

Circularization and multimerization of synthetic ribozymes

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Outline

- **The topic**

Design of self-polymerizing RNA

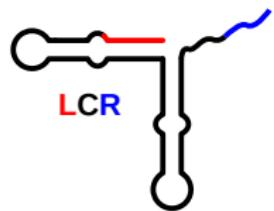
- **The strategy**

Biochemistry, Physics and Computational Biology

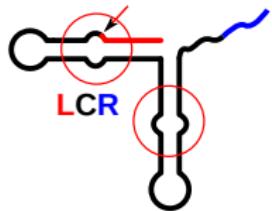
- **The loop**

{Design, Implementation, Evaluation}

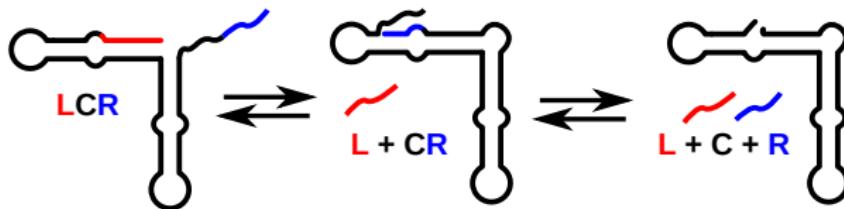
The topic: Design of self-polymerizing RNA



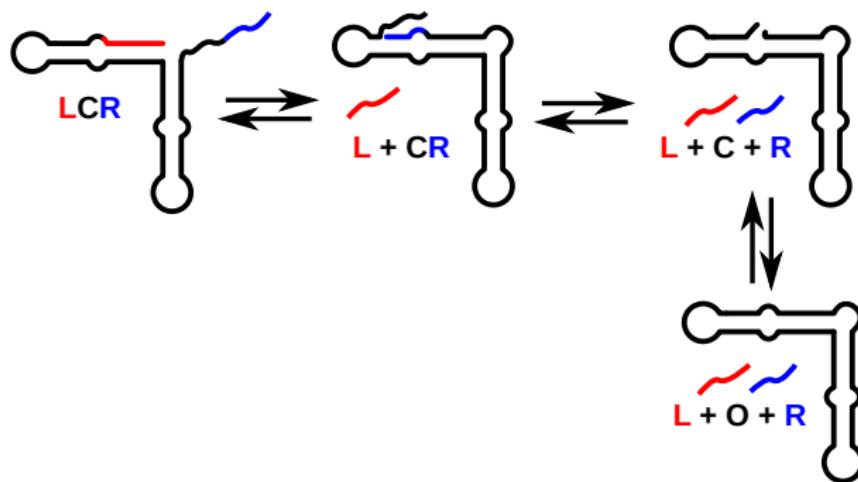
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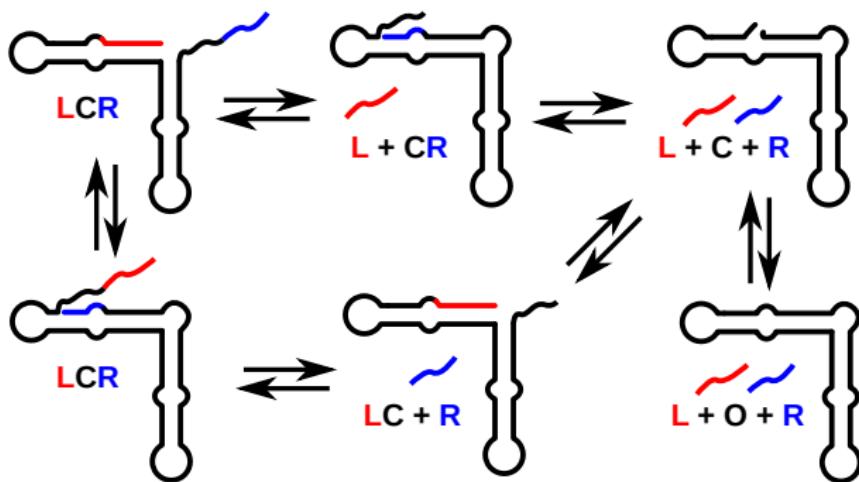
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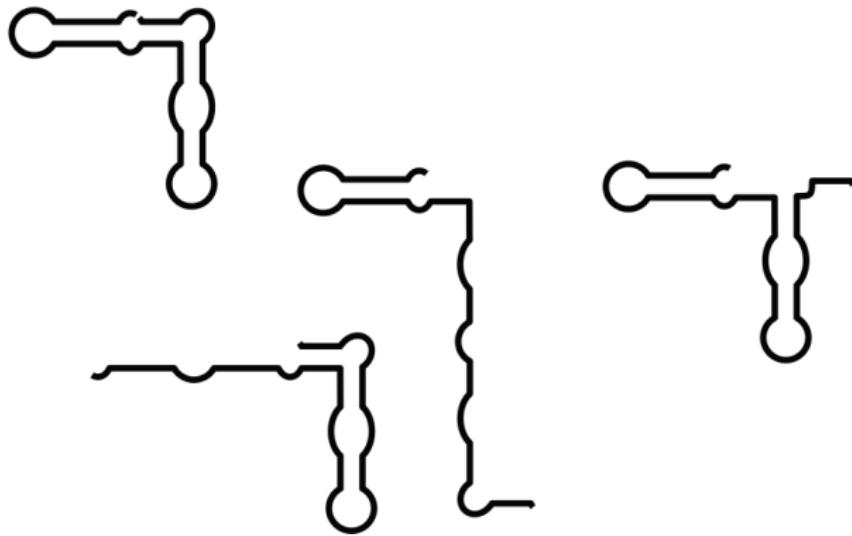
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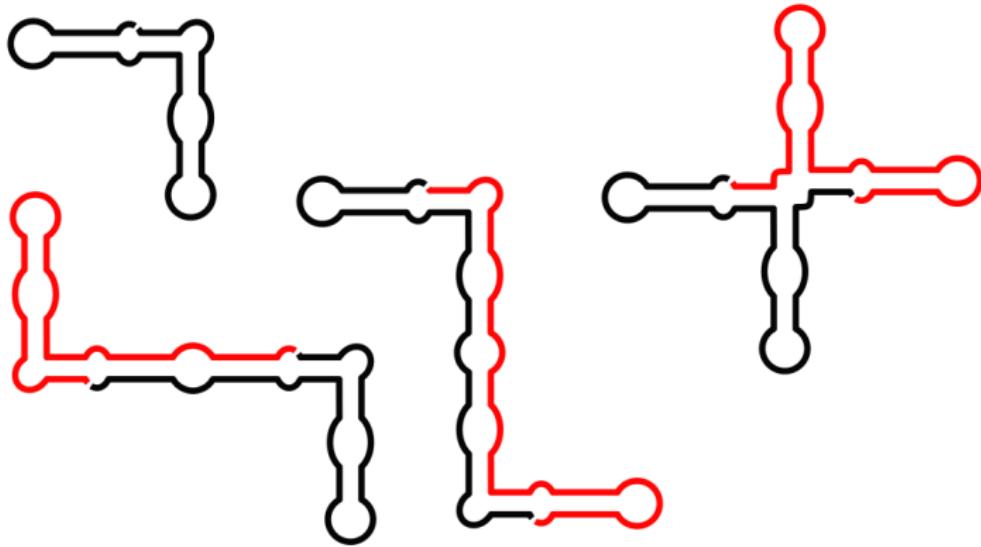
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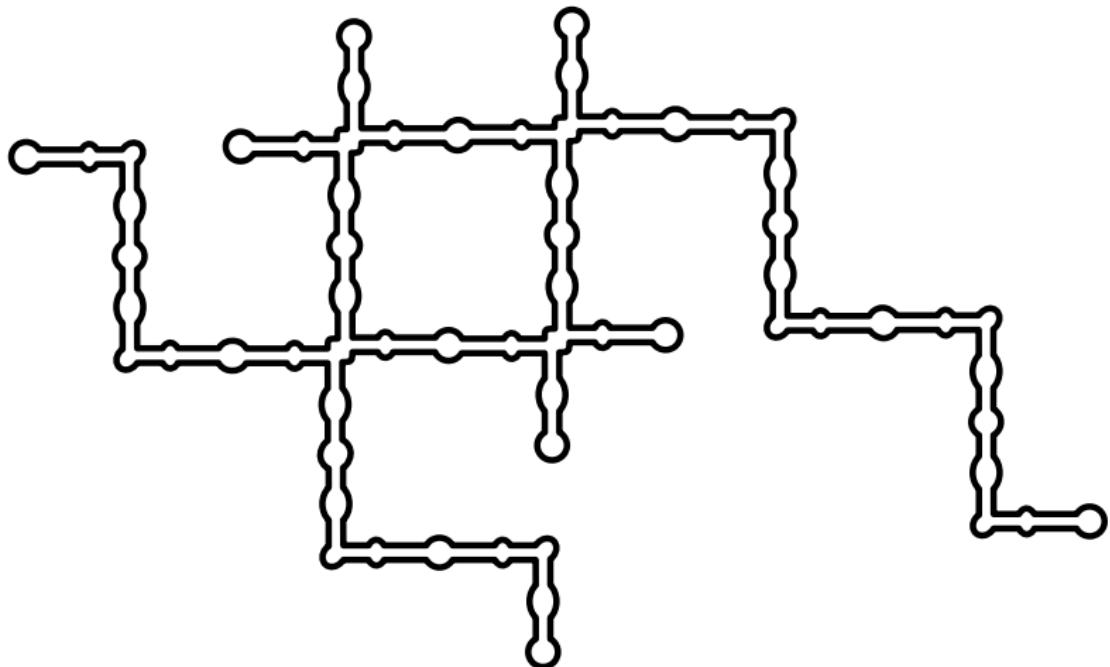
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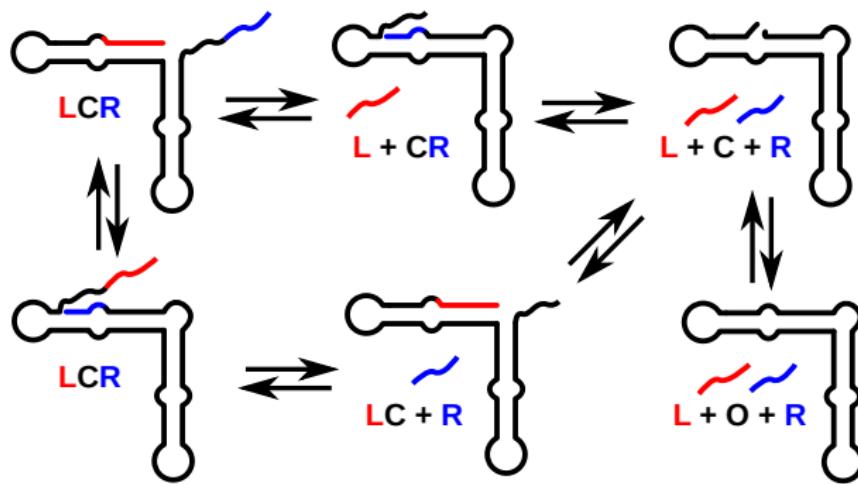
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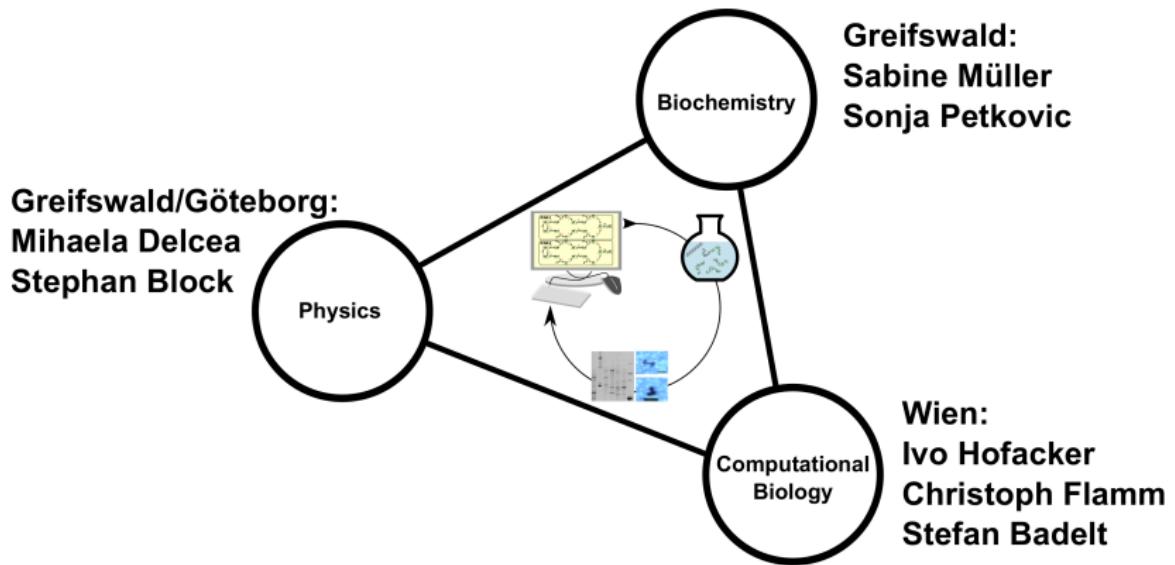


The topic: Design of self-polymerizing RNA

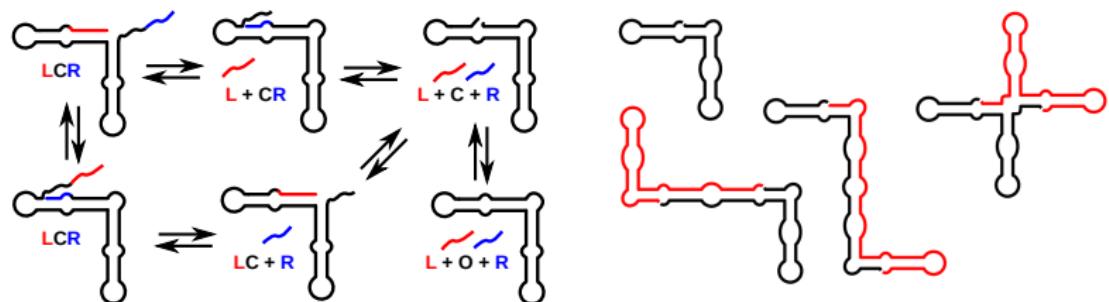


- Does this system exist in nature?
- What is the biological relevance?

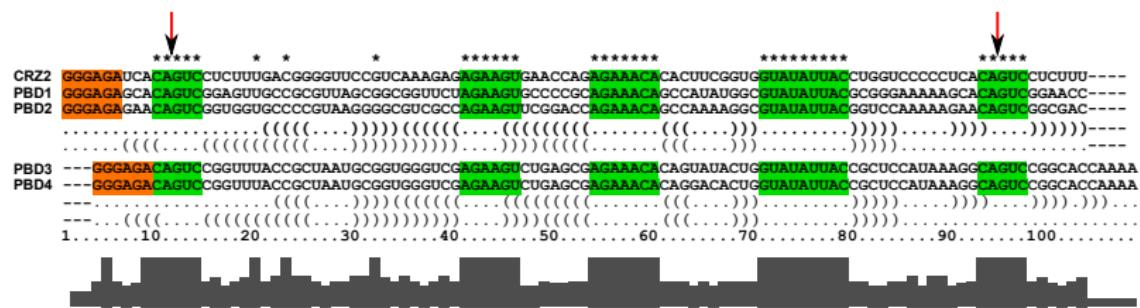
The strategy: Biochem., Physics and Comp.Bio.



The loop: Design, Implementation, Evaluation



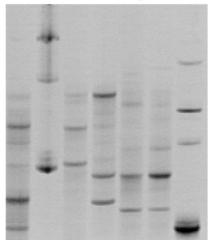
- ⇒ compute a set of candidate molecules (switch.pl)
- ⇒ maximize probabilities to form reactive conformations
- ⇒ differ between probabilities to form active monomers and dimers



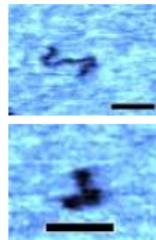
The loop: Design, **Implementation**, Evaluation

- Sample Preparation – Sonja Petkovic
 - ① dsDNA synthesis
 - ② *in vitro* Transcription (T7 RNA polymerase)
 - ③ full-length ribozyme extraction
 - ④ cleavage/ligation/denaturing conditions
- 1D and 2D Gel electrophoresis – Sonja Petkovic
- Atomic Force Microscopy – Stephan Block

Gel Electrophoresis



AFM



The loop: Design, Implementation, **Evaluation**

- Matching Results to Design Objective
- Identify deficiencies of Design Functions
- Include results into the second round of sequence design

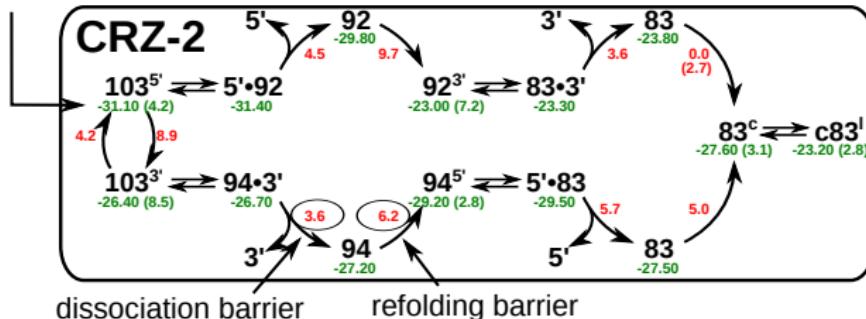
The loop: Design, Implementation, Evaluation

Gel Electrophoresis (Lane Intensity, Nr. of lanes):

RNA	103nt	91-97nt	83nt	circ83nt	> 83nt
CRZ2	-	+++	++	-	6×
PBD1	-	-	-	+++	2×
PBD2	+	+	-	++	4×
PBD3	+	+	-	+	2×
PBD4	++	+	-	+	2×

Computational Analysis:

free energy (activation energy)

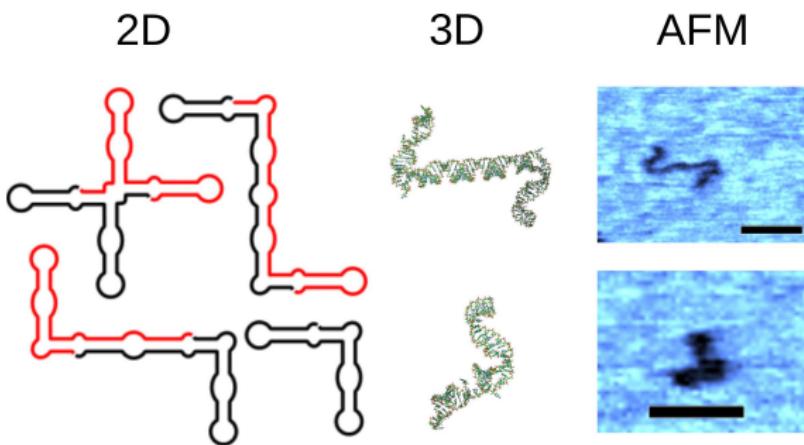


The loop: Design, Implementation, Evaluation

Preliminary Results – Atomic Force Microscopy:

RNA	Ratio (Monomers : Dimers : Trimmers)
CRZ2	(7 :1:0)
PBD1	(10:3:1)
PBD4	(5 :1:1)

Computational Analysis:



Conclusion

Results:

- Design approach is sufficient to yield circular monomers
- Dissociation barriers do have an impact on efficiency
- Diversity of dimers has to be considered for optimization
- Optimizing for dimer-ligation does not lead to multimerization

Remaining Questions:

- Do molecules tend to cleave first and then multimerize (need of time resolved results)
- Will we be able to fit tertiary structure predictions to AFM images?

thanks to

	Supervisor	PhD committee	Experimenter
Ivo Hofacker	x	x	
Christoph Flamm	x		
Sabine Müller		x	x
Sonja Petkovic			x
Stephan Block			x
Peter Stadler		x	

and the whole TBI group for everything else



universität
wien



FWF

Der Wissenschaftsfonds.

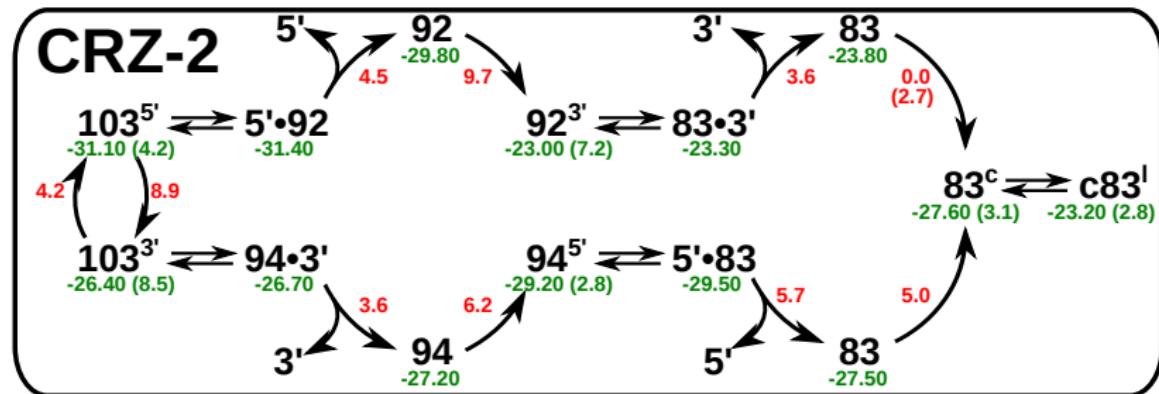
The research was funded by the Austrian Science Fund (FWF): W1207-B09, I670-B11

Fitness Functions

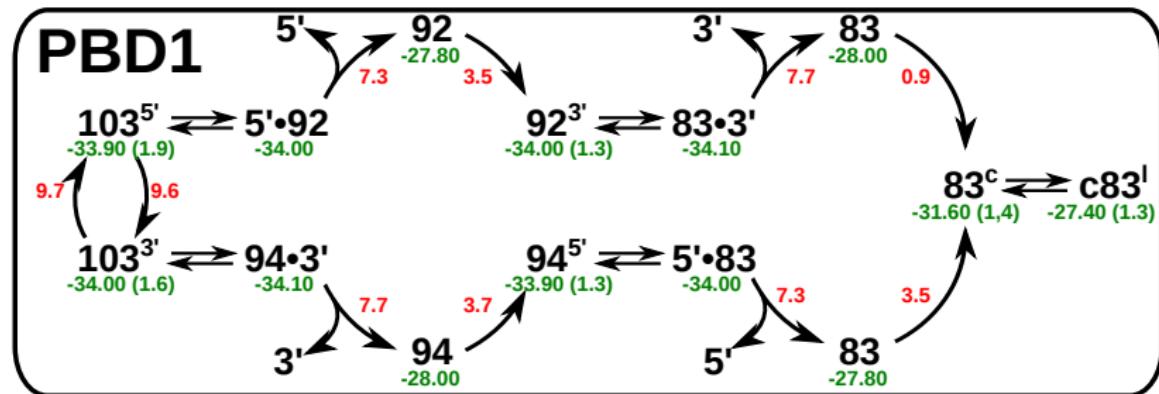
$$\kappa_1 = \left(\min \begin{cases} P(LCR^L), \\ P(CR^R) \end{cases} + \min \begin{cases} P(LCR^R), \\ P(LC^L) \end{cases} \right) \times e^{\frac{E(C^O) - E(O^C)}{kT}} \quad (1)$$

$$\kappa_2 = \frac{[LCR_{\text{dimer}}]_\theta}{[LCR_{\text{monomer}}]_\theta} \times \left(\min \begin{cases} P(LCR_{\text{dimer}}^L), \\ P(CR_{\text{dimer}}^R) \end{cases} + \min \begin{cases} P(LCR_{\text{dimer}}^R), \\ P(LC_{\text{dimer}}^L) \end{cases} \right) \times e^{\frac{E(C_{\text{dimer}}^O) - E(O_{\text{dimer}}^C)}{kT}} \quad (2)$$

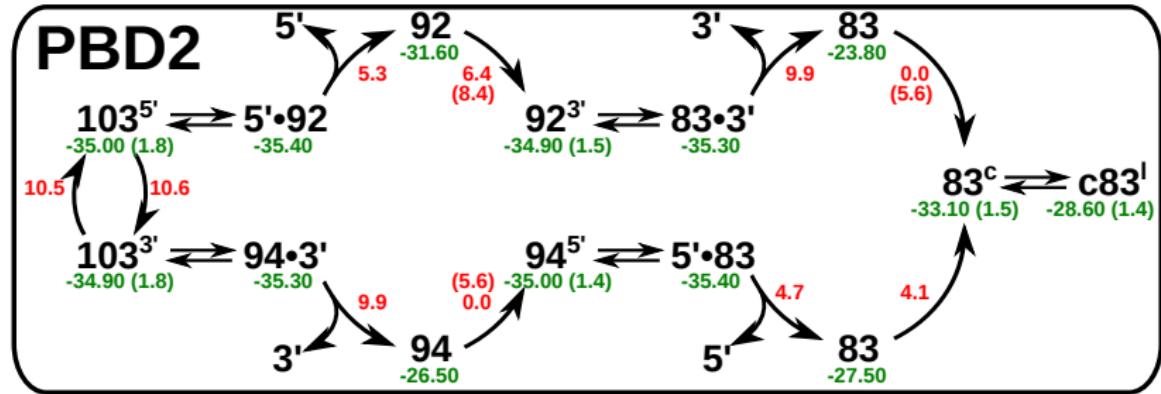
Cascades



Cascades



Cascades



Cascades

