

Mechanisms of molecular cooperation

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„*Homo Sociobiologicus*“ – Evolution of human cooperation

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Selforganization of Matter
and the Evolution of Biological Macromolecules

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I. Introduction

I.1. "Cause and Effect"

The question about the origin of life often appears as a question about "cause and effect". Physical theories of macroscopic processes usually involve answers to such questions, even if a statistical interpretation is given to the relation between "cause" and "effect". It is mainly due to the nature of this question that many scientists believe that our present physics does not offer any obvious explanation for the existence of life.

* Partially presented at the "Robbins Lectures" at Pennzo College, Chicago, in spring 1970.

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The Hypercycle

A Principle of Natural Self-Organization

Part A: Emergence of the Hypercycle

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This paper is the first part of a trilogy, which comprises a detailed study of a special type of functional organization and demonstrates its relevance with respect to the origin and evolution of life. Self-replicating macromolecules, such as RNA or DNA in a suitable environment exhibit a behavior, which we may call Darwinian and which can be formally represented by the concept of the quasi-species. A quasi-species is defined as a given distribution of macromolecular species with closely interrelated sequences, dominated by one or several (topological) master copies. External constraints enforce the selection of the best adapted distribution, continuously referred to as the wild-type. Most important for Darwinian behavior are the criteria for internal stability of the quasi-species. If these criteria are violated, the information stored in the nucleotide sequence of the master copy will disseminate irreversibly leading to an error catastrophe. As a consequence, selection and evolution of RNA or DNA molecules is limited with respect to the amount of information that can be stored in a single replicative unit. An analysis of experimental data regarding RNA and DNA replication at various levels of organization reveals that a sufficient amount of information for the build-up of a translation machinery can be gained only via integration of several different replicative units (replicative cycles) through reciprocal linkages. A stable functional integration then will raise the system to a new level of organization and thereby enlarge its information capacity considerably. The hypercycle appears to be such a form of organization.

Preview on Part B: The Abstract Hypercycle

The mathematical analysis of dynamical systems using methods of differential topology yields the result that there are only one type of mechanisms which fulfill the following requirements. The information stored in each single replicative unit (or reproductive cycle) must be maintained, i.e., the respective master copies must compete favorably with their error distributions. Despite their competitive behavior these units must establish a cooperation which includes all functionally integrated species. On the other hand, the cycle as a whole must continue to compete strongly with any other single entity or isolated ensemble which does not contribute to its integrated function.

These requirements are crucial for a selection of the best adapted functionally linked ensemble and its evolutive optimization. Only

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Molecular Quasi-Species¹

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The molecular quasi-species model describes the physicochemical organization of monomers into an ensemble of heteropolymers with combinatorial complexity by ongoing template polymerization. Polynucleotides belong to the simplest class of such molecules. The quasi-species limit represents the stationary distribution of macromolecular sequences maintained by chemical reactions effecting error-prone replication and by transport processes. It is obtained deterministically, by mass-action kinetics, as the dominant eigenvector of a value matrix, W , which is derived directly from chemical rate coefficients, but it also exhibits stochastic features, being composed of a significant fraction of unique individual macromolecular sequences. The quasi-species model demonstrates how macromolecular information originates through specific nonequilibrium autocatalytic reactions and thus forms a bridge between reaction kinetics and molecular evolution. Selection and evolutionary optimization appear as new features in physical chemistry. Concentration bias in the production of mutants is a new concept in population genetics, relevant to frequently mutating populations, which is shown to greatly enhance the optimization processes. The present theory relates to actually replicating ensembles, but this restriction is not essential. A sharp transition is exhibited between a drifting population of essentially random macromolecular sequences and a localized population of close relatives. This transition at a threshold error rate was found to depend on sequence lengths, distributions of selective values, and population sizes. It has been determined generally for complex landscapes and for special cases, and, it was shown to persist generally in the presence of nearly neutral mutants. Replication dynamics has much in common with the equilibrium statistics of complex spin systems: the error threshold is equivalent to a magnetic order-disorder transition. A rational function of the replication accuracy plays the role of temperature. Experimental data obtained from test-tube evolution of polynucleotides and from studies of natural virus populations support the quasi-species model. The error threshold seems to set a limit to the genome lengths of several classes of RNA viruses. In addition, the results are relevant even in eucaryotes where they contribute to the exon-intron debate.

Preview on Part C: The Abstract Hypercycle

A realistic model of a hypercycle relevant with respect to the origin of the genetic code and the translation machinery is presented. It includes the following features referring to natural systems:

- 1) The hypercycle has a sufficiently simple structure to admit an organization with finite probability under prebiotic conditions.
- 2) It permits a continuous emergence from slowly accumulated in RNA-like precursors, originally being members of a stable RNA quasi-species and having been amplified to a level of higher abundance.
- 3) The organizational structure and the properties of single functional units of this hypercycle are well reflected in the present genetic code in the translation apparatus of the prokaryotic cell, as well as in certain bacterial viruses.

I. The Paradigm of Unity and Diversity in Evolution

Why do millions of species, plants and animals, exist, while there is only one basic molecular machinery of the cell: one universal genetic code and unique chemicalities of the macromolecules?

The geneticists of our day would not hesitate to give an immediate answer to the first part of this question. Diversity of species is the outcome of the tremendous branching process of evolution with its myriads of single steps of reproduction and mutation. It in-

cludes very small numbers of different bases, even just two, readily provides numbers of different entities so enormous that neither consecutive nor parallel physical realization is possible. The physical chemistry of finite systems of such macromolecules must deal with both known regularities and the advent of unique copolymeric sequences. Normally this would present no difficulty in a statistical mechanical analysis of typical behavior, where rare events play no significant role, but with autocatalytic polymerization processes even unique single molecules may be amplified to determine the fate of the entire system. Potentially creative, self-organizing around unique events, the dynamics of the simplest living chemical system is invested with regularities that both allow and limit efficient adaptation. The quasi-species model is a study of these regularities.

The fundamental regularity in living organisms that has invited explanation is adaptation. Why are organisms so well fitted to their environments? At a more chemical level, why are enzymes

optimal catalysts? Darwin's theory of natural selection has provided biologists with a framework for the answer to this question. The present model is constructed along Darwinian lines but in terms of specific macromolecules, chemical reactions, and physical processes that make the notion of survival of the fittest precise. Not only does the model give an understanding of the physical limitations of adaptation, but also it provides new insight into the role of chance in the process. For an understanding of the structure of this minimal chemical model it is first necessary to recall the conceptual basis of Darwin's theory.

Darwin recognized that new inheritable adaptive properties were not induced by the environment but arose independently in the production of offspring. Lasting adaptive changes in a population could only come about by natural selection of the heritable traits or genotype based on the full characteristics or phenotype relevant for producing offspring. A process of chance, i.e., uncorrelated with the developed phenotype, control changes in the genotype from one generation to the next and generates the diversity necessary for selection. Three factors have probably prevented chemists from gaining a clear insight into these phenomena in the past, despite the discovery of the polymeric nature of the genotype (DNA): the complexity of a minimum replication phenotype, the problem of dealing with a huge number of variants, and the nonequilibrium nature of these ongoing processes.

The formulation of a tractable chemical model based on Darwin's principle may be understood in several steps:

1. The major constituents of the system have to be inherently self-reproductive. Only two classes of molecules are presently

¹ This is an abridged account of the quasi-species theory that has been submitted in comprehensive form to *Advances in Chemical Physics*.

² E. Eigen, M. McCaskill, J. S. Schuster, P. Adam, *Chem. Phys.*, in press.

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1. Cyclic reaction networks " catalysts
2. Cyclic catalytic networks " autocatalysts
3. Cyclic autocatalytic networks " hypercycles
4. Neutrality - a source for coexistent competitors

1. **Cyclic reaction networks ' catalysts**
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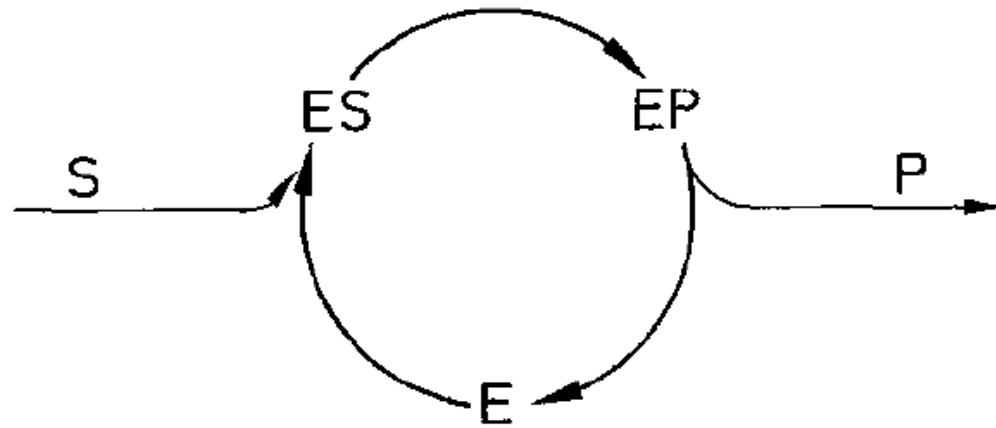
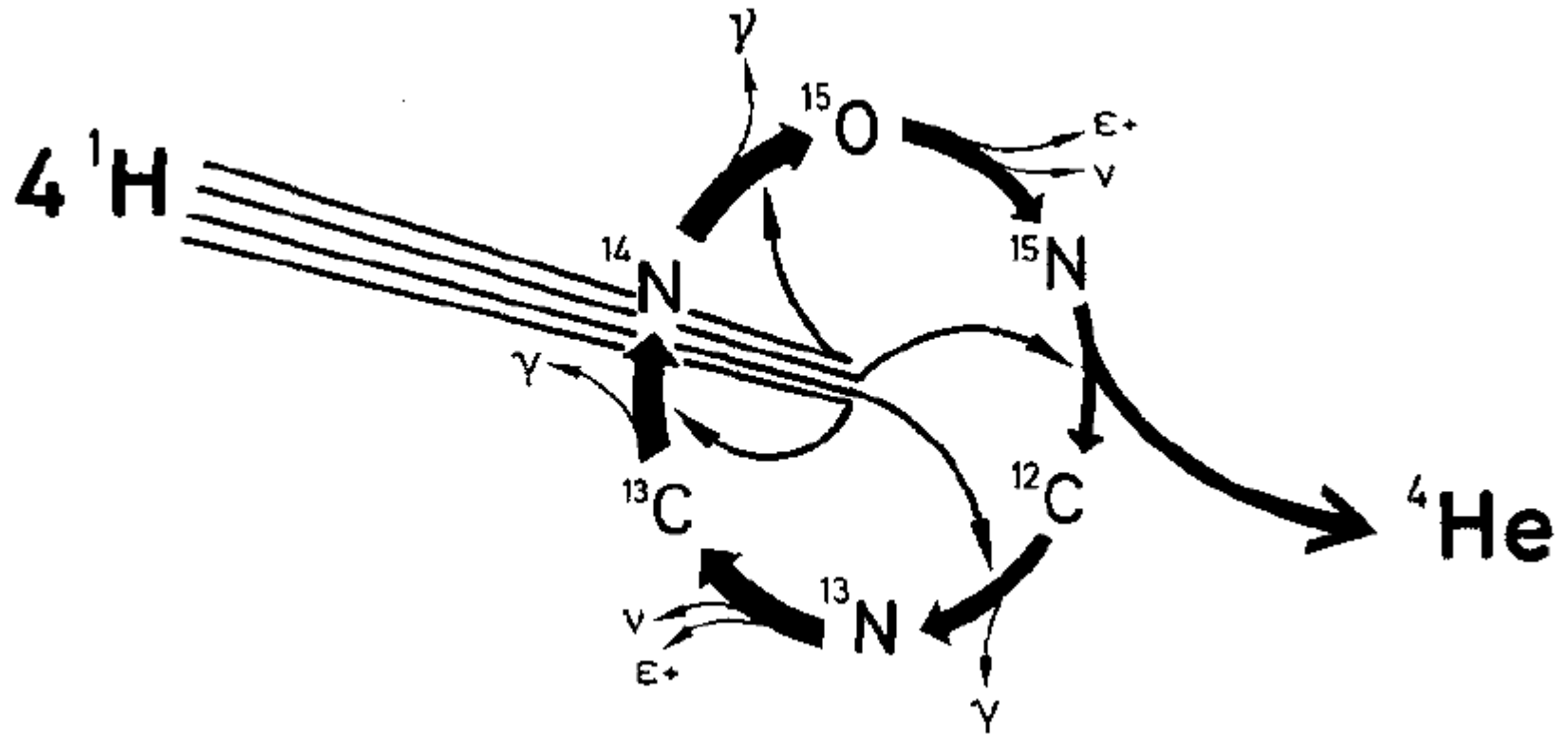
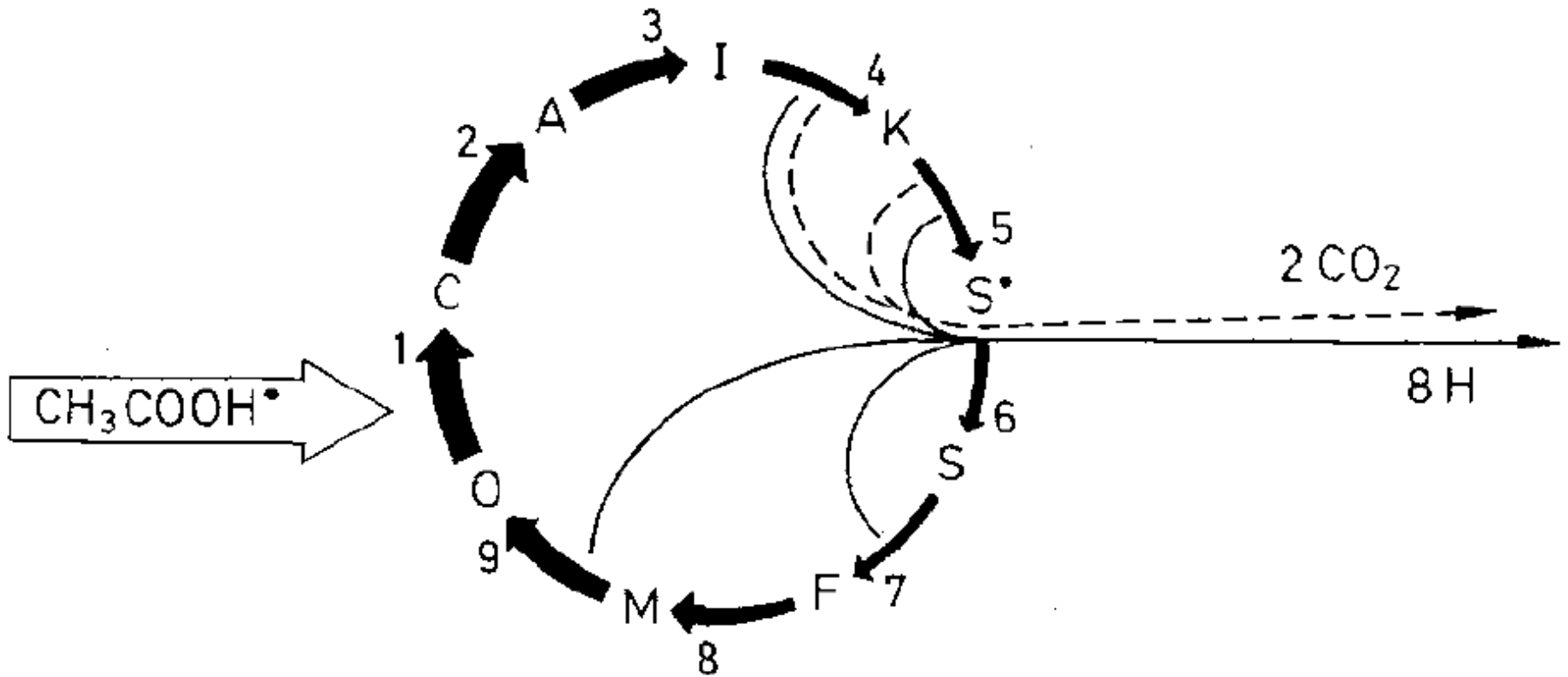


Fig. 1. *The common catalytic mechanism of an enzyme according to Michaelis and Menten involves (at least) three intermediates: the free enzyme (E), the enzyme-substrate (ES) and the enzyme-product complex (EP). The scheme demonstrates the equivalence of catalytic action of the enzyme and cyclic restoration of the intermediates in the turnover of the substrate (S) to the product (P). Yet, it provides only a formal representation of the true mechanism which may involve a stepwise activation of the substrate as well as induced conformation changes of the enzyme.*

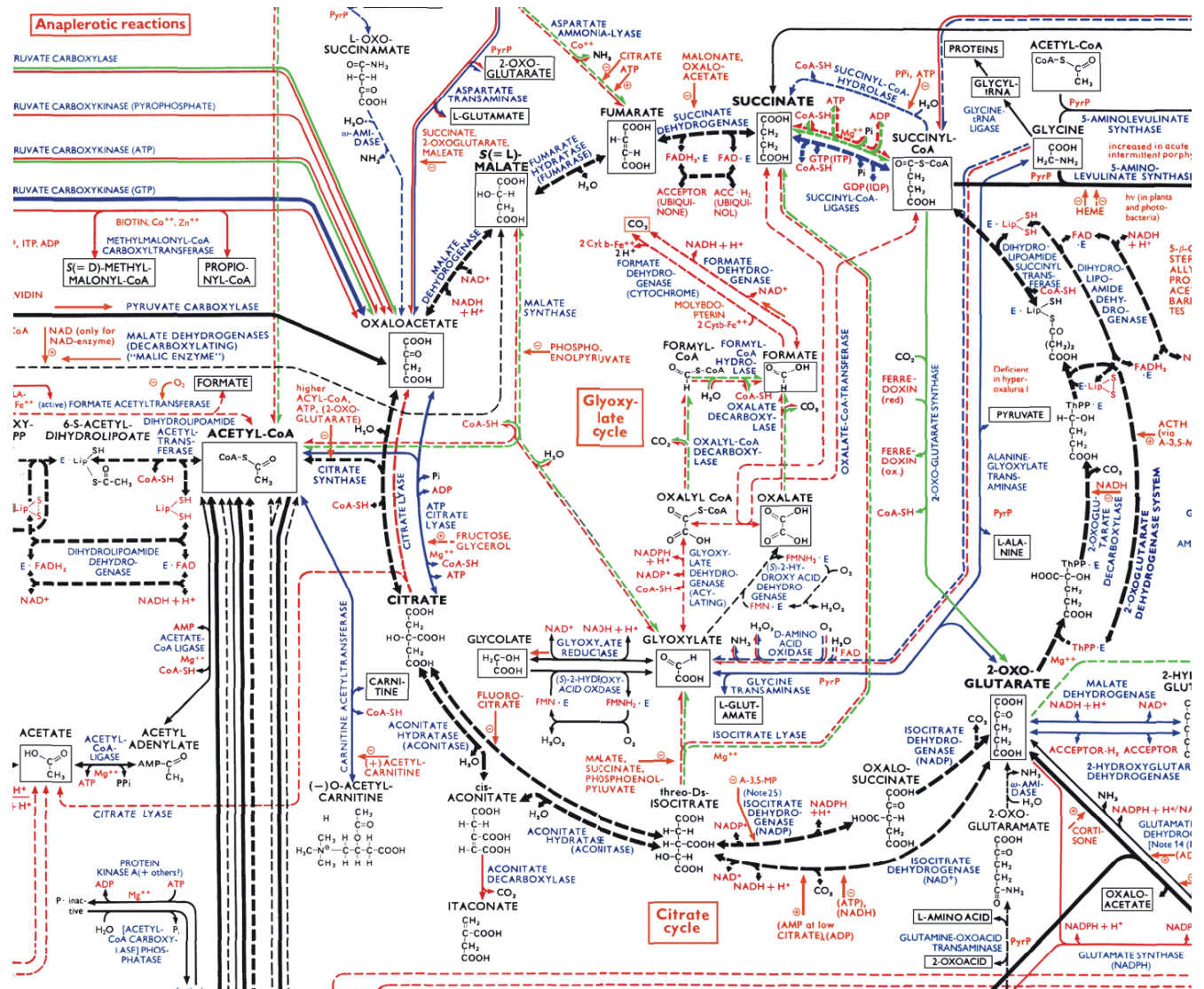


The Bethe - vonWeizsäcker catalytic cycle ist responsible
- in part - for the energy production in massive stars.

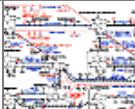
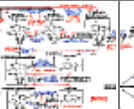
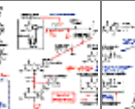
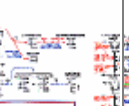
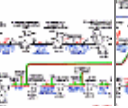
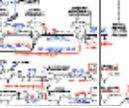
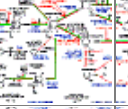
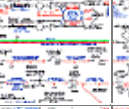
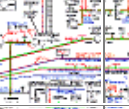
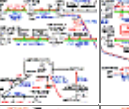
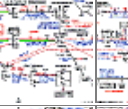
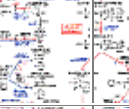
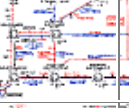
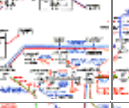
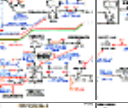
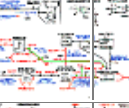
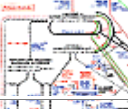
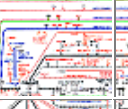
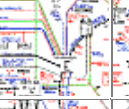
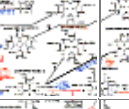
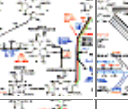
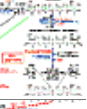
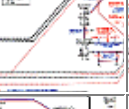
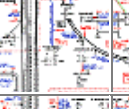
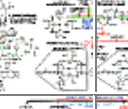
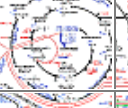
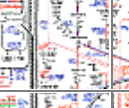
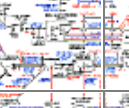
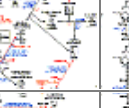
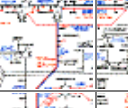
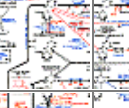
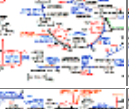
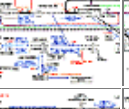
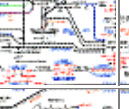
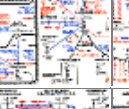
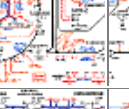
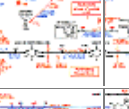
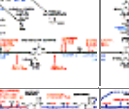
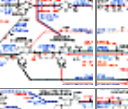
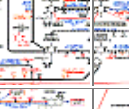
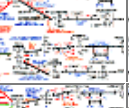
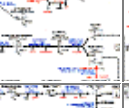
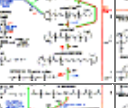
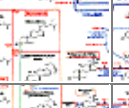
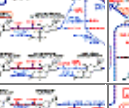
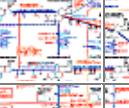
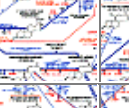
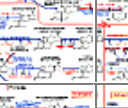
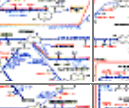
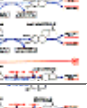
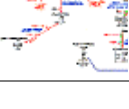
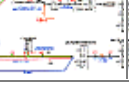
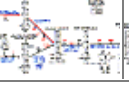
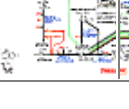
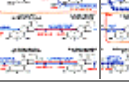
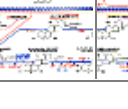
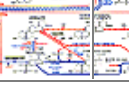


The tricarboxylic acid or citric acid cycle is fuelling the metabolic reactions of the cell.

The citric acid
or Krebs cycle
(enlarged)



The reaction network of cellular metabolism published by Boehringer-Mannheim.

	A	B	C	D	E	F	G	H	I	J	K	L
1	Biochemical Pathways											
2												
3												
4												
5												
6												
7												
8												
9												
10												

The reaction network of cellular metabolism published by Boehringer-Mannheim.

1. Cyclic reaction networks " catalysts
2. **Cyclic catalytic networks ' autocatalysts**
3. Cyclic autocatalytic networks " hypercycles
4. Neutrality - a source for coexistent competitors

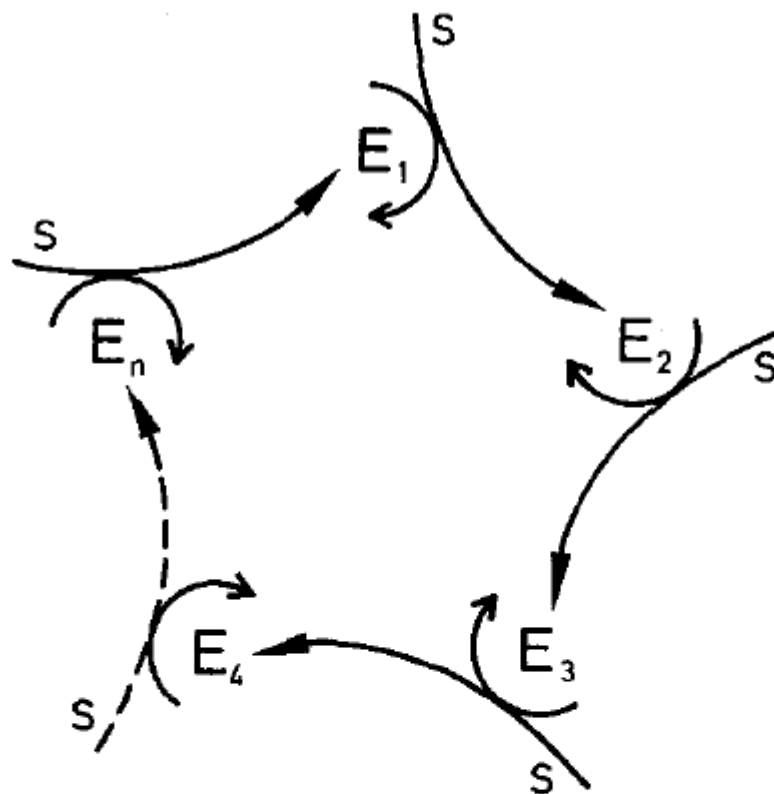
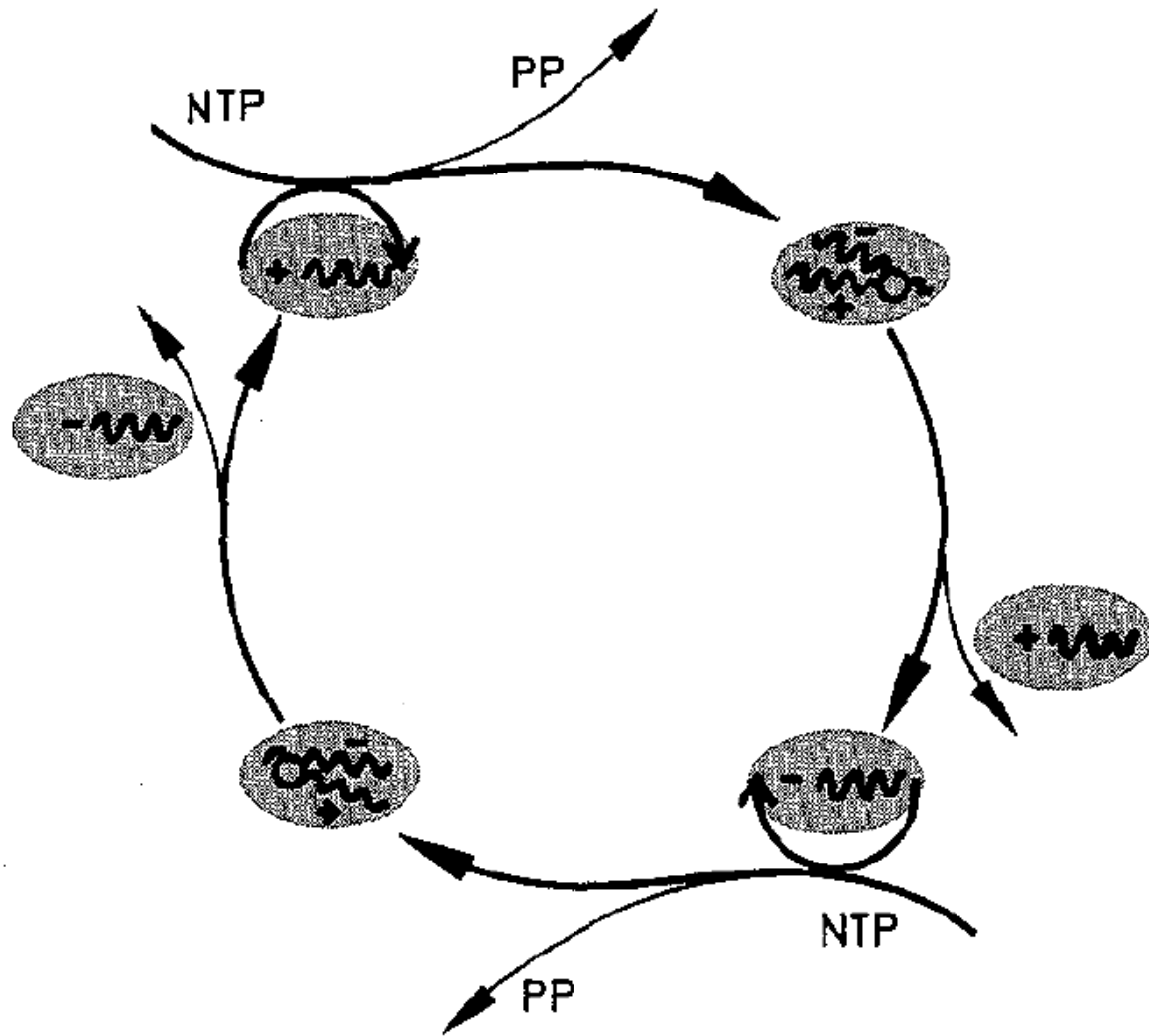
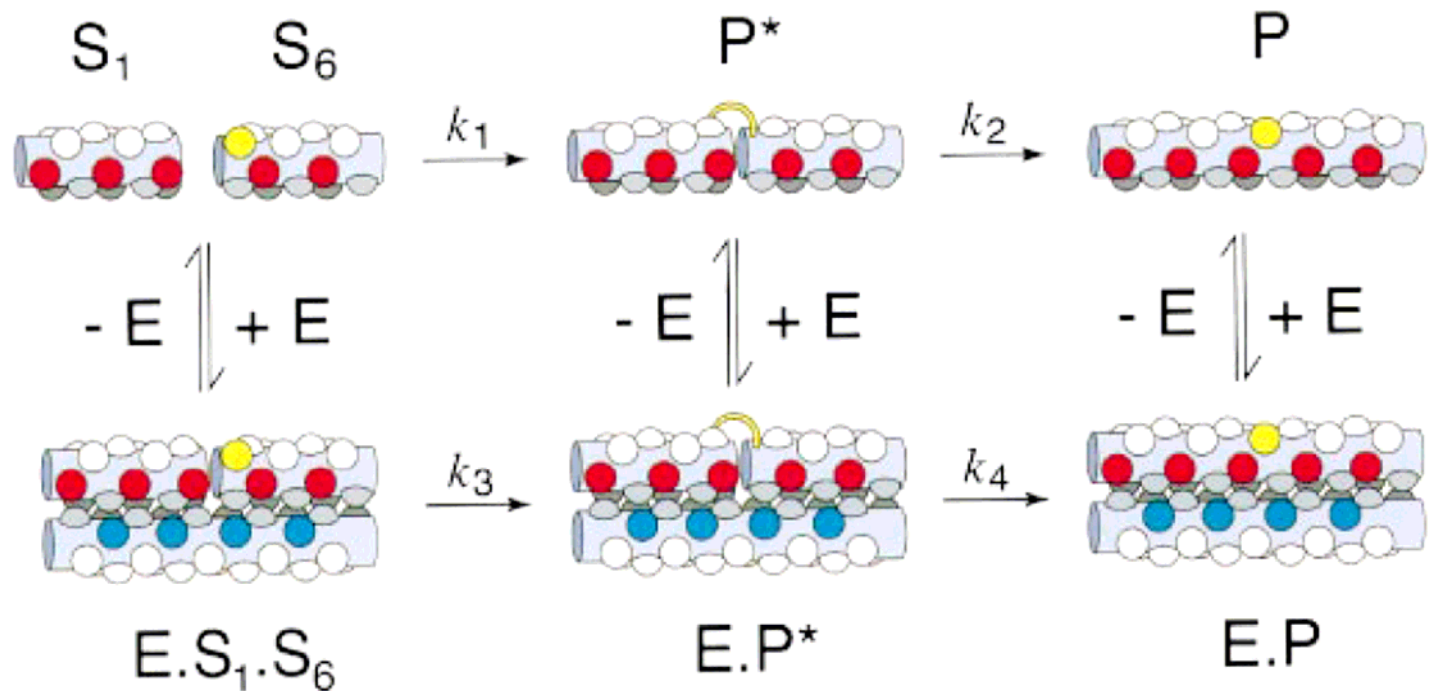


Fig. 4. *The catalytic cycle* represents a higher level of organization in the hierarchy of catalytic schemes. The constituents of the cycle $E_1 \rightarrow E_n$ are themselves catalysts which are formed from some energy-rich substrates (S), whereby each intermediate E_i is a catalyst for the formation of E_{i+1} . The catalytic cycle seen as an entity is equivalent to an autocatalyst, which instructs its own reproduction. To be a catalytic cycle it is sufficient, that only one of the intermediates formed is a catalyst for one of the subsequent reaction steps.



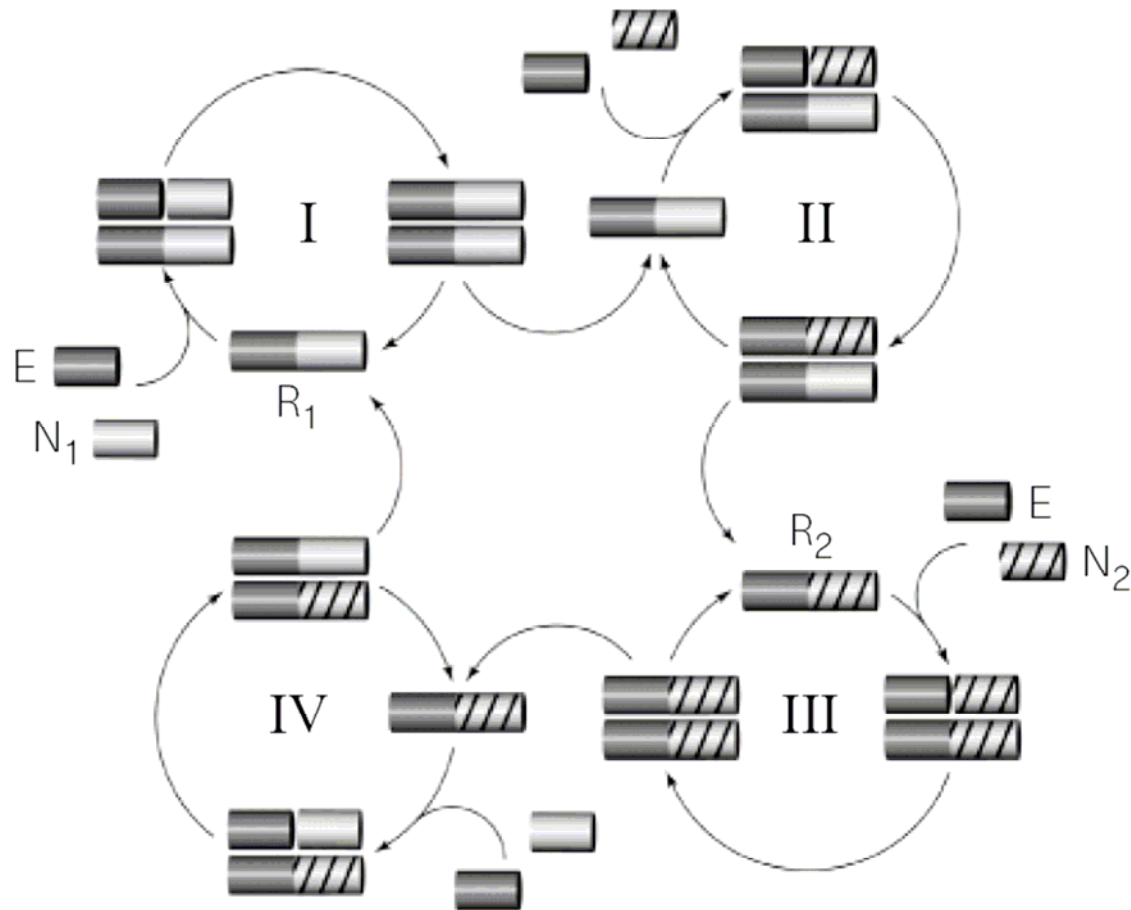
Complementary ($\pm$) replication of RNA as an example of an autocatalytic cycle.



A synthetic oligopeptide ligase
becomes a replicator for $E = P$

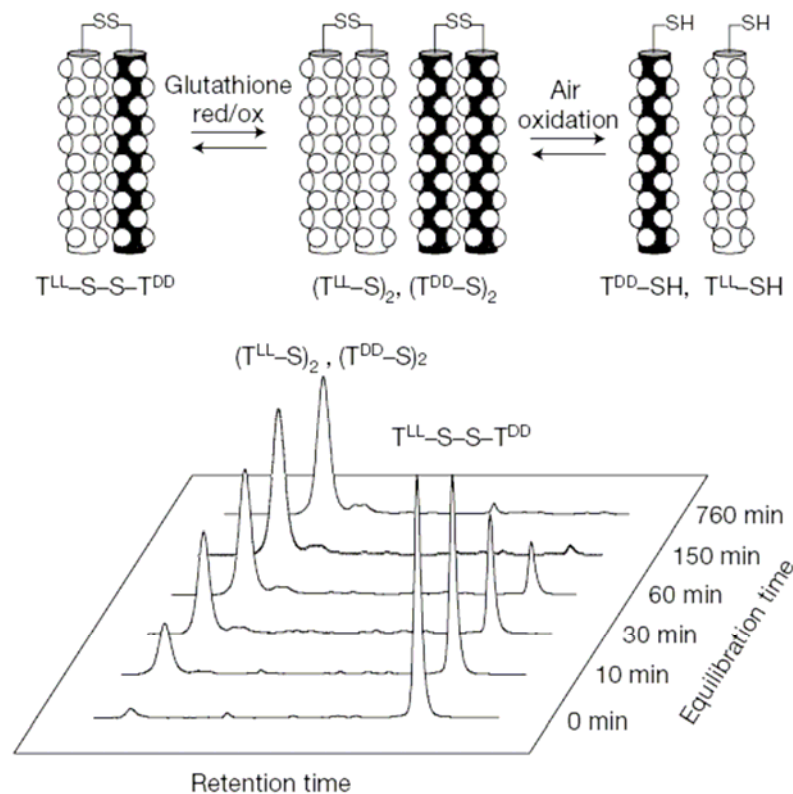
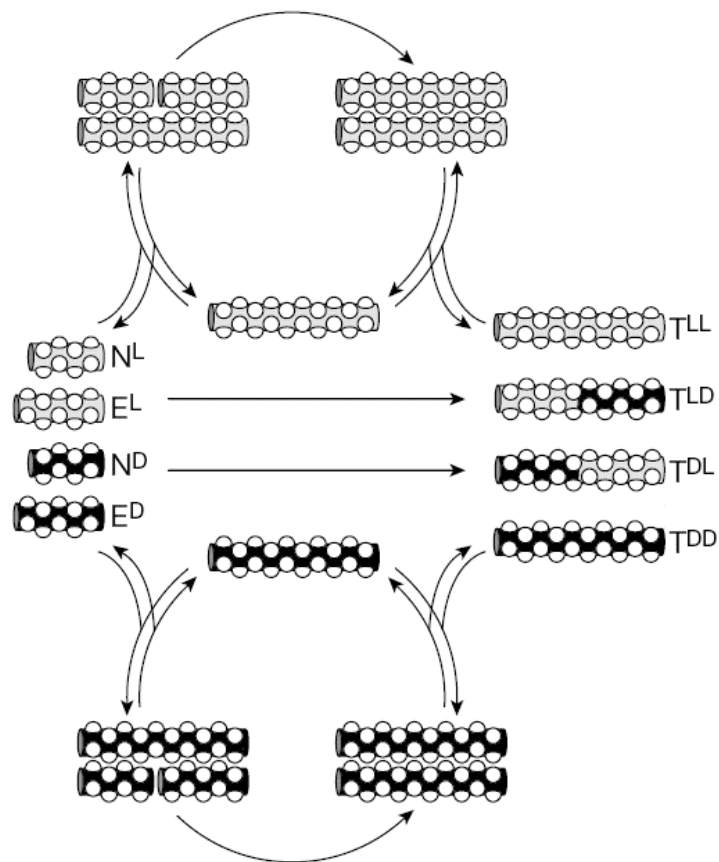
K. Severin, D.H. Lee, A.J. Kennan, M.R. Ghadiri, *Nature* **389**, 706-709, 1997

D.H. Lee, J.R. Granja, J.A. MartinezK. Severin, M.R. Ghadiri, *Nature* **382**, 525-528, 1996

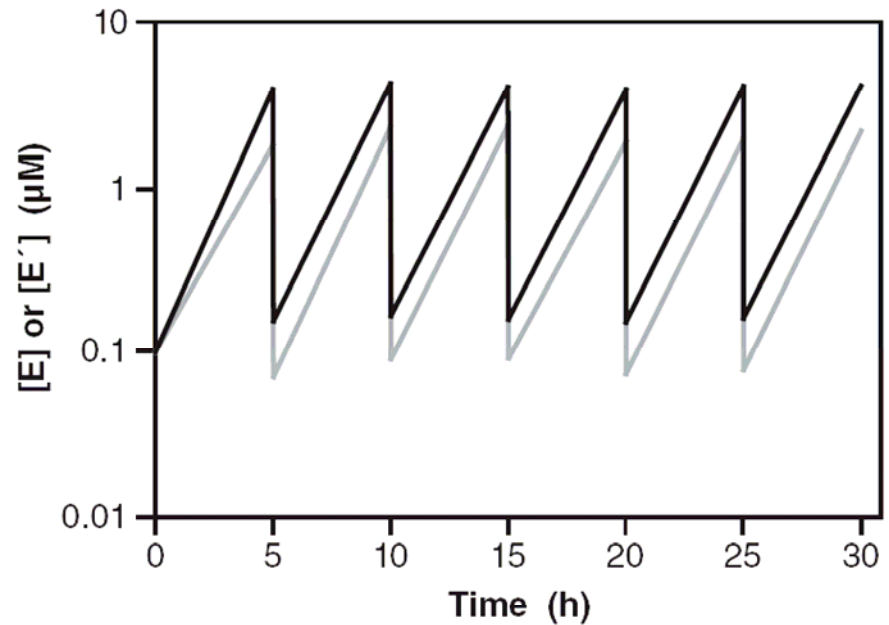
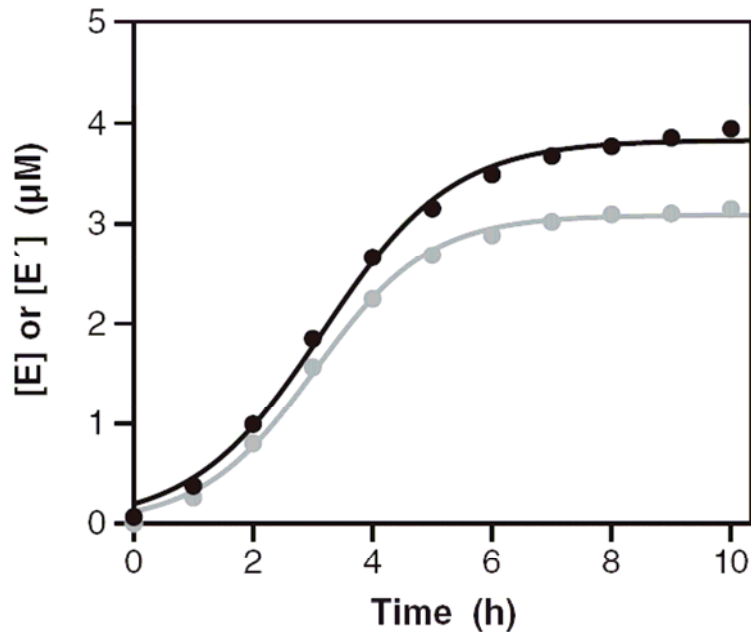


Cross-catalysis of peptide replicators

D.H. Lee, K. Severin, Y. Yokobayashi, M.R. Ghadiri, *Nature* **390**, 591-594, 1997

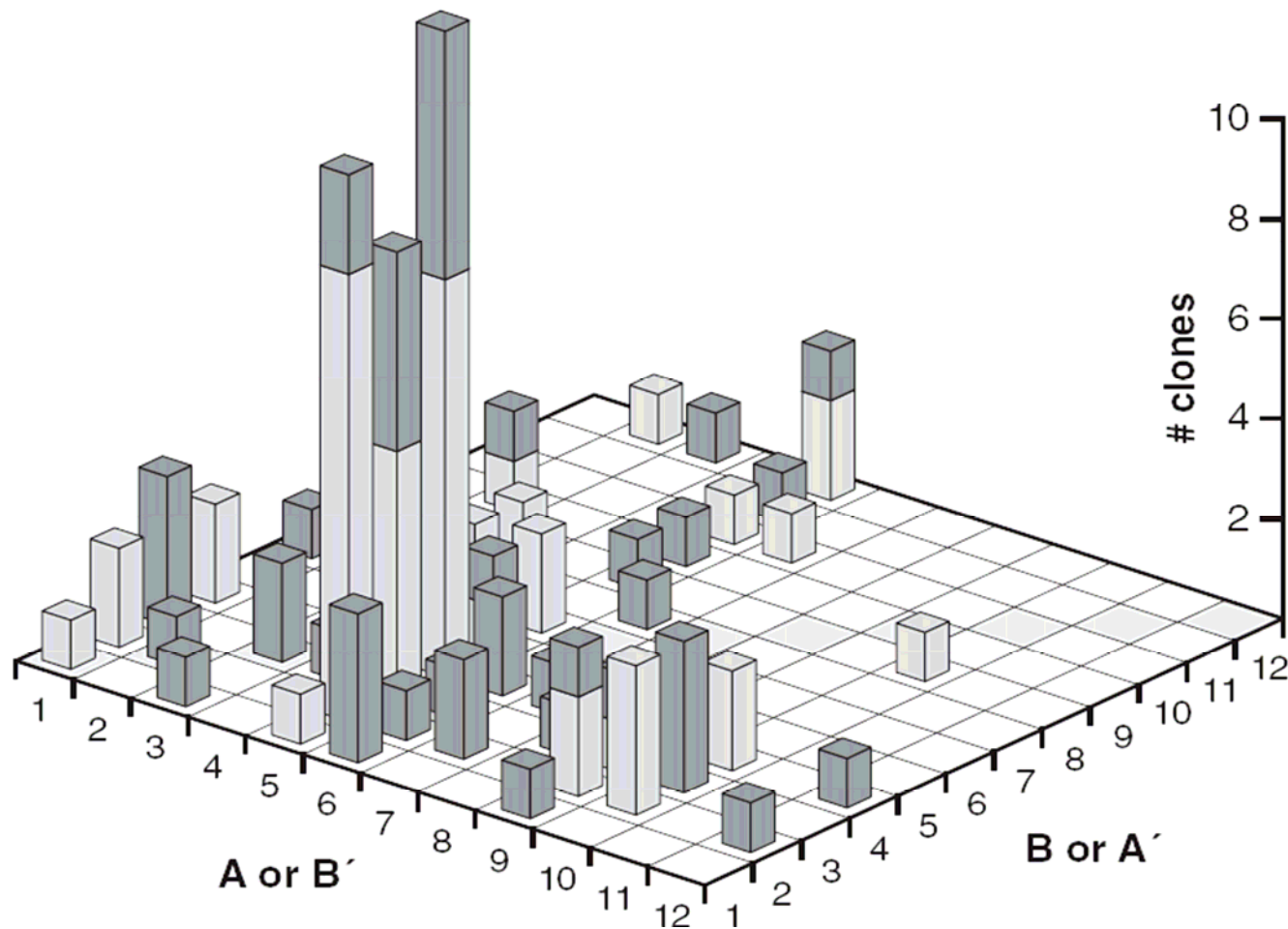


A chiroselective peptide replicator



Exponential growth levels off when the reservoir is exhausted (l.h.s.).

RNA production in serial transfer experiments (r.h.s.)



RNA evolution of recombinant replicators

Tracey A. Lincoln, Gerald F. Joyce, *Science* **323**, 1229-1232, 2009

1. Cyclic reaction networks " catalysts
2. Cyclic catalytic networks " autocatalysts
3. **Cyclic autocatalytic networks ´ hypercycles**
4. Neutrality - a source for coexistent competitors

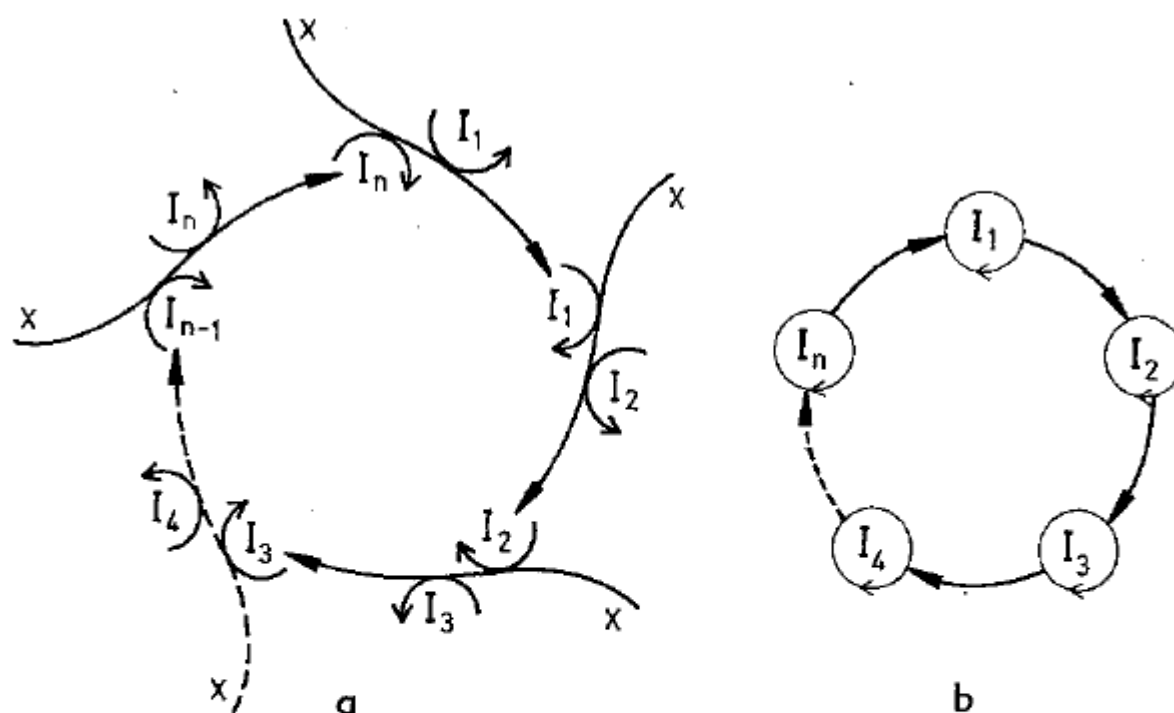
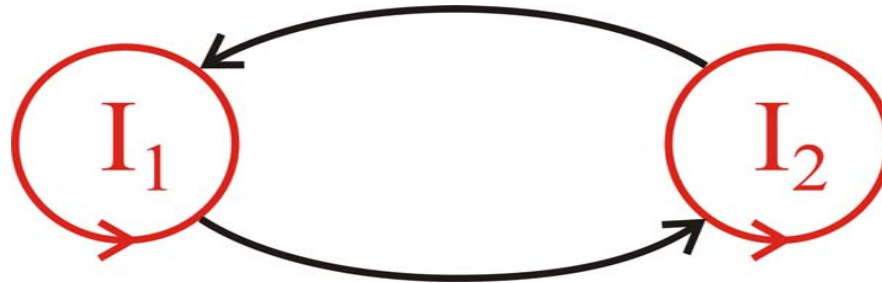
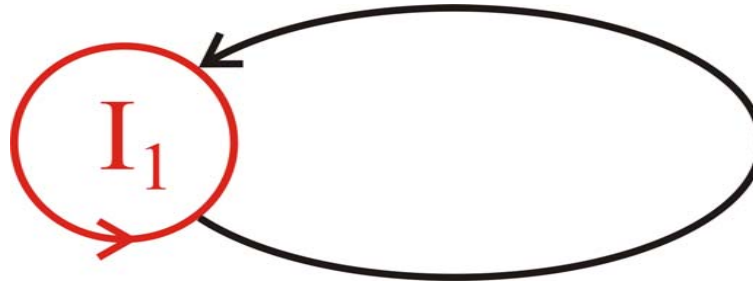
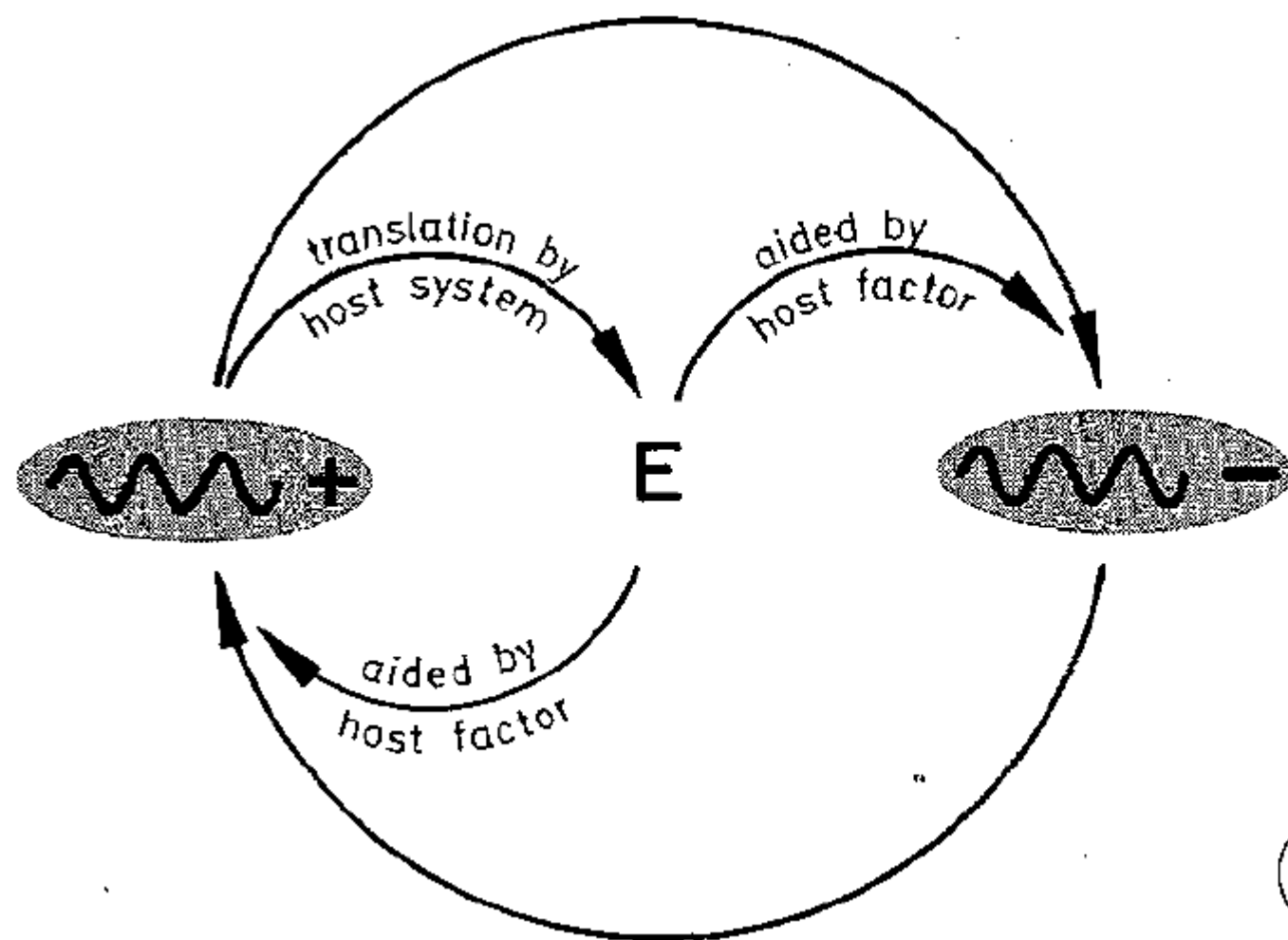
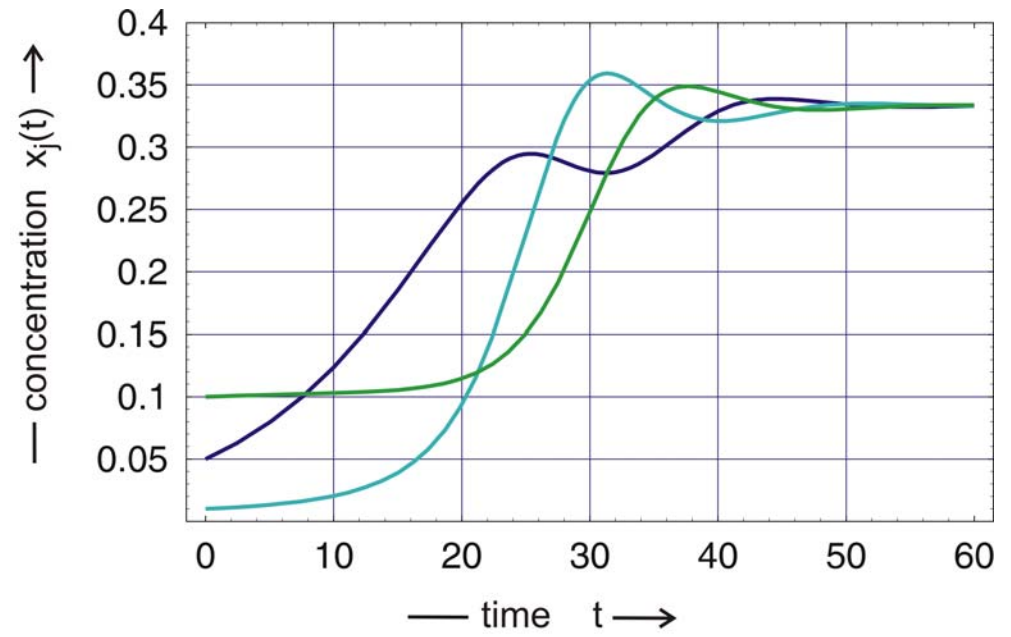
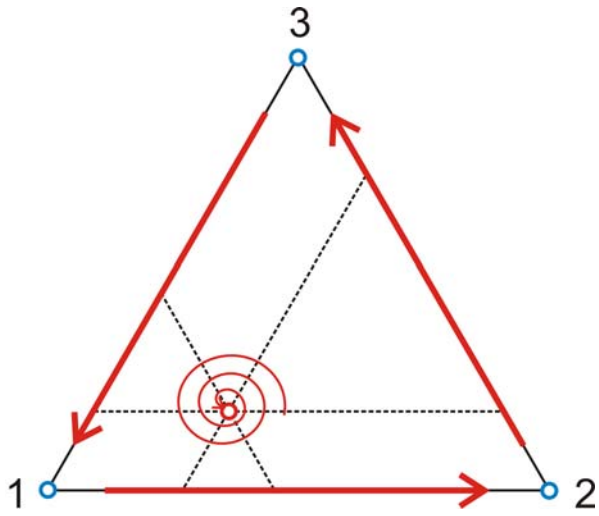


Fig. 7. A *catalytic hypercycle* consists of self-instructive units I_i with two-fold catalytic functions. As autocatalysts or—more generally—as catalytic cycles the intermediates I_i are able to instruct their own reproduction and, in addition, provide catalytic support for the reproduction of the subsequent intermediate (using the energy-rich building material X). The simplified graph (b) indicates the cyclic hierarchy

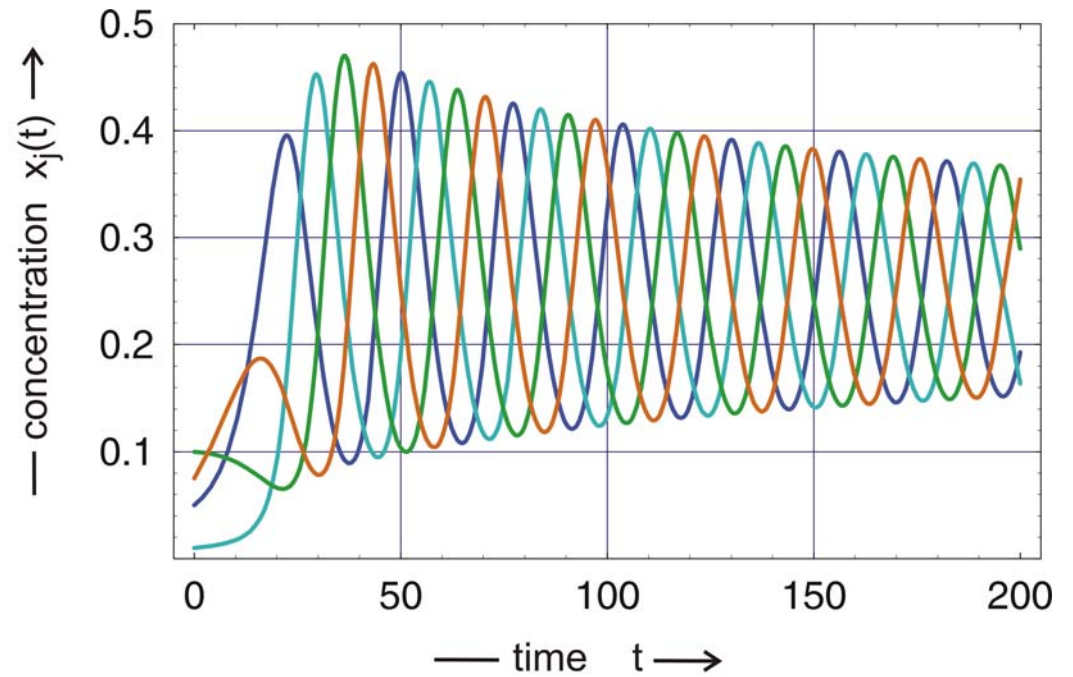
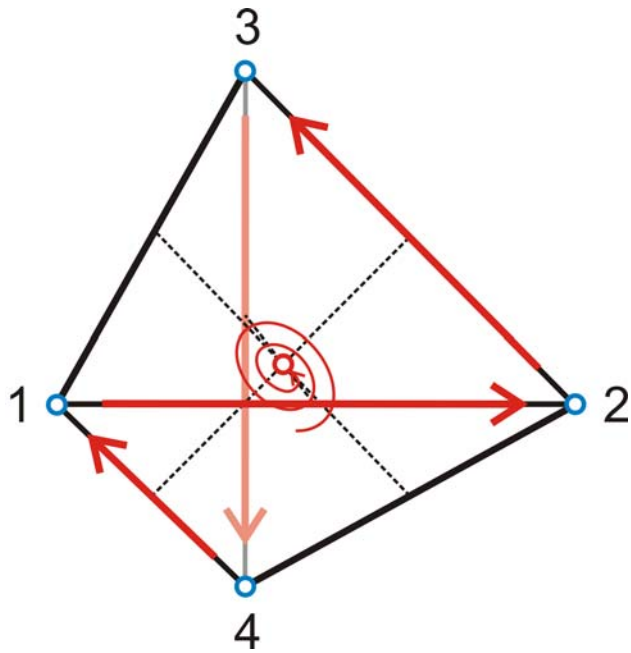


Hypercycles with one and two members are common in nature.

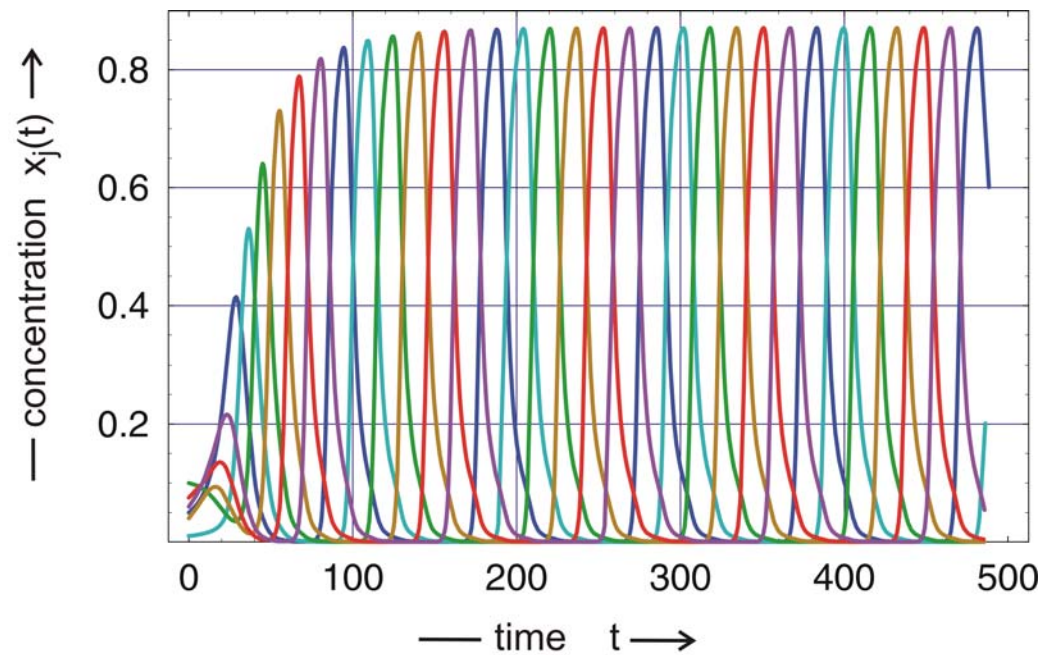
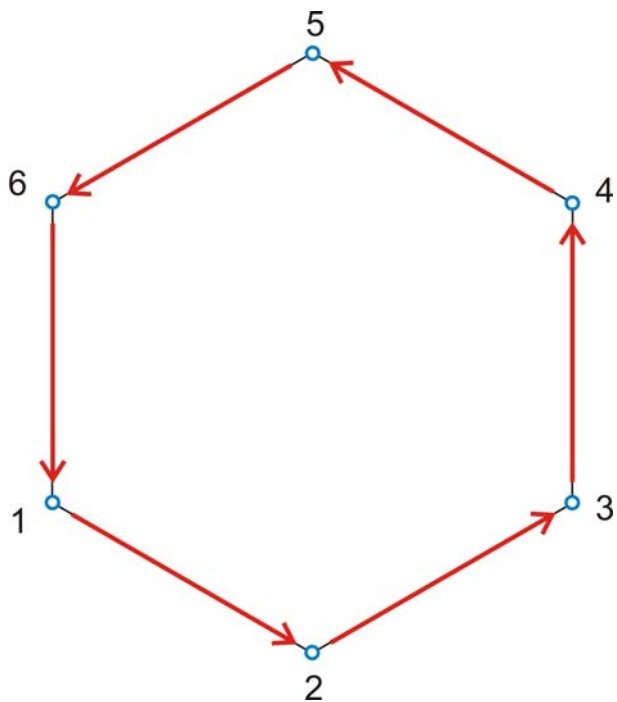




Hypercycle dynamics for $n=3$

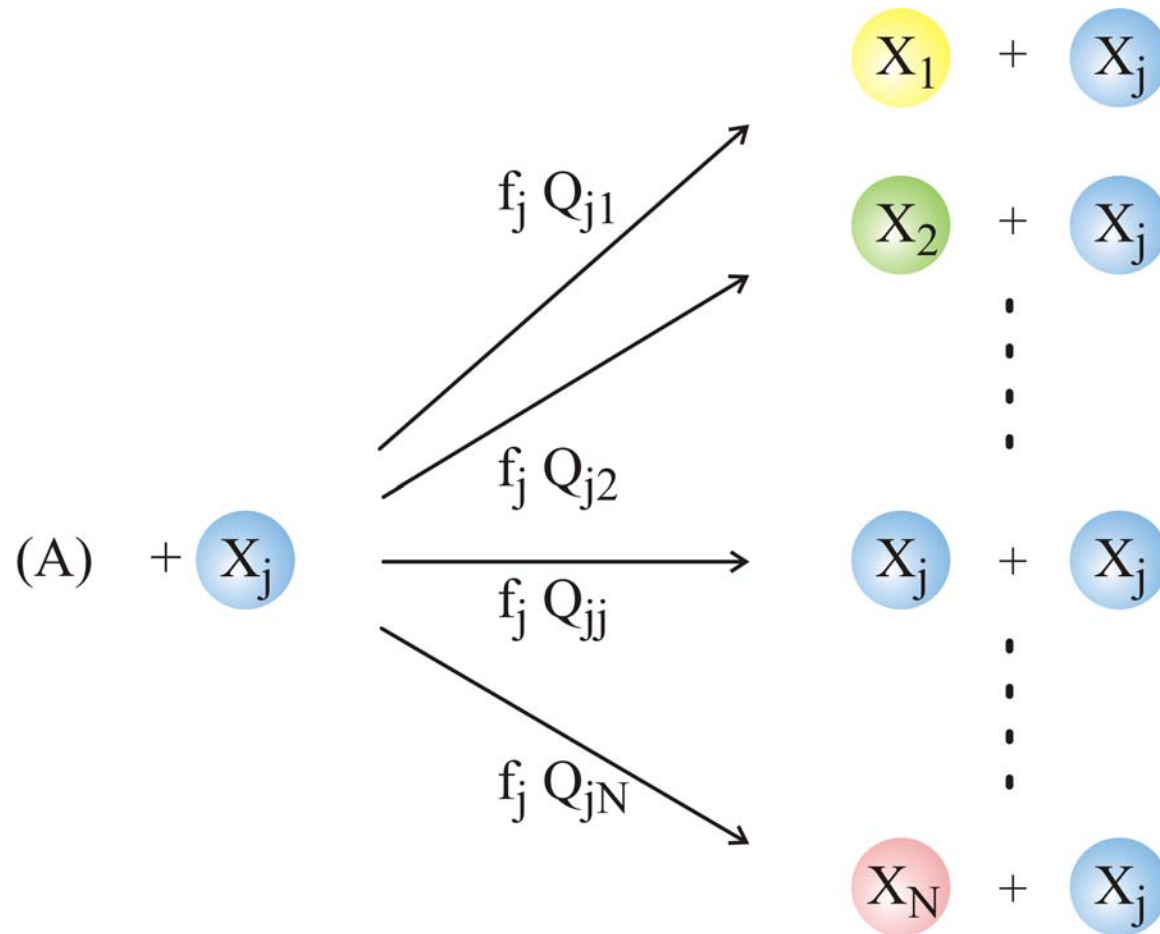


Hypercycle dynamics for $n=4$

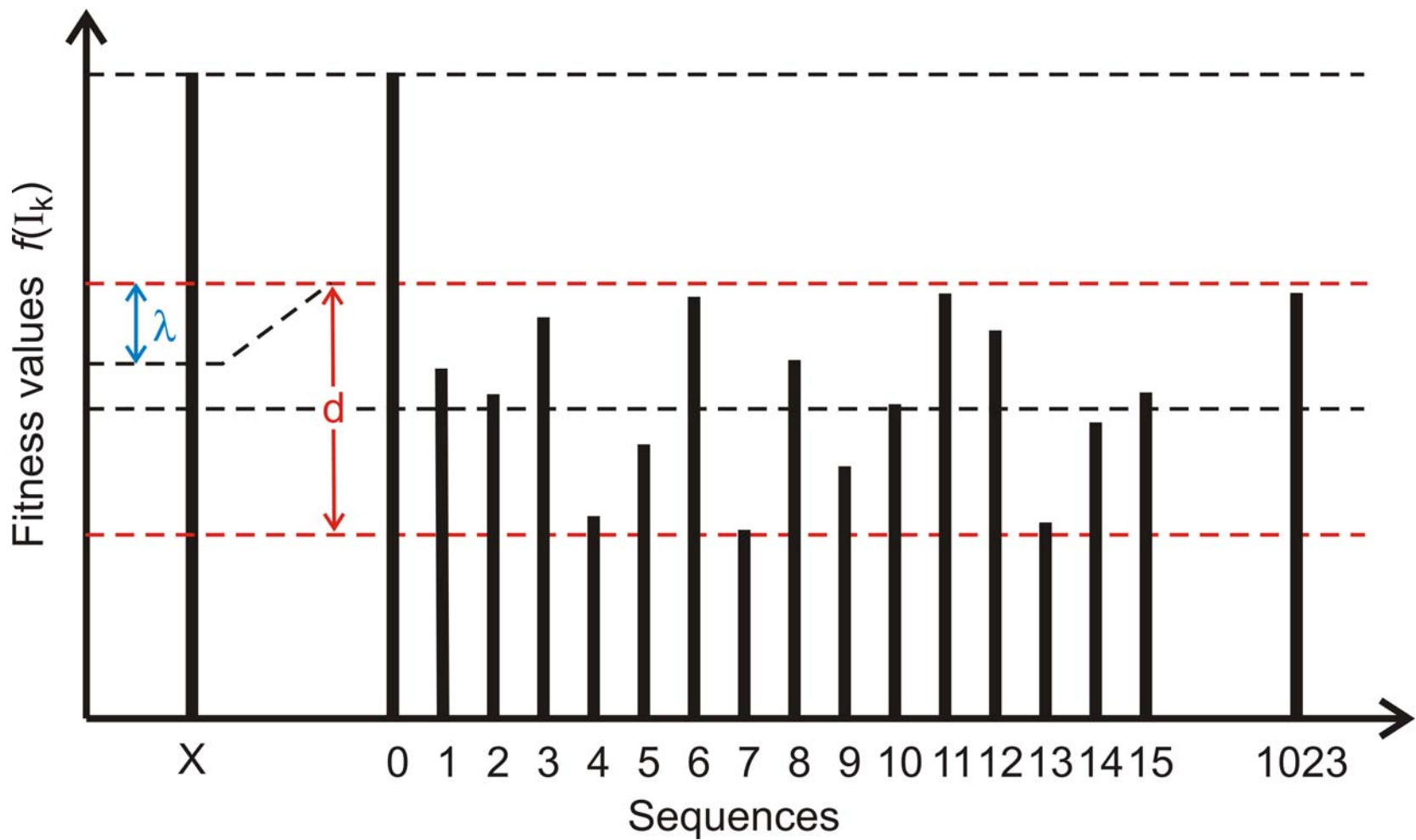


Hypercycle dynamics for $n=6$

1. Cyclic reaction networks " catalysts
2. Cyclic catalytic networks " autocatalysts
3. Cyclic autocatalytic networks " hypercycles
4. **Neutrality - a source for coexistent competitors**

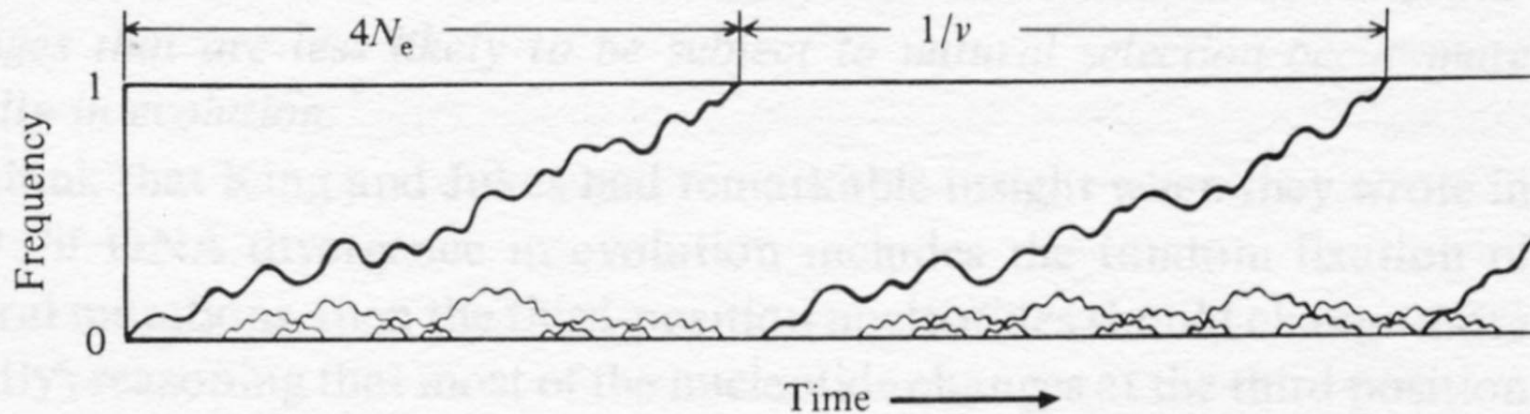


Chemical kinetics of replication and mutation as parallel reactions



A fitness landscape including neutrality

Fig. 3.1. Behavior of mutant genes following their appearance in a finite population. Courses of change in the frequencies of mutants destined to fixation are depicted by thick paths. N_e stands for the effective population size and v is the mutation rate.



Motoo Kimura

Is the Kimura scenario correct for frequent mutations?

STATIONARY MUTANT DISTRIBUTIONS AND EVOLUTIONARY OPTIMIZATION

■ PETER SCHUSTER and JÖRG SWETINA
Institut für theoretische Chemie
und Strahlenchemie der Universität Wien,
Währingerstraße 17,
A 1090 Wien,
Austria

Molecular evolution is modelled by erroneous replication of binary sequences. We show how the selection of two species of equal or almost equal selective value is influenced by its nearest neighbours in sequence space. In the case of perfect neutrality and sufficiently small error rates we find that the Hamming distance between the species determines selection. As the error rate increases the fitness parameters of neighbouring species become more and more important. In the case of almost neutral sequences we observe a critical replication accuracy at which a drastic change in the "quasispecies", in the stationary mutant distribution occurs. Thus, in frequently mutating populations fitness turns out to be an ensemble property rather than an attribute of the individual.

In addition we investigate the time dependence of the mean excess production as a function of initial conditions. Although it is optimized under most conditions, cases can be found which are characterized by decrease or non-monotonous change in mean excess productions.

1. Introduction. Recent data from populations of RNA viruses provided direct evidence for vast sequence heterogeneity (Domingo *et al.*, 1987). The origin of this diversity is not yet completely known. It may be caused by the low replication accuracy of the polymerizing enzyme, commonly a virus specific, RNA dependent RNA synthetase, or it may be the result of a high degree of selective neutrality of polynucleotide sequences. Eventually, both factors contribute to the heterogeneity observed. Indeed, mutations occur much more frequently than previously assumed in microbiology. They are by no means rare events and hence, neither the methods of conventional population genetics (Ewens, 1979) nor the neutral theory (Kimura, 1983) can be applied to these virus populations. Selectively neutral variants may be close with respect to Hamming distance and then the commonly made assumption that the mutation backflow from the mutants to the wilde type is negligible does not apply.

A kinetic theory of polynucleotide evolution which was developed during the past 15 years (Eigen, 1971; 1985; Eigen and Schuster, 1979; Eigen *et al.*, 1987; Schuster, 1986); Schuster and Sigmund, 1985) treats correct replication and mutation as parallel reactions within one and the same reaction network

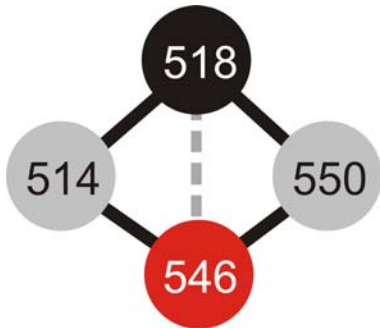


Neutral network

$\lambda = 0.01, s = 367$

$$d_H = 1$$

$$\lim_{p \rightarrow 0} x_1(p) = x_2(p) = 0.5$$



Neutral network

$\lambda = 0.01, s = 877$

$$d_H = 2$$

$$\lim_{p \rightarrow 0} x_1(p) = a$$

$$\lim_{p \rightarrow 0} x_2(p) = 1 - a$$

$$d_H \geq 3$$

$$\lim_{p \rightarrow 0} x_1(p) = 1, \lim_{p \rightarrow 0} x_2(p) = 0 \quad \text{or}$$

$$\lim_{p \rightarrow 0} x_1(p) = 0, \lim_{p \rightarrow 0} x_2(p) = 1$$

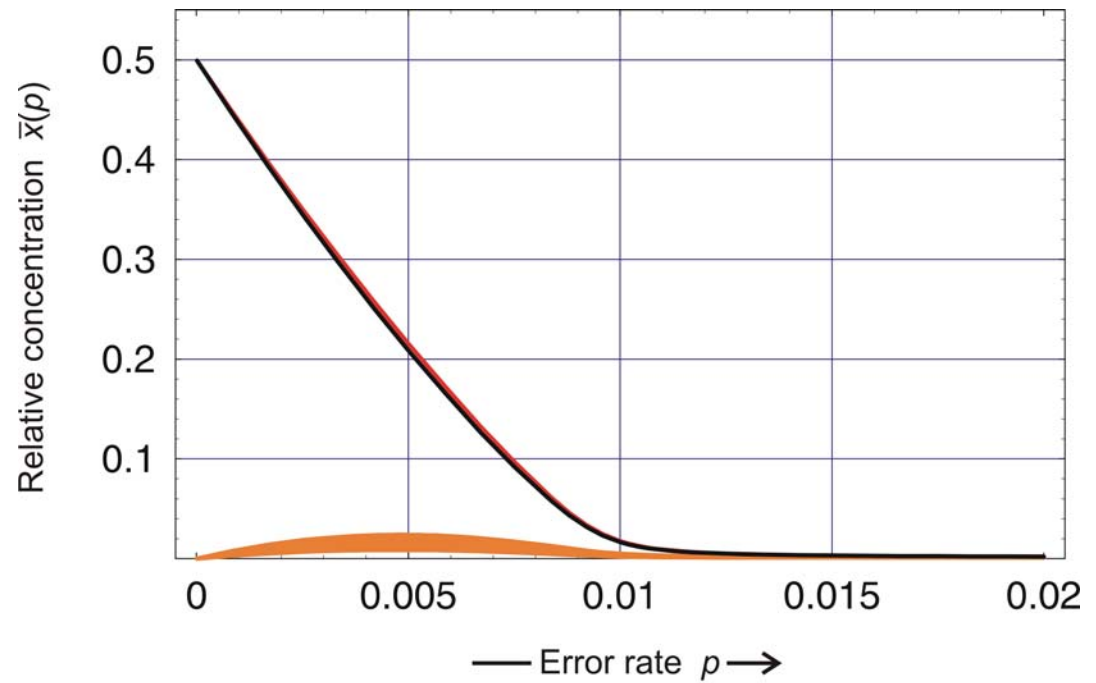
Pairs of genotypes in neutral replication networks

Random fixation in the
sense of Motoo Kimura



Neutral network

$\lambda = 0.01$, $s = 367$



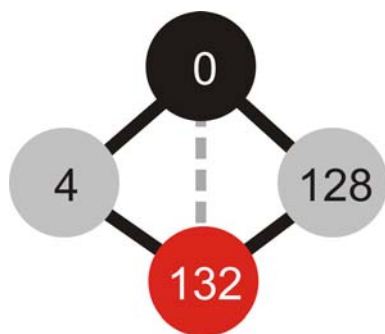
Neutral network: Individual sequences

$n = 10$, $\sigma = 1.1$, $d = 1.0$

..... ACAUGCGAA
 AUAUACGAA
 ACAUGCGCA
 GCAUACGAA
 ACAUGCUGAA
 ACAUGCGAG
 ACACGCGAA
 ACUGACGAA
 ACAUAGGAA
 ACAUACGAA

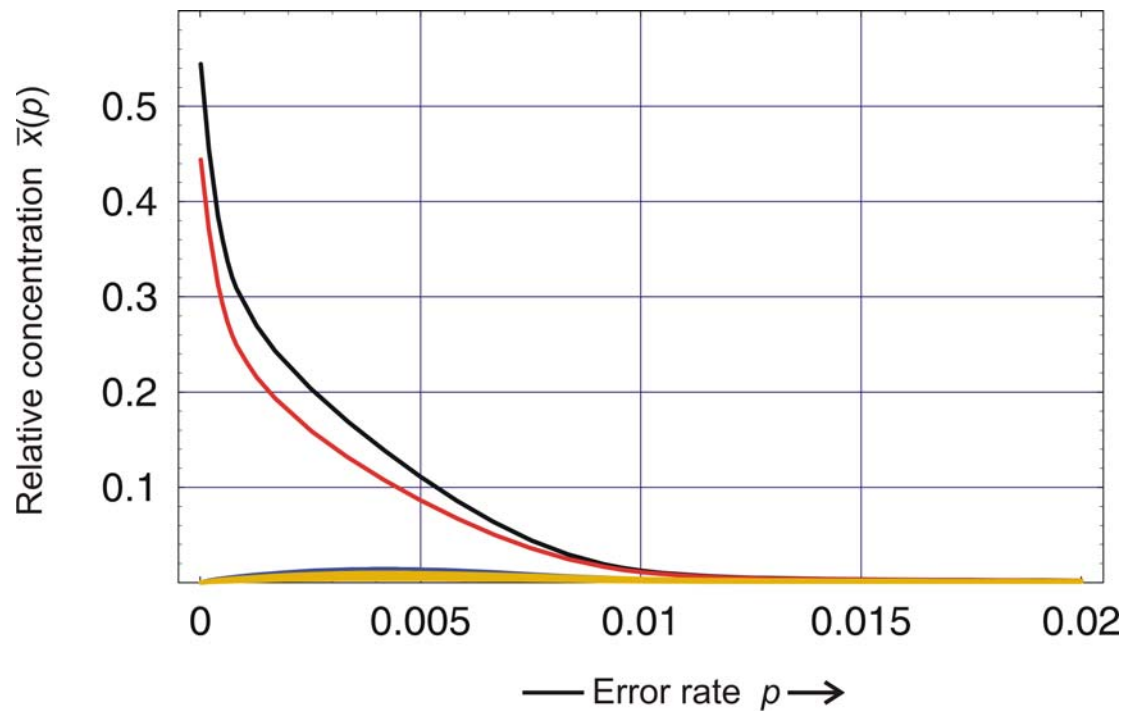
..... ACAU $\begin{matrix} G \\ A \end{matrix}$ CGAA

Consensus sequence of a quasispecies of two strongly coupled sequences of
 Hamming distance $d_H(X_i, X_j) = 1$.



Neutral network

$\lambda = 0.01$, $s = 877$



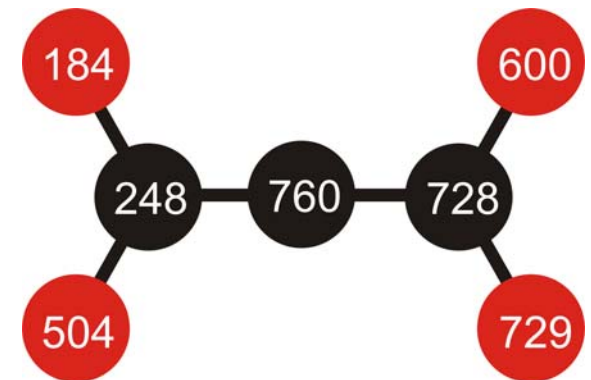
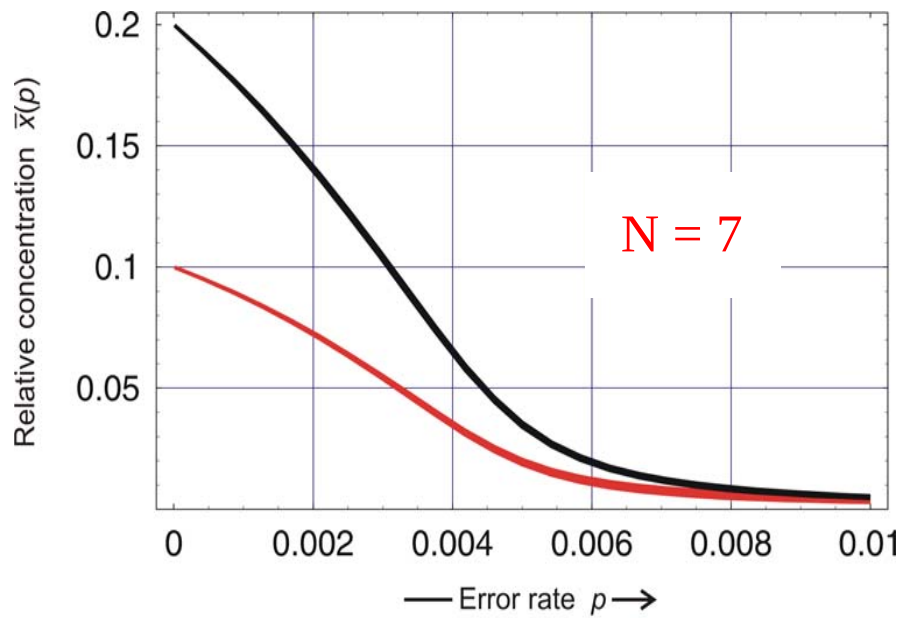
Neutral network: Individual sequences

$n = 10$, $\sigma = 1.1$, $d = 1.0$

..... ACAU**G**AU**U**CC**C**CGAA
 AU**A**UA**A**U**A**CC**U**CGAA
 ACAU**A**AU**U**CC**C**CG**A**
 **G**CAU**A**AU**U**U**U**C**U**CGAA
 ACAU**G**AU**U**CC**C**C**U**AA
 ACAU**A**A**G**U**C**CC**C**CG**A**G
 AC**A**C**G**AU**U**CC**C**CGAA
 AC**G**U**A**AU**U**CC**U**CGAA
 ACAU**G**C**U**U**C**C**U**A**G**AA
 ACAU**A**AU**U**CC**C**CGAA
 AU**A**UA**A**U**U**C**U**C**G**AA
 ACA**A**AU**G**CC**C**CG**U**A

..... ACAU^{**A**}_{**G**}AU**U**CC^{**C**}_{**U**}CGAA

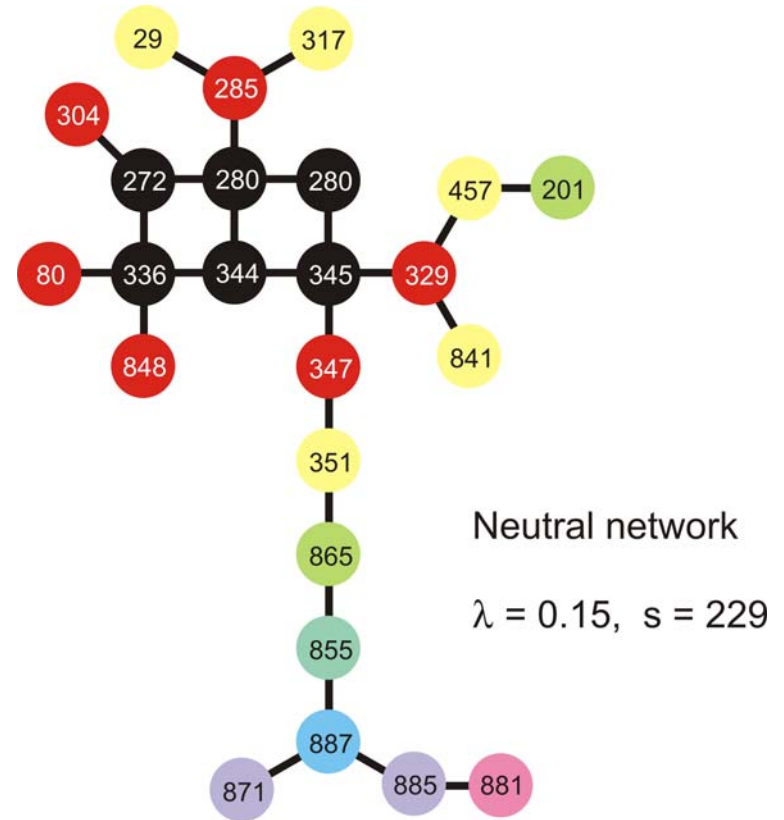
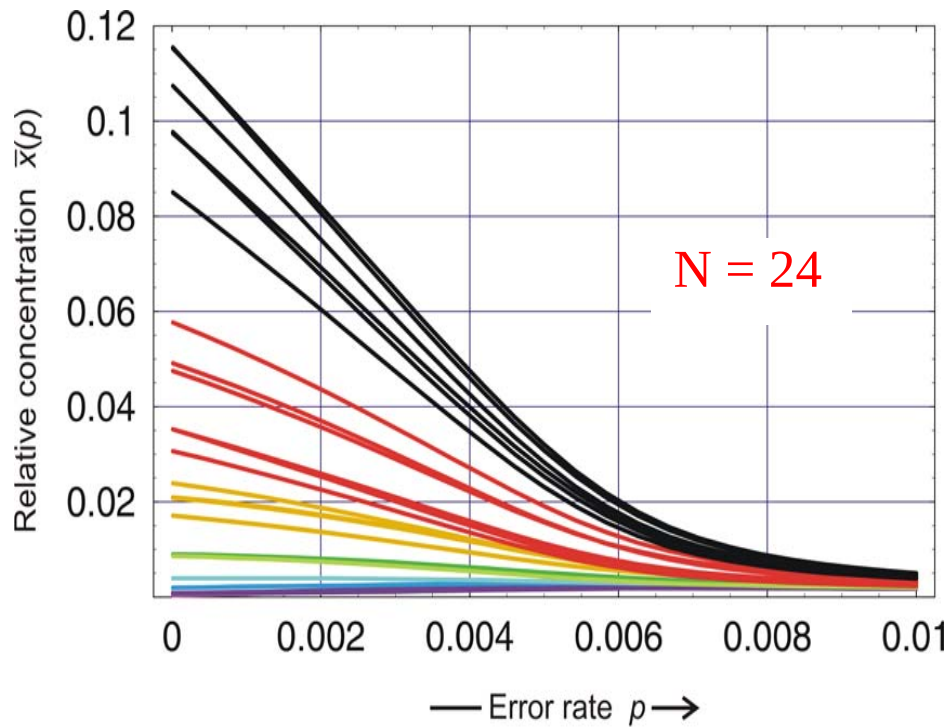
Consensus sequence of a quasispecies of two strongly coupled sequences of
 Hamming distance $d_H(X_i, X_j) = 2$.



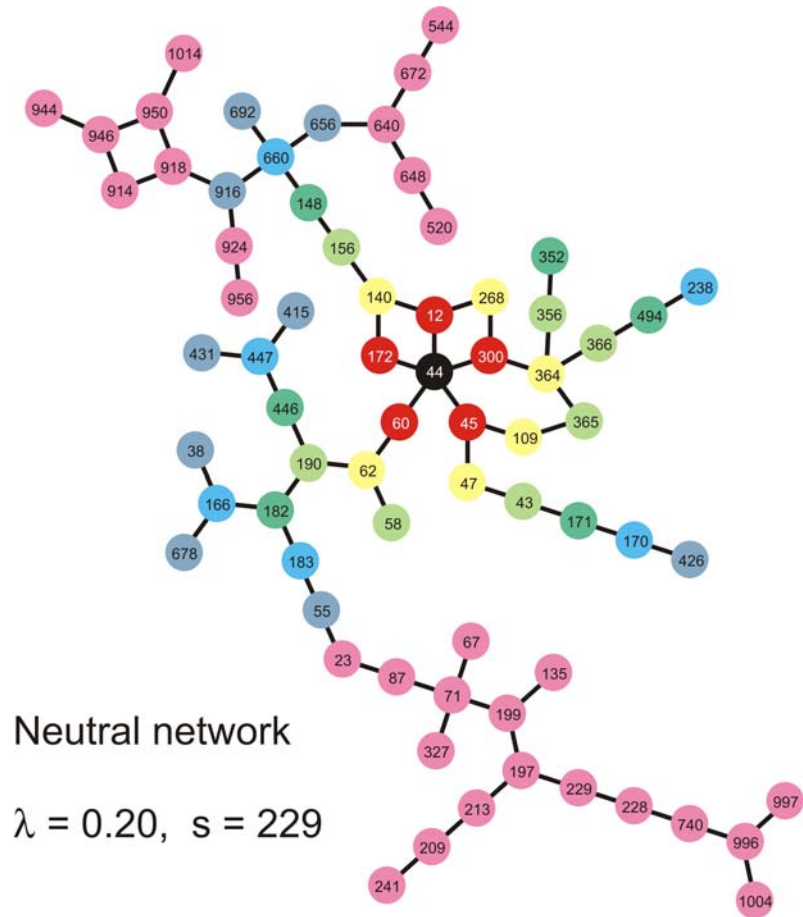
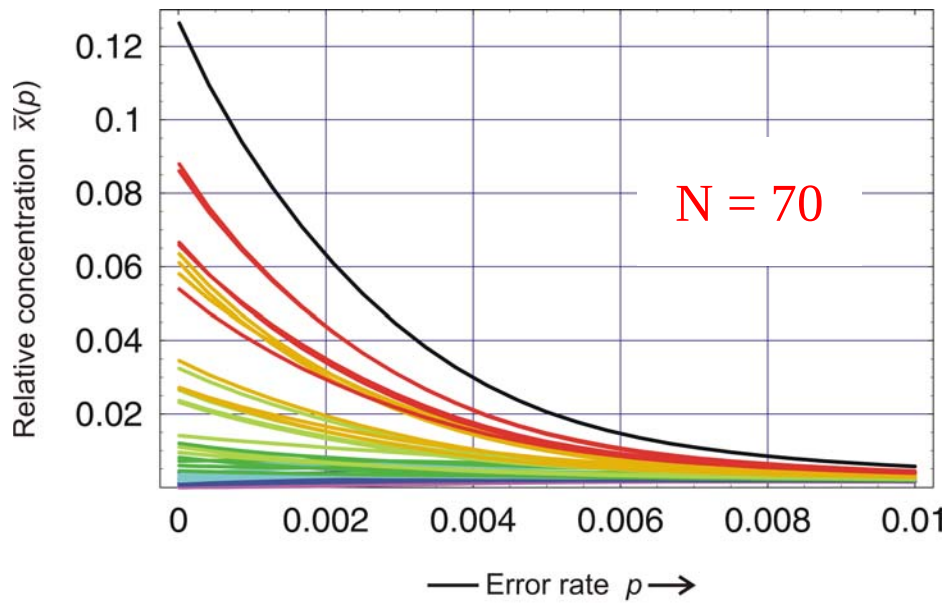
Neutral network

$\lambda = 0.10, s = 229$

Neutral networks with increasing λ : $\lambda = 0.10, s = 229$

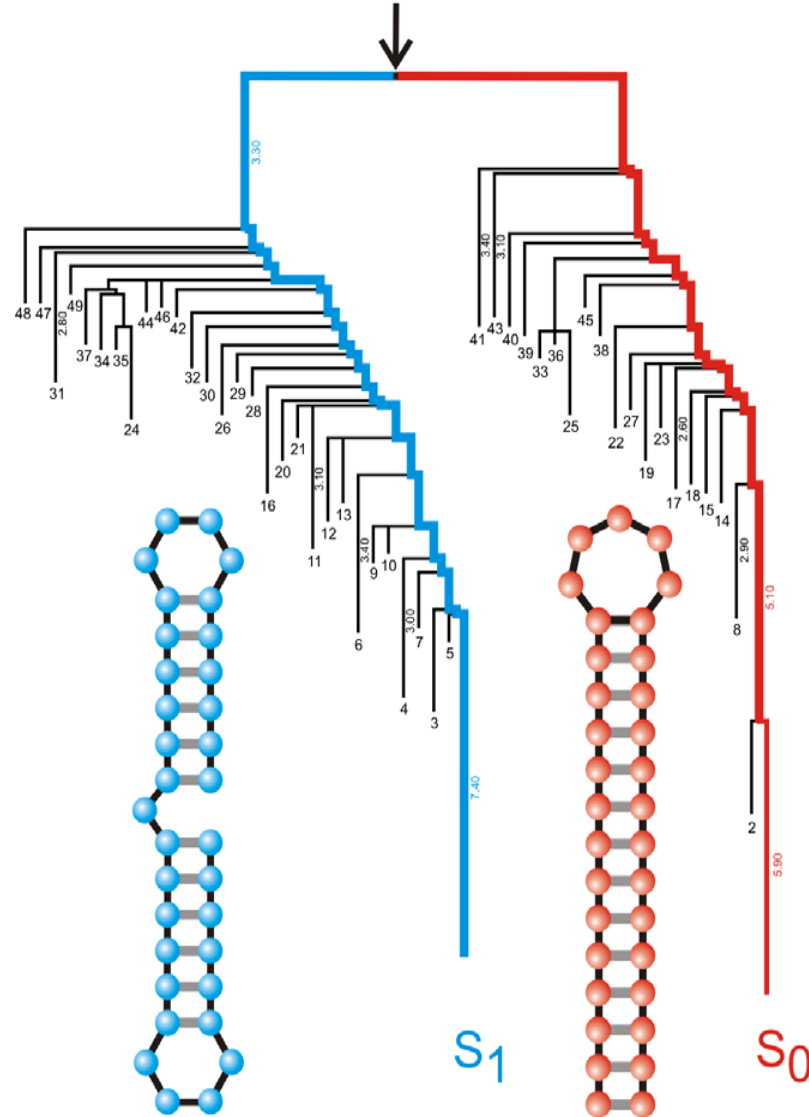


Neutral networks with increasing λ : $\lambda = 0.15$, $s = 229$



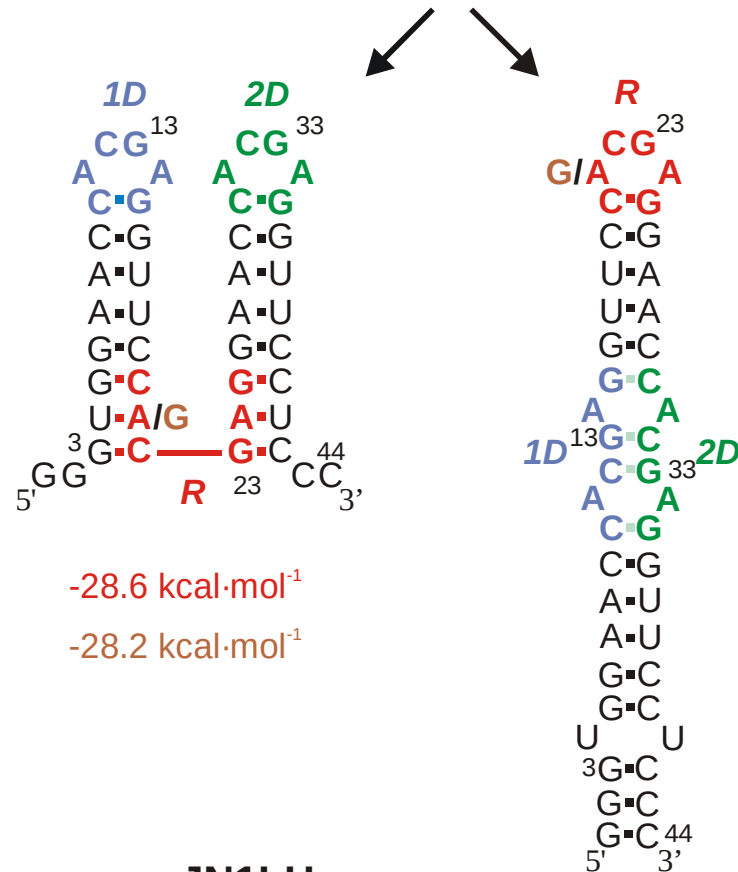
Neutral networks with increasing λ : $\lambda = 0.20, s = 229$

Metastable structures Conformational switches



Kinetic structures

$1D$ R $2D$
 GGGUGGAAC**CACGAG**GUUC**CACGAG**GAAC**CACGAG**GUUCCUCCC
 3 13 23 33 44



An RNA switch

JN1LH

J.H.A. Nagel, C. Flamm, I.L. Hofacker, K. Franke,
M.H. de Smit, P. Schuster, and C.W.A. Pleij.

Structural parameters affecting the kinetic competition of
RNA hairpin formation. *Nucleic Acids Res.* **34**:3568-3576,
2006.

A ribozyme switch

E.A.Schultes, D.B.Bartel, Science
289 (2000), 448-452

minus the background levels observed in the HSP in the control (Sar1-GDP-containing) incubation that prevents COPII vesicle formation. In the microsome control, the level of p115-SNARE associations was less than 0.1%.

46. C. M. Carr, E. Grote, M. Munson, F. M. Hughson, P. J. Novick, *J. Cell Biol.* **146**, 333 (1999).

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50. GST-SNARE proteins were expressed in bacteria and purified on glutathione-Sepharose beads using standard methods. Immobilized GST-SNARE protein (0.5 μ M) was incubated with rat liver cytosol (20 mg) or purified recombinant p115 (0.5 μ M) in 1 ml of NS buffer containing 1% BSA for 2 hours at 4°C with rotation. Beads were briefly spun (3000 rpm for 10 s) and sequentially washed three times with NS buffer and three times with NS buffer supplemented with 150 mM NaCl. Bound proteins were eluted three times in 50 μ l of 50 mM Tris-HCl (pH 8.5), 50 mM reduced glutathione, 150 mM NaCl, and 0.1% Triton

REPORTS

X-100 for 15 min at 4°C with intermittent mixing, and elutes were pooled. Proteins were precipitated by MeOH/CH₂Cl₂ and separated by SDS-polyacrylamide gel electrophoresis (PAGE) followed by immunoblotting using p115 mAb 13F12.

51. V. Rybin et al., *Nature* **383**, 266 (1996).

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62. M. R. Peterson, C. G. Burd, S. D. Emr, *Curr. Biol.* **9**, 159 (1999).

63. M. G. Waters, D. O. Clary, J. E. Rothman, *J. Cell Biol.* **118**, 1015 (1992).

64. D. M. Walter, K. S. Paul, M. G. Waters, *J. Biol. Chem.* **273**, 29565 (1998).

65. N. Hui et al., *Mol. Biol. Cell* **8**, 1777 (1997).

66. T. E. Kreis, *EMBO J.* **5**, 931 (1986).

67. H. Plutner, H. W. Davidson, J. Saraste, W. E. Balch, *J. Cell Biol.* **119**, 1097 (1992).

68. D. S. Nelson et al., *J. Cell Biol.* **143**, 319 (1998).

69. We thank G. Waters for p115 cDNA and p115 mAbs; G. Warren for p97 and p47 antibodies; R. Scheller for rbt1, membrin, and sec22 cDNAs; H. Plutner for excellent technical assistance; and P. Tan for help during the initial phase of this work. Supported by NIH grants GM 33301 and GM42336 and National Cancer Institute grant CA58689 (W.E.B.), a NIH National Research Service Award (B.D.M.), and a Wellcome Trust International Traveling Fellowship (B.B.A.).

20 March 2000; accepted 22 May 2000

One Sequence, Two Ribozymes: Implications for the Emergence of New Ribozyme Folds

Erik A. Schultes and David P. Bartel*

We describe a single RNA sequence that can assume either of two ribozyme folds and catalyze the two respective reactions. The two ribozyme folds share no evolutionary history and are completely different, with no base pairs (and probably no hydrogen bonds) in common. Minor variants of this sequence are highly active for one or the other reaction, and can be accessed from prototype ribozymes through a series of neutral mutations. Thus, in the course of evolution, new RNA folds could arise from preexisting folds, without the need to carry inactive intermediate sequences. This raises the possibility that biological RNAs having no structural or functional similarity might share a common ancestry. Furthermore, functional and structural divergence might, in some cases, precede rather than follow gene duplication.

Related protein or RNA sequences with the same folded conformation can often perform very different biochemical functions, indicating that new biochemical functions can arise from preexisting folds. But what evolutionary mechanisms give rise to sequences with new macromolecular folds? When considering the origin of new folds, it is useful to picture, among all sequence possibilities, the distribution of sequences with a particular fold and function. This distribution can range very far in sequence space (1). For example, only seven nucleotides are strictly conserved among the group I self-splicing introns, yet secondary (and presumably tertiary) structure within the core of the ribozyme is preserved (2). Because these dis-

crete isolates have the same fold and function, it is thought that they descended from a common ancestor through a series of mutational variants that were each functional. Hence, sequence heterogeneity among divergent isolates implies the existence of paths through sequence space that have allowed neutral drift from the ancestral sequence to each isolate. The set of all possible neutral paths composes a "neutral network," connecting in sequence space those widely dispersed sequences sharing a particular fold and activity, such that any sequence on the network can potentially access very distant sequences by neutral mutations (3-5).

Theoretical analyses using algorithms for predicting RNA secondary structure have suggested that different neutral networks are interwoven and can approach each other very closely (3, 5-8). Of particular interest is whether ribozyme neutral networks approach each other so closely that they intersect. If so, a single sequence would be capable of folding into two different conformations, would

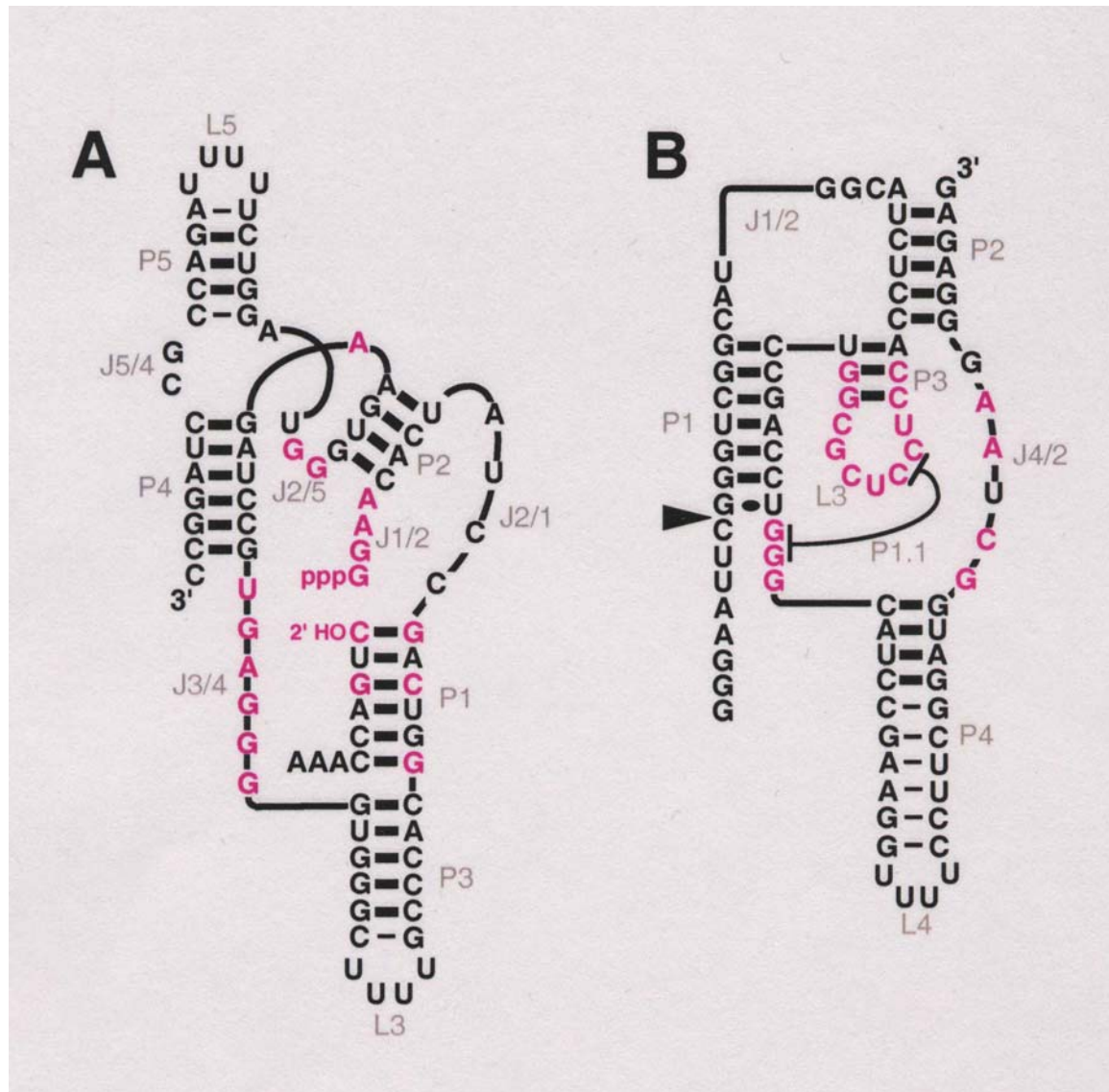
have two different catalytic activities, and could access by neutral drift every sequence on both networks. With intersecting networks, RNAs with novel structures and activities could arise from previously existing ribozymes, without the need to carry non-functional sequences as evolutionary intermediates. Here, we explore the proximity of neutral networks experimentally, at the level of RNA function. We describe a close apposition of the neutral networks for the hepatitis delta virus (HDV) self-cleaving ribozyme and the class III self-ligating ribozyme.

In choosing the two ribozymes for this investigation, an important criterion was that they share no evolutionary history that might confound the evolutionary interpretations of our results. Choosing at least one artificial ribozyme ensured independent evolutionary histories. The class III ligase is a synthetic ribozyme isolated previously from a pool of random RNA sequences (9). It joins an oligonucleotide substrate to its 5' terminus. The prototype ligase sequence (Fig. 1A) is a shortened version of the most active class III variant isolated after 10 cycles of in vitro selection and evolution. This minimal construct retains the activity of the full-length isolate (10). The HDV ribozyme carries out the site-specific self-cleavage reactions needed during the life cycle of HDV, a satellite virus of hepatitis B with a circular, single-stranded RNA genome (11). The prototype HDV construct for our study (Fig. 1B) is a shortened version of the antigenomic HDV ribozyme (12), which undergoes self-cleavage at a rate similar to that reported for other antigenomic constructs (13, 14).

The prototype class III and HDV ribozymes have no more than the 25% sequence identity expected by chance and no fortuitous structural similarities that might favor an intersection of their two neutral networks. Nevertheless, sequences can be designed that simultaneously satisfy the base-pairing requirements

Whitehead Institute for Biomedical Research and Department of Biology, Massachusetts Institute of Technology, 9 Cambridge Center, Cambridge, MA 02142, USA.

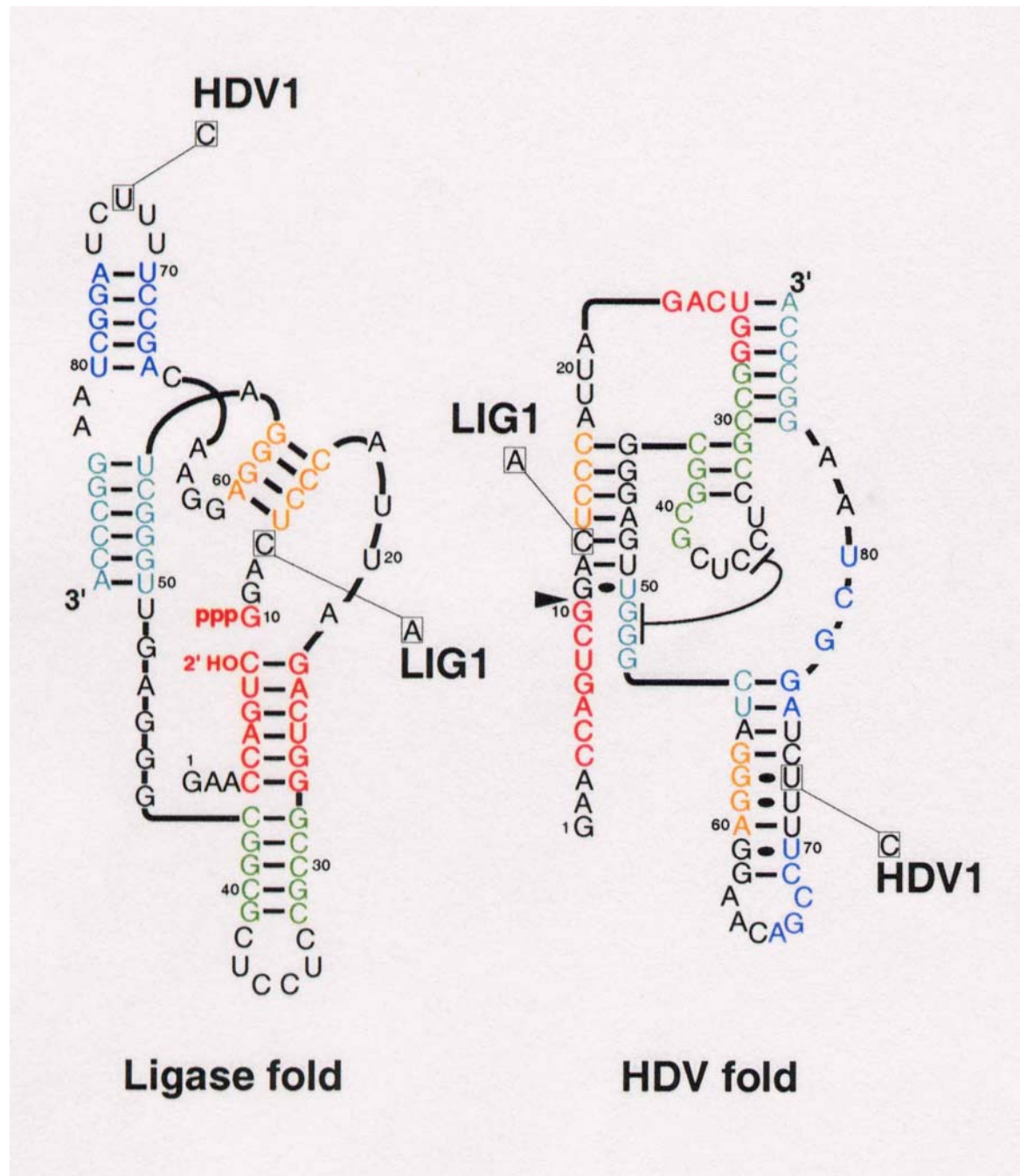
*To whom correspondence should be addressed. E-mail: dbartel@wi.mit.edu

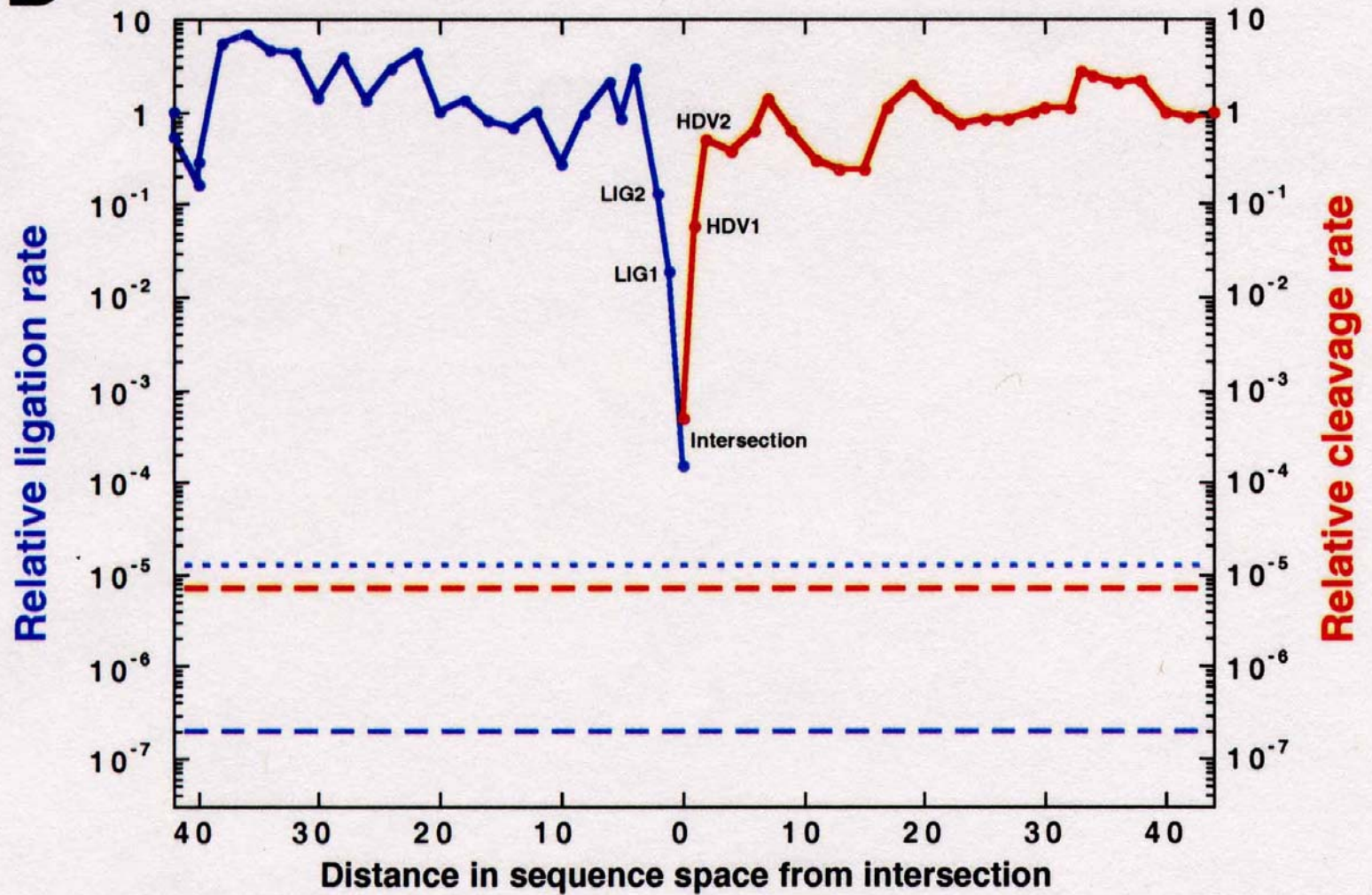


Two ribozymes of chain lengths $n = 88$ nucleotides: An artificial ligase (**A**) and a natural cleavage ribozyme of hepatitis- δ -virus (**B**)

The sequence at the *intersection*:

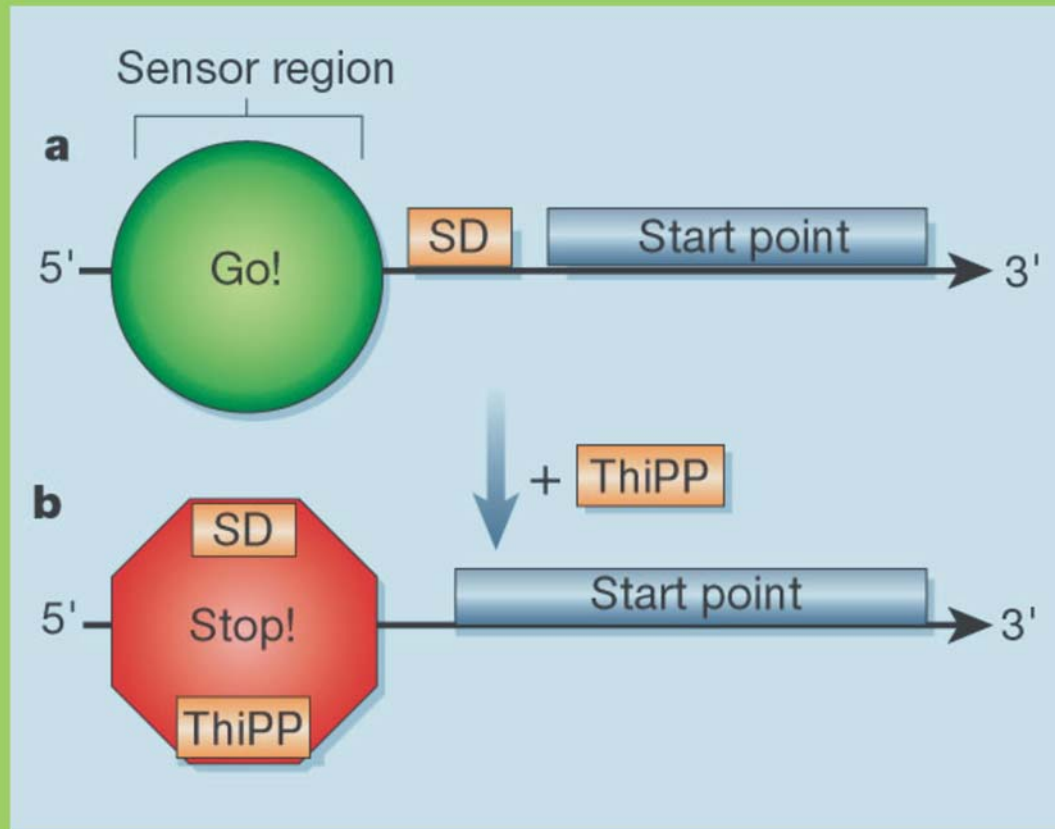
An RNA molecules which is 88 nucleotides long and can form both structures



B

Two neutral walks through sequence space with conservation of structure and catalytic activity

Allosteric control of transcribed RNA



Riboswitches controlled by metabolites

Web-Page for further information:

<http://www.tbi.univie.ac.at/~pks>

