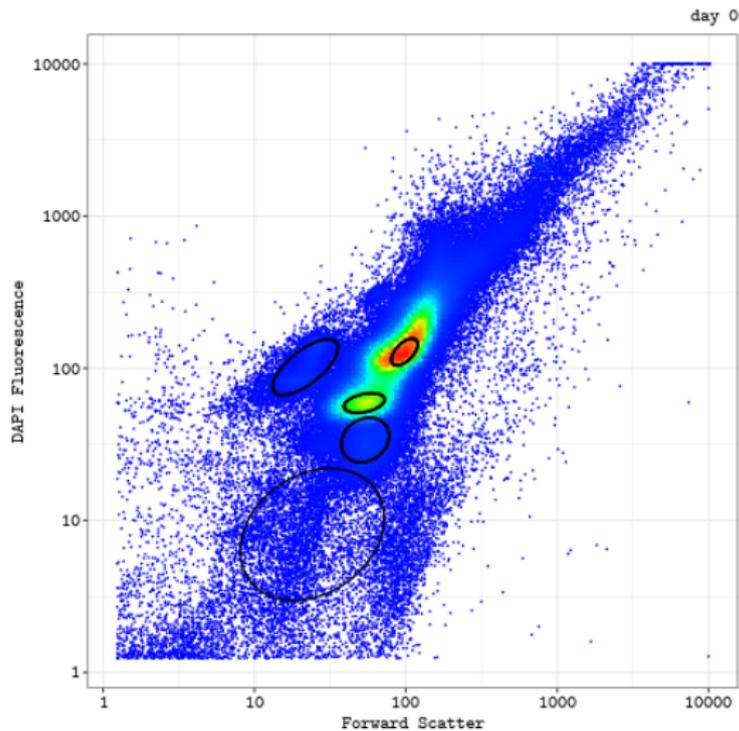


# Example



## Example 2

### Automated gating of single samples



- If we are given a time series of flow cytometry data, we would like to have a gate template that approximately fits every time point.

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- Input: The ellipses of all the samples (combined into a single set)
- Take out ellipses with a weight below a given threshold

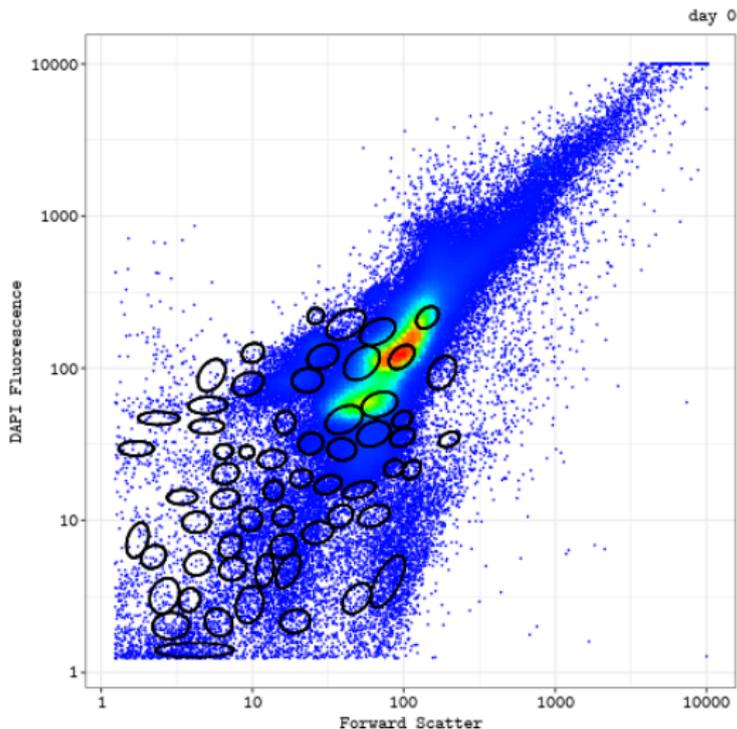
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- Remove the overlaps in this set of ellipses

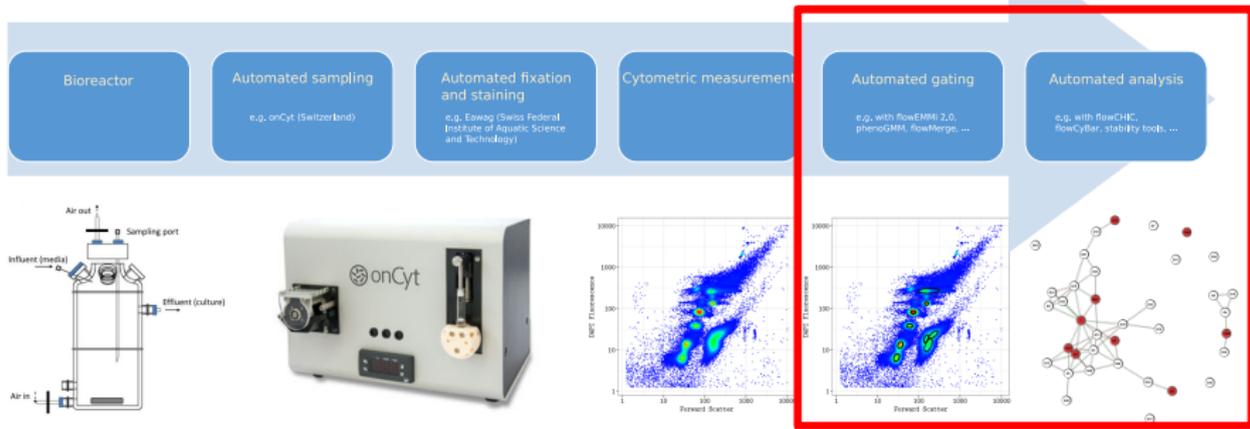
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- With the gate template we can calculate *cell numbers per gate and sample*

# Example 3

## Automatically generated gate template





- Liu et al., **Neutral mechanisms and niche differentiation in steady-state insular microbial communities revealed by single cell analysis**, *Environmental Microbiology*, 2019
- onCyt, <https://www.eawag.ch/de/news-agenda/news-plattform/newsarchiv/archiv-detail/online-durchflusszytometrie-sensor-fuer-bakterienkonzentrationen/>
- Cichocki et al., **Bacterial mock communities as standards for reproducible cytometric microbiome analysis**, *Nature Protocols*, 2020
- Ludwig et al., **flowEMMI: an automated model-based clustering tool for microbial cytometric data**, *BMC Bioinformatics*, 2019
- Koch et al., **flowCyBar: Analyze flow cytometric data using gate information**, *R package version 1.30.0*, 2019

The screenshot shows the BiTCa Analyze Tool interface in an Electron window. The title bar indicates the date and time as 14. Feb. 10:39. The application menu includes File, Edit, View, Window, and Help. The main interface is titled "flowEMMi v2" and features a sidebar on the left with navigation options: flowCHIC, flowEMMi v2 (selected), flowCyBar, and settings. The main area contains several configuration panels:

- channel x:** A dropdown menu showing "no fcs file found".
- x start:** A text input field containing "1500".
- x end:** A text input field containing "50000".
- channel y:** A dropdown menu showing "no fcs file found".
- y start:** A text input field containing "1500".
- y end:** A text input field containing "50000".
- use a cell-gate from FlowJo
- threshold for deletion:** A text input field containing "0,8".
- threshold for merging:** A text input field containing "0,6".
- inits:** A text input field containing "10".
- init fraction:** A text input field containing "0,01".
- final fraction:** A text input field containing "1".
- alpha:** A text input field containing "0,7".
- log
- disable parallelism
- remove overlaps
- when to remove overlaps:** A text input field containing "20".
- min minor:** A text input field containing "700".
- convergence:** A text input field containing "0,01".
- max cluster:** A text input field containing "15".
- min cluster:** A text input field containing "3".
- cluster bracket:** A text input field containing "3".
- 
- 
- Monitor Directory:**  Monitor Directory
- filepath (fcs directory):**

At the top of the main area, there are three buttons: "Gating of a single sample", "Gate template", and "Calculate cell numbers | Plot".

# Acknowledgements



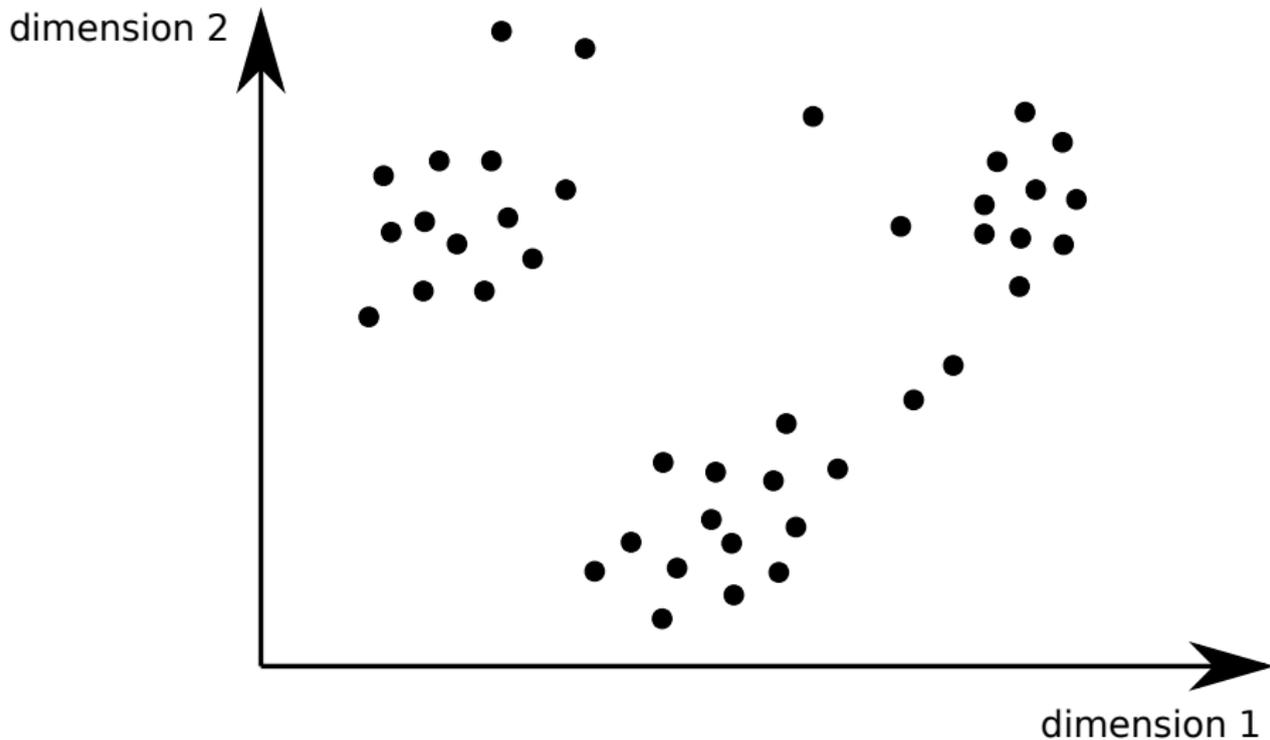
Bundesministerium  
für Wirtschaft  
und Energie



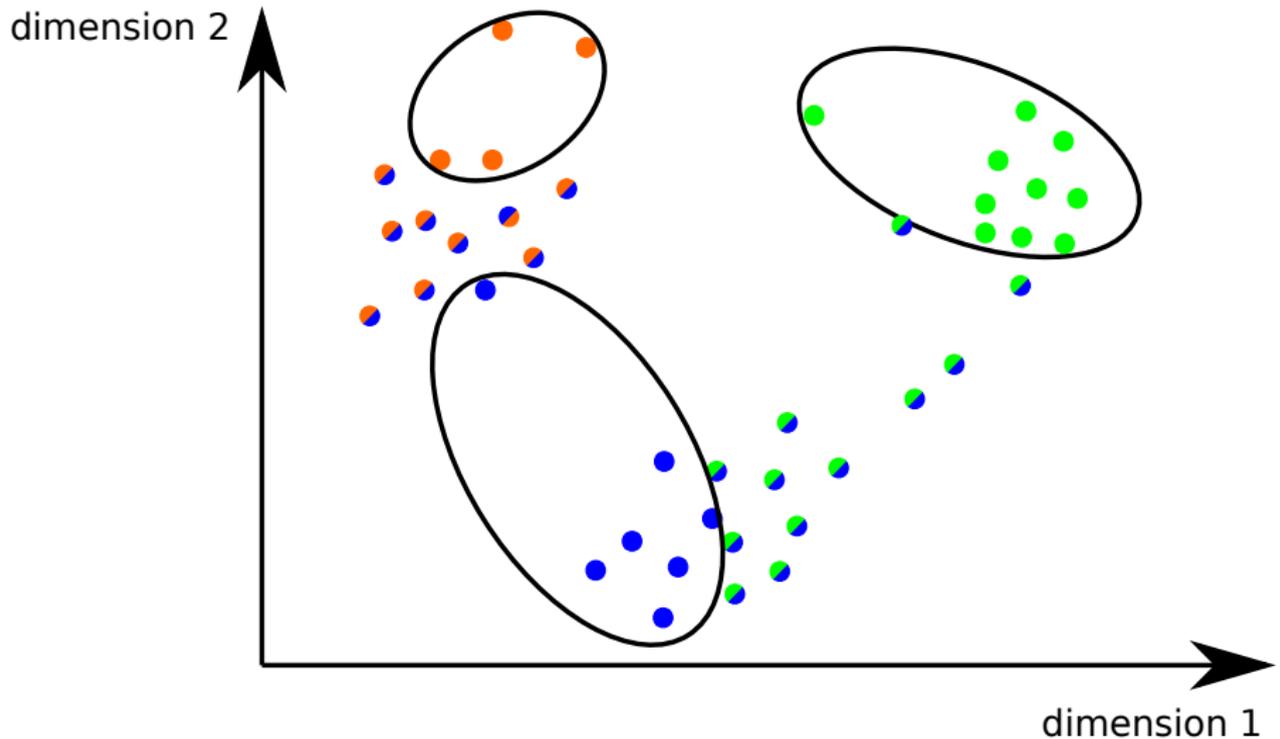
ScaDS.AM  
DRESDEN LEIPZIG

**Thank you for your attention and your support!**

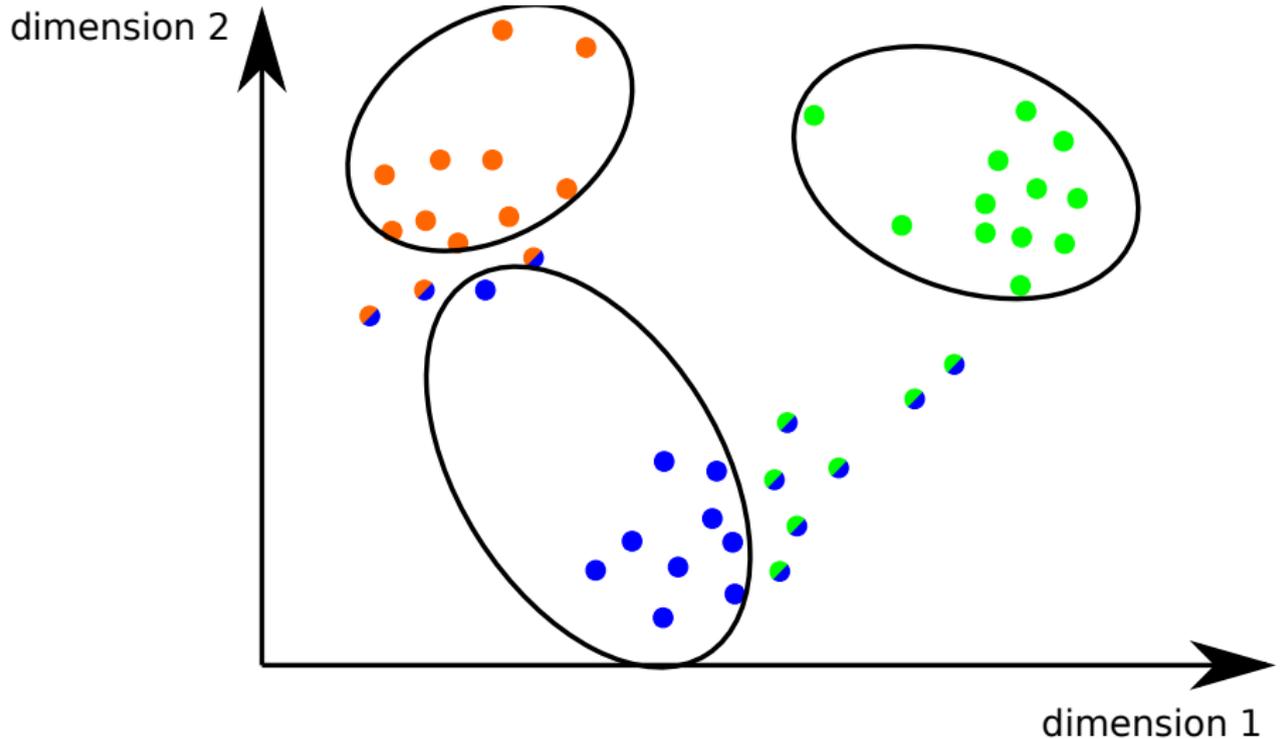
# Appendix: The EM-algorithm



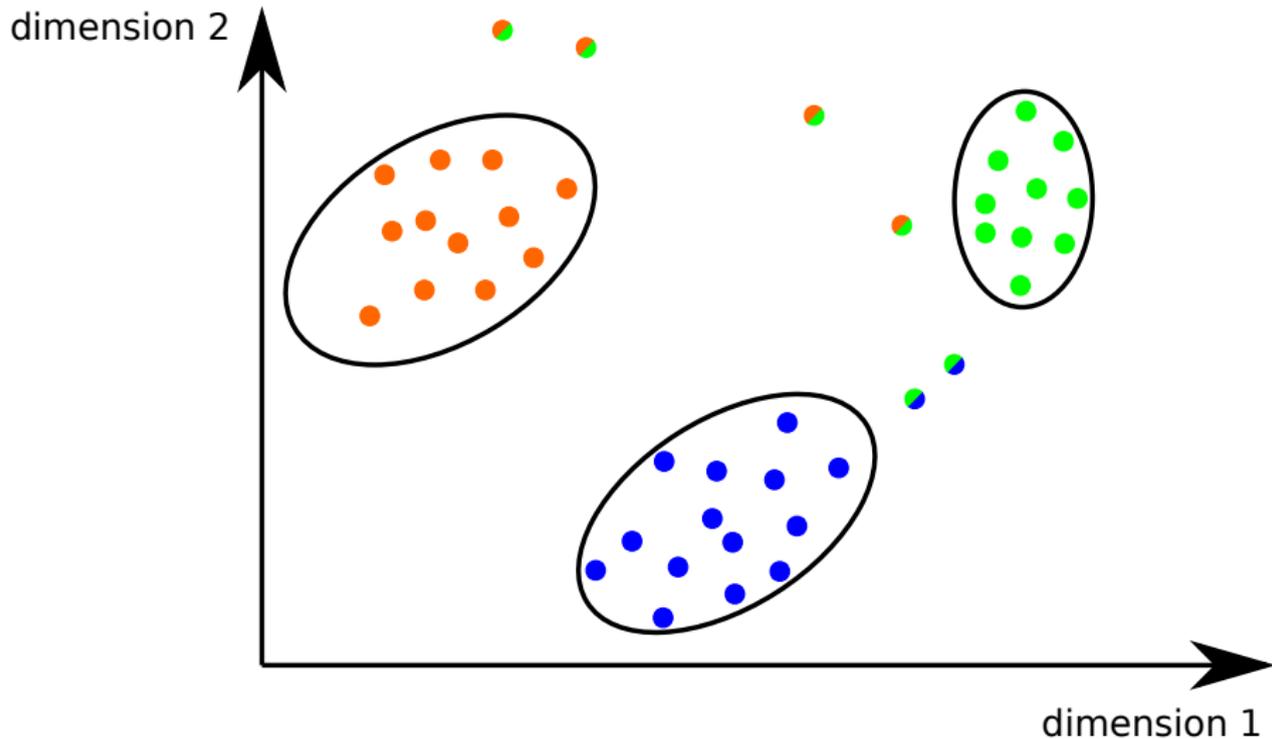
# Appendix: The EM-algorithm



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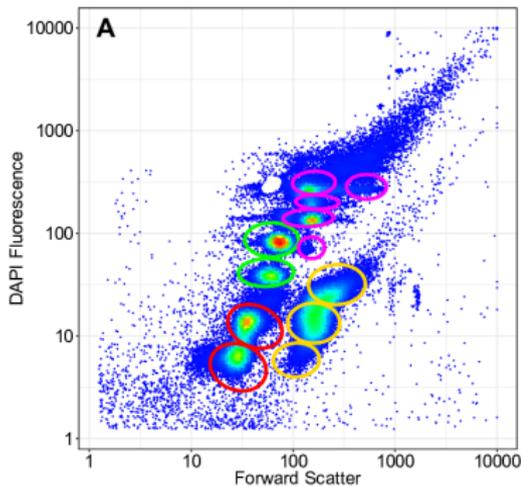


# Appendix: The EM-algorithm



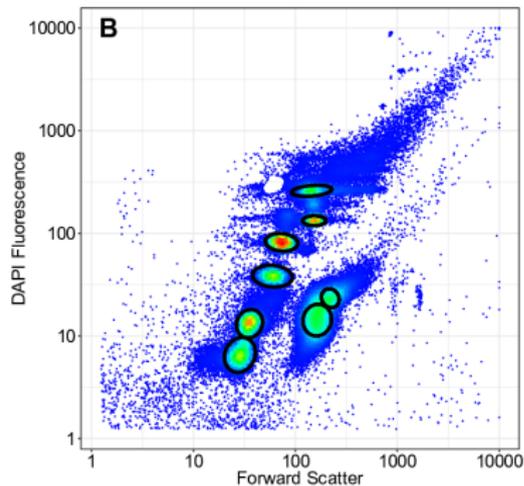
# mock communities

## manual gating



- *Stenotrophomonas rhizophila*
- *Escherichia coli*
- *Kocuria rhizophila*
- *Paenibacillus polymyxa*

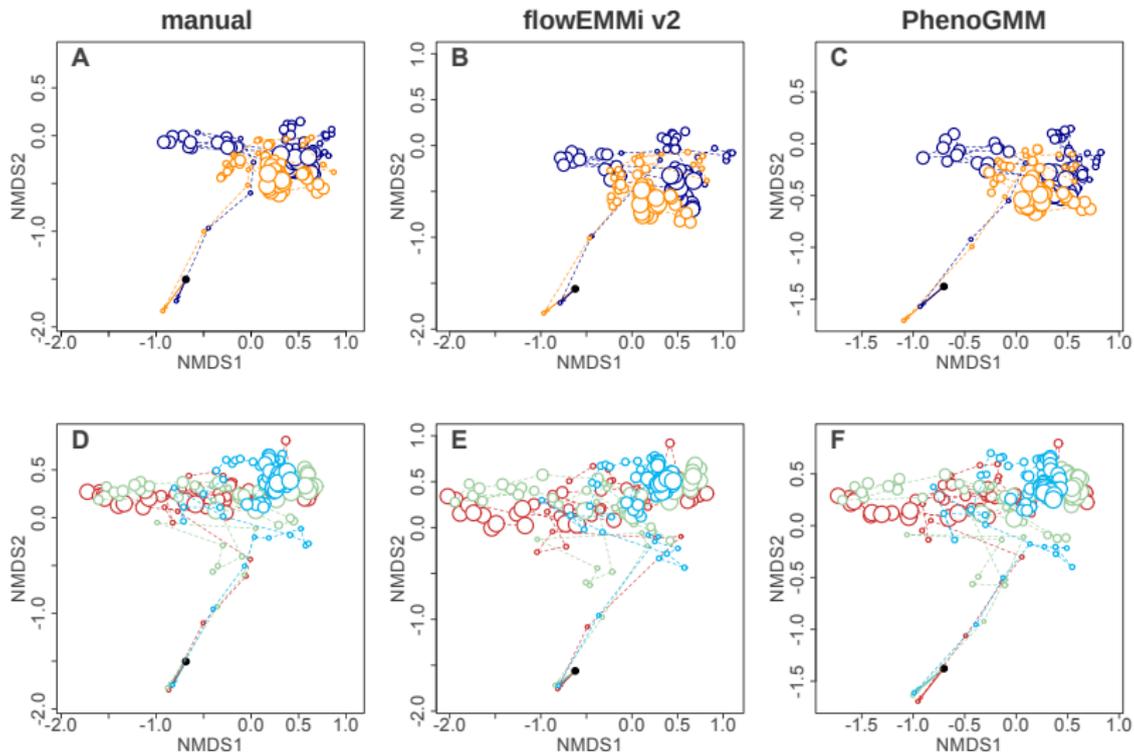
## flowEMMi v2



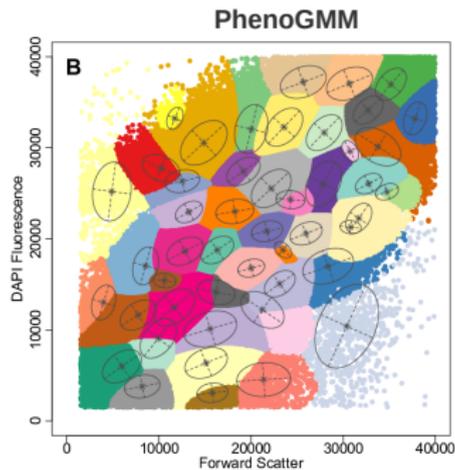
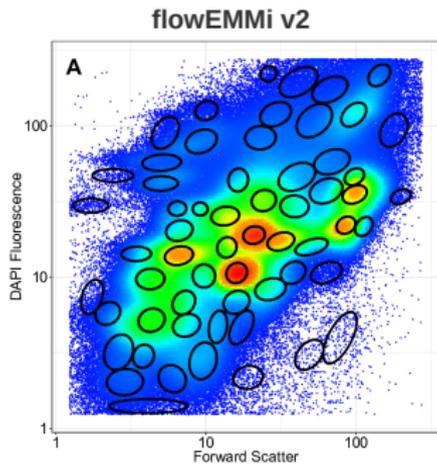
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Cichocki et al., **Bacterial mock communities as standards for reproducible cytometric microbiome analysis**, *Nature Protocols*, 2013

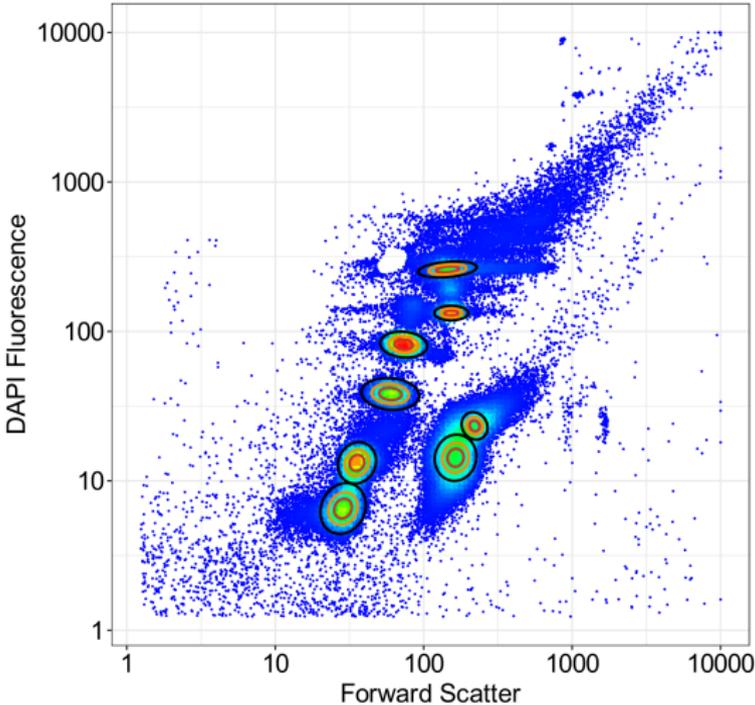
# Appendix: Community dynamics



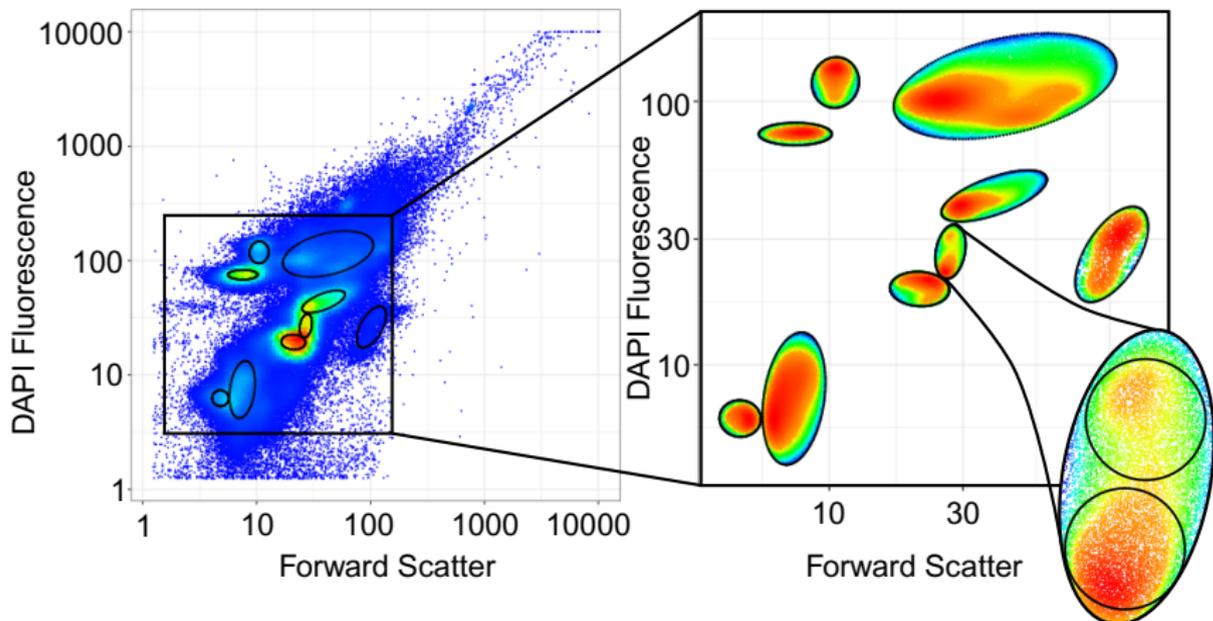
# Appendix: Community dynamics



# Appendix: Ellipse sizes



## Appendix: Hierarchical Gating



# Merging

## Merging: The mathematical details

- $\Sigma_1$  and  $\Sigma_2$  of  $E_1$  and  $E_2$  are both symmetric and positive definite  
⇒ use the Cholesky decomposition and re-scale them:

$$\Sigma_i = L_i \cdot D_i \cdot L_i^T \quad \text{for } i = 1, 2 \quad (1)$$

$$\Sigma'_i = L_i \cdot \frac{1}{w_i} \cdot D_i \cdot L_i^T \quad \text{for } i = 1, 2 \quad (2)$$

- Then, the new  $\mu_c$  and  $\Sigma'_c$  of  $E'_c$ , which will be re-scaled to  $E_c$  later, can be derived as the product of two Gaussians:

$$\mu_c = \left( \Sigma_1'^{-1} + \Sigma_2'^{-1} \right) \cdot \left( \Sigma_1'^{-1} \cdot \mu_1 + \Sigma_2'^{-1} \cdot \mu_2 \right) \quad (3)$$

$$\Sigma'_c = \left( \Sigma_1'^{-1} + \Sigma_2'^{-1} \right)^{-1} = \Sigma'_1 \cdot \left( \Sigma'_1 + \Sigma'_2 \right)^{-1} \cdot \Sigma'_2 \quad (4)$$

- For the re-adjustment of the weights, we again need to decompose  $\Sigma'_c$  and multiply the diagonal elements by the sum of the weights.

$$\Sigma'_c = L'_c \cdot D'_c \cdot L'^T_c \quad (5)$$

$$\Sigma_c = L'_c \cdot (w_1 + w_2) \cdot D'_c \cdot L'^T_c \quad (6)$$